

Effect of Salinities Stress on Seed Germination of *Glycine. Soja*

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Abstract: Wild soybeans (*Glycine soja*), the ancestors of cultivated soybean (*Glycine max*), are important sources of major genes for resistance to pests, diseases and environmental stresses. The study of salt stress on the seed germination is invaluable for efficient utilization of saline-alkali soil resources and wild soybeans. Wild soybean, collected from Dezhou, Shandong province, were treated with NaCl, Na₂SO₄, Na₂CO₃ and the mixture of three solutions. Seed germination percentage, viability, index and the growth of radicle and hypocotyl of annual wild soybean were measured and analyzed. Results showed seed germination percentage, germination viability, germination index and the growth of radicle and hypocotyl annual wild soybean reduced with increasing of salt concentrations of Na₂CO₃ and the mixture. Low concentrations of Na₂SO₄ (10-50 mmol · L⁻¹), Na₂CO₃ (0-10 mmol · L⁻¹) promoted germination of seeds, while high concentrations of NaCl (>200 mmol · L⁻¹), Na₂SO₄ (≥200 mmol · L⁻¹) and Na₂CO₃ (≥75 mmol · L⁻¹) restrained it. Under low concentration of salt solution (10 mmol · L⁻¹ Na₂SO₄), the radicle length of *Glycine soja* was longer than that of the control (distilled water). Results suggest that lower concentration of salt solution simulated the growth of radicle and hypocotyl of wild soybean, and the radicle is more sensitive to salt stress than that of hypocotyl.

Key words: Annual wild soybean; Salt stress; Germination rate; Germination viability

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不同盐分胁迫对野生大豆种子发芽的影响

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摘要: 一年生野生大豆是栽培大豆的野生近缘种和基因资源, 具有耐盐碱、抗寒、抗病等优良性状。研究其种子萌发的耐盐性, 为利用盐碱地资源和野大豆的引种驯化供依据。以山东德州一年生野大豆为材料, 研究 NaCl、Na₂SO₄、Na₂CO₃ 及三者的混合盐的胁迫对野大豆种子的发芽率、发芽势、发芽指数及胚生长的影响。随盐溶液浓度的增加, 野大豆种子的发芽率、发芽速度、发芽指数均呈下降趋势, 而低浓度的 Na₂SO₄ (10 ~ 50 mmol · L⁻¹), Na₂CO₃ (0 ~ 10 mmol · L⁻¹) 促进种子萌发, 高浓度的 NaCl (>200 mmol · L⁻¹)、Na₂SO₄ (≥200 mmol · L⁻¹)、Na₂CO₃ (≥75 mmol · L⁻¹) 抑制种子萌发; 10 mmol · L⁻¹ Na₂SO₄ 处理下, 其胚根胚轴长度都大于未经盐处理的, 低浓度的盐分促进了胚根和胚轴的生长。结果表明低浓度的盐分促进一年生野生大豆胚的生长, 野生大豆的胚根比胚轴对盐分更敏感。

关键词: 野生大豆; 盐胁迫; 发芽率; 发芽势

Germination is a key part of plant life histories in saline environments as it determines whether or not the plants can establish successfully in certain areas^[1]. In general, non-halophyte and halophyte respond to salinity in a similar way during the germination stage; the in-

itial germination process is delayed under salt stress^[2]. Seed of halophyte under natural condition are subjected to saline stress and usually it is dominated by NaCl. However, other sulfate and carbonate salt and their interaction plays a significant role in affecting seed ger-

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mination^[3]. Salt can affected seed germination either by restricting the supply of water (osmotic affect) or causing specific injury through ions to the metabolic machinery(ionic effect). The mechanism for how salinity inhibits seed germination of halophytes in a highly saline environment has received a lot of attention in recent years. For example Bajji et al.^[4] reported that the inhibitory effect of NaCl stress on seed germination is mainly due to an osmotic effect, rather than specific ion toxicity, in halophytes of *A. halimus*. However, others reported both osmotic and ionic effects^[5].

In general, Authors have concluded that salinity is inhibitory to the germination in following ways: either causing a complete inhibition of the germination process at salinities beyond the tolerance limits of a species, or delaying the germination of seeds, at salinities that cause some stress to seeds but do not prevent germination, causing the loss of seeds viability by high salinity^[6-7].

Wild soybean (*Glycine soja* Sieb. et Zucc) is an annual herb, it is a slender twiner and grows in fields, along roadsides and riverbanks^[8]. The plant has trifoliate leaves. Seeds are oval-oblong. Flower color is pink, lilac or purple. Wild soybean has been thought to be the wild progenitor of the domesticated soybean (*G. max* [L.] Merr.) which was thought to have originated in China^[9] and is grown in China and adjacent countries or regions. Some wild soybeans have been used as a genetic resource for improvement on the cultivars.

Annual wild soybeans (*Glycine soja*) are distributed throughout East Asia, including Korea, Japan and the far eastern regions of Russia, and are centered in China^[10]. The geographical distribution of wild soybeans in China is very wide. The horizontal distribution is from Tahe County, Heilongjiang (latitude: 53) in the north to Xiangzhou County, Guangxi (latitude: 24) in the south and from Raohe County, Heilongjiang (longitude: 134), in the east to Chayu County, Tibet (longitude: 97), in the west. The vertical distribution is from 1.8 to 2650 m above sea level^[11].

Annual wild soybean, the ancestors of cultivated soybeans (*G. max*), is important sources of major genes for resistance to pests, diseases and environmental

stresses. It can be used for improving the soil organic matter. In addition, it is a good source of food and forage, its grain, stem and leaf can be used as feedstuffs. In the current, salt tolerance of cultivated soybeans (*G. max*) is limited and the genetic basis of more and more narrow circumstances, wild soybeans is therefore considered to be an important gene source of soybean^[12]. Because soil mainly contain NaCl, Na₂SO₄ and Na₂CO₃, NaCl is the most abundant salt in many salinized lacotions in the North China. We used them in our experiment. The present report describes the effect of various concentrations of sodium chloride, sodium sulfate, sodium carbonate and the mixture of three solutions on seed germination, seedling growth of annual wild soybean.

1 Materials and Methods

Annual wild soybeans were collected from a natural grassland located in the east of Dezhou, Shandong province (China) (38° 0' N, 117° 34' E) in October 2006. Dry seeds were stored in a refrigerator at <4°C for 6 months before being used. Because the seed coat was hard, the seed coat was break by mechanical scarification. The trial was conducted at Biology Department, Dezhou University, Dezhou, Shandong, China, in April 2007. After being surface sterilized with 0.1% KMnO₄ solution for 20 min and rinsed in tap water. 40 seeds of annual wild soybean were placed on two layers of filter paper in a 1200 mm Petri dishes. The filter paper was moistened with about 10 mL distilled water (control) or solution of salt. Each experiment was replicated three times. Seeds were considered to have germinated when the emerging radicle was at least 5 mm.

Germination tests were conducted in incubators with a regime of 13 h light at 25°C and 11 h darkness at 20°C. The filter paper was moistened with distilled water for the controls, or with aqueous solution of NaCl, Na₂SO₄, Na₂CO₄ and the mixture of three solutions. To determine the effect of different salt on germination, solution of four salts were used, containing increasing levels of NaCl, Na₂SO₄, Na₂CO₄ and the mixture of three solutions (Table 1). In the course of the

experiment, the Petri dishes were covered with lids and kept in a controlled environment. About two-thirds of volume of the water or solution in each Petri dish was replaced daily to remain the filter paper under the same concentration. The number of germinated seeds was

Table 1 The composition, concentration ($\text{mmol} \cdot \text{L}^{-1}$) and pH value of a set of salt treatment

No. of Treatment	NaCl	pH	Na ₂ SO ₄	pH	Na ₂ CO ₃	pH	Mixed salt				pH
							Na ⁺	Cl ⁻	SO ₄ ²⁻	CO ₃ ²⁻	
CK	0	7	0	7	0	7	0	0	0	0	7
1	50	7.06	10	7.32	10	9.38	90	50	10	10	9.26
2	200	6.68	50	7.46	50	10.21	400	200	50	50	10.01
3	400	6.26	150	7.60	75	10.41	850	400	150	75	10.12
4	600	6.02	200	7.80	150	10.63	1300	600	200	150	10.26
5	800	5.80	400	8.50	200	10.73	2000	800	400	200	10.34

The germination percentage, germination viability, germination index and relative germination percentages, were determined by the following formulae, respectively:

$$\text{Germination percentage}(\%) = [a/b]100,$$

$$\text{Germination viability}(\%) = [d/b]100,$$

$$\text{Germination index} = \sum G_t/D_t,$$

$$\text{Relative germination percentages} = f/e,$$

Where a is the total number of seeds germinated on each day, b is the total number of seeds, d is the total number of seeds germinated on second day, G_t is the number of seeds germinated on t days, D_t is the number of germination days corresponding, f is germination percentage in salts solution, e is germination percentage of control.

Statistical analyses were carried out using SPSS 13.0. A one-way ANOVA was carried out to determine the difference among treatment group means for germination percent, germination viability, germination index and relative percentages, and significance is indicated in the form of superscripted letters in the tables.

2 Results and discussion

2.1 Effects of salt stress on seed germination

The germination pattern varied under different salts in this experiment. Seed germination was affected by different concentrations of NaCl, Na₂SO₄, Na₂CO₃

counted daily, and the final percentage germination was recorded 6 d after incubation when the number of germination is constant. The lengths of radicle and hypocotyl were measured 15 times in each repeat when seed germination ended on the sixth day.

and the mixture of three solutions. An obvious decrease was observed in annual wild soybean when the concentration of NaCl, Na₂SO₄, Na₂CO₃ and the mixture of three solutions was increased. But the germination percentage was not significantly different among 0, 50 $\text{mmol} \cdot \text{L}^{-1}$ NaCl, 10, 50 $\text{mmol} \cdot \text{L}^{-1}$ Na₂SO₄ and 10 $\text{mmol} \cdot \text{L}^{-1}$ Na₂CO₃. In distilled water, germination percentage of annual wild soybean was 96.67%. Low concentrations of Na₂SO₄ (10 – 50 $\text{mmol} \cdot \text{L}^{-1}$) and Na₂CO₃ (0 – 10 $\text{mmol} \cdot \text{L}^{-1}$) promoted the germination percentage of seeds, while high concentrations of NaCl (> 200 $\text{mmol} \cdot \text{L}^{-1}$), Na₂SO₄ (\geq 200 $\text{mmol} \cdot \text{L}^{-1}$), Na₂CO₃ (> 75 $\text{mmol} \cdot \text{L}^{-1}$) and mixture salt 2 restrained it. There were 90.00% seeds germinated at 200 $\text{mmol} \cdot \text{L}^{-1}$ NaCl treatment, 59.17% seeds germinated at 200 $\text{mmol} \cdot \text{L}^{-1}$ Na₂SO₄ treatment, 84.17% seeds germinated at 75 $\text{mmol} \cdot \text{L}^{-1}$ Na₂CO₃ treatment and 94.17% seeds germinated at mixture salt 1 treatment. At the higher level of salinity (\geq 400 $\text{mmol} \cdot \text{L}^{-1}$ NaCl and Na₂SO₄ concentration, \geq 150 $\text{mmol} \cdot \text{L}^{-1}$ Na₂CO₃ and mixture 2) no seed germinated. The high salt stress inhibits the seed germination completely (Table 2). The germination viability, index of annual wild soybeans decreased significantly with increasing of salt concentration of NaCl, Na₂SO₄, Na₂CO₃ and the mixture of three solutions, and have significant difference with the control.

Table 2 The effect of different salt stress on germination percentage, viability, index and relative germination rate

Salt sort	Concentration/ mmol · L ⁻¹	Germination percentage/%	Germination viability	Germination index	Relative germination percentage
NaCl	0	96.67 ± 2.89a	65.00 ± 6.61a	32.11 ± 1.51a	100.00 ± 2.99a
	50	95.83 ± 1.44a	13.33 ± 11.81b	21.69 ± 2.56a	99.14 ± 1.49a
	200	90.00 ± 5.00b	0c	16.06 ± 1.55b	93.10 ± 5.17b
	400	0c	0c	0c	0c
	600	0c	0c	0c	0c
	800	0c	0c	0c	0c
Na ₂ SO ₄	0	96.67 ± 2.89a	65.00 ± 6.61a	32.11 ± 1.51a	100.00 ± 2.99a
	10	95.83 ± 5.20a	34.17 ± 10.10b	25.84 ± 2.33a	99.14 ± 5.38a
	50	98.33 ± 1.44a	0c	19.42 ± 0.36a	101.72 ± 1.40a
	150	80.00 ± 5.00b	0c	9.36 ± 1.16b	82.76 ± 5.17b
	200	59.17 ± 14.22c	0c	5.64 ± 1.36b	61.21 ± 14.71c
	400	0d	0c	0b	0d
Na ₂ CO ₃	0	96.67 ± 2.89a	65.00 ± 6.61a	32.11 ± 1.51a	100.00 ± 2.99a
	10	99.17 ± 1.44a	51.67 ± 8.04b	30.08 ± 1.38a	102.59 ± 1.49a
	50	96.67 ± 3.82a	7.50 ± 2.50c	20.14 ± 0.13a	100.00 ± 3.95a
	75	84.17 ± 1.44b	2.50 ± 2.50c	13.28 ± 2.77b	87.07 ± 1.49b
	150	0c	0c	0c	0c
	200	0c	0c	0c	0c
Mixture salt	0	96.67 ± 2.89a	65.00 ± 6.61a	32.11 ± 1.51a	100.00 ± 2.99a
	Mixture 1	94.17 ± 1.44b	0b	18.22 ± 0.57b	97.41 ± 1.49b
	Mixture 2	0c	0b	0c	0c
	Mixture 3	0c	0b	0c	0c
	Mixture 4	0c	0b	0c	0c
	Mixture 5	0c	0b	0c	0c

Optimal germination for most halophyte has been reported in non-saline conditions^[13-14]. Seeds of annual wild soybeans germinated better in non-saline controls and any increase in salinity progressively inhibited seed germination and no seed germinated beyond the 400 mmol · L⁻¹ NaCl, Na₂SO₄, 150 mmol · L⁻¹ Na₂CO₃ and the mixture 2 salt treatment. An increase in salinity can induce reduction in the percentage of seeds germinated, a delay in the initiation of the germination process, and dormancy of seeds^[6,15]. Salt induced inhibition of seed germination could be attributed to osmotic stress or to specific ion toxicity^[16]. Salinity stress affects seed germination either by decreasing the rate of water uptake (osmotic effect) and or by facilitating the intake of ions, which may change certain enzymatic or hormonal activities of the seed (ion toxicity)^[16]. Our data clearly indicate that seed germination percentage increased with the increase of salt concentrations in suitable range of salt concentration (Table 2).

Not only germination percentage but also the speed of germination was affected by salt. In distilled

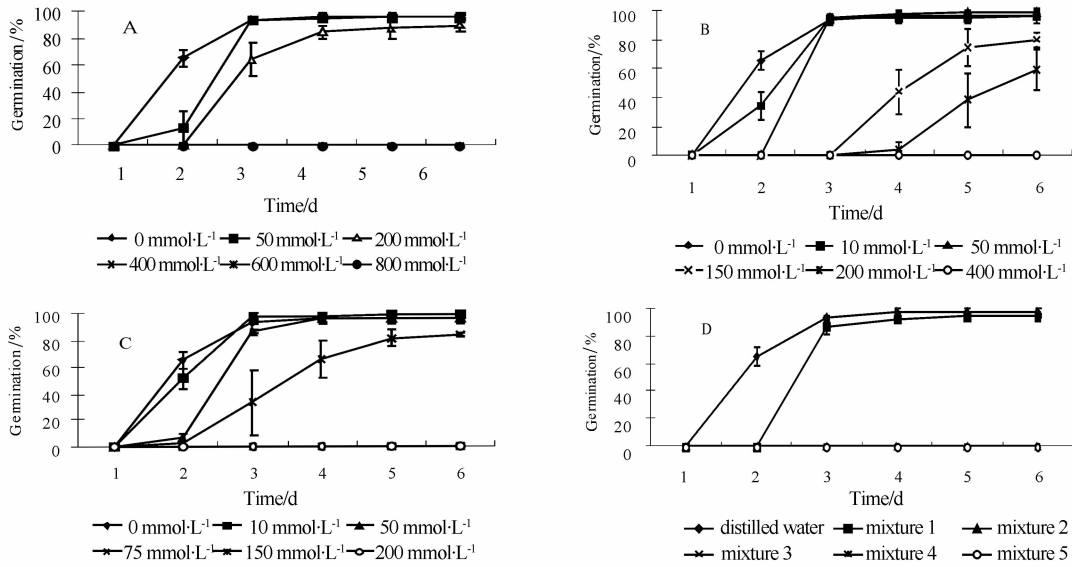
water and the low concentration of salt solution, seed germinated on second day, in 200 mmol · L⁻¹ NaCl, 50 mmol · L⁻¹ Na₂SO₄, 75 mmol · L⁻¹ Na₂CO₃ and the mixture 2 seed germination on third day. Higher concentration of salt delay in the initiation of the germination process. Salinity beyond the tolerance limits can cause complete inhibition of the germination process, even lead to loss of seeds viability (Fig. 1A, Fig. 1B, Fig. 1C, Fig. 1D).

2.2 Effects of salinity on radicle and hypocotyl growth

The radicle growth for annual wild soybeans under salt stress were depressed with increasing of salt concentration. For annual wild soybeans, significant differences were found in radicle and hypocotyle lengths depending on salinity levels. Elongation of radicle and hypocotyle were affected by increasing salt concentration severely (Table 3). Concentration of 200 mmol · L⁻¹ NaCl treatment reduced radicle in annual wild soybeans from 5.07 to 0.81 cm. Concentration of 75 mmol · L⁻¹ Na₂CO₃ treatment reduced radicle in annual wild soybeans from 5.07 to 0.59 cm. The inhibition of radicle

growth under salt stress may be due to the reduction in

the turgor of the radicle cells^[17].



A: NaCl A; B: Na₂SO₄; C: Na₂CO₃; D: Mixture of three solutions

Fig. 1 Effect of different salt on germination rate of annual wild soybean

Under salt stress, the length of radicle and hypocotyl decreased as salt concentrations increased. But the radicle length was longer than those of control at 10 mmol · L⁻¹ Na₂ SO₄, the proper lower concentration stimulated the growth of radicle. The hypocotyle length was longer than those of control at 50 mmol · L⁻¹ NaCl, 10 mmol · L⁻¹ Na₂SO₄ and 10 mmol · L⁻¹ Na₂CO₃, the

proper lower concentration stimulated the growth of hypocotyl. Radicle and hypocotyl length depended on the type and concentration of salts, our result indicate the radicle is more sensitive to salinity than hypocotyl(Table 3). The sensitivity of radicle and hypocotyl of annual wild soybeans to different kind and gradient salt stress were different.

Table 3 Effect of different salt stress on radicle length, hypocotyl length of annual wild soybean

Salt sort	Concentration /mmol · L ⁻¹	Radicle length /cm	Hypocotyl length /cm	Salt sort	Concentration /mmol · L ⁻¹	Radicle length /cm	Hypocotyl length /cm
NaCl	0	5.07 ± 1.42	2.77 ± 0.47	Na ₂ CO ₃	0	5.07 ± 1.42	2.77 ± 0.47
	50	3.98 ± 0.92	3.45 ± 0.40		10	2.73 ± 1.06	3.10 ± 0.71
	200	0.81 ± 0.23	1.00 ± 0.20		50	0.59 ± 0.16	1.11 ± 0.36
	400	0	0		75	0.59 ± 0.18	0.73 ± 0.05
	600	0	0		150	0.00 ± 0.00	0.00 ± 0.00
	800	0	0		200	0.00 ± 0.00	0.00 ± 0.00
Na ₂ SO ₄	0	5.07 ± 1.42	2.77 ± 0.47	Mixture salt	0	5.07 ± 1.42	2.77 ± 0.47
	10	5.23 ± 1.11	4.32 ± 1.56		mixture 1	0.96 ± 0.47	1.44 ± 0.38
	50	2.24 ± 0.47	2.71 ± 0.98		mixture 2	0	0
	150	0.33 ± 0.07	0.72 ± 0.26		mixture 3	0	0
	200	0.43 ± 0.09	0.05 ± 0.01		mixture 4	0	0
	400	0	0		mixture 5	0	0

What constitutes the favorable level of salinity might, however, vary greatly with species due to the differences in their physiological mechanism of resisting salt stress. Euhalophytes have great salt-tolerance than recretohalophytes. Liu et al^[18] reported that *Limonium bicolor* (a recretohalophyte) had a lower range of favora-

ble salinity than *S. salsa* (an euhalophyte). In many other halophytes, most small seedlings survived and radicle continued to elongate at salinities that allowed seeds to germinate. Appropriate salinity in soil is necessary for the growth of most halophytes^[19].

3 Conclusion

All salts significantly inhibited seed germination of annual wild soybean. The magnitude of inhibition of different salts was in the order; mixture salt > NaCl > Na₂CO₃ > Na₂SO₄.

Low salinity (10 – 50 mmol · L⁻¹ Na₂SO₄ and 0 – 10 mmol · L⁻¹ Na₂CO₃) promoted seed germination and high salinity inhibited seed germination. The high salt stress inhibits the seed germination completely. The germination viability and germination index of annual wild soybeans decreased with increasing concentration of NaCl, Na₂SO₄, Na₂CO₃ and the mixture of three solutions.

The radicle and hypocotyl had different response to the salts; the radicle is more sensitive than that of hypocotyl.

References

- [1] Song J, Feng G, Tian C Y, et al. Strategies for adaptation of *Suaeda physophora*, *Haloxylon ammodendron* and *Haloxylon persicum* to a saline environment during seed-germination stage [J]. *Annals of Botany*, 2005, 96(3):399-405.
- [2] Khan M A, Ungar I A. Effects of thermoperiod on recovery of seed germination of halophytes from saline conditions [J]. *American Journal of Botany*, 1997, 84(2):279-283.
- [3] Khan M A. Halophyte seed germination; Success and Pitfalls [C]// Hegazi A M. International symposium on optimum resource utilization in salt affected ecosystems in arid and semi arid regions. Cairo, Egypt, 2002:346-358.
- [4] Bajji M, Kinet J M, Lutts S. Osmotic and ionic effects of NaCl on germination, early seedling growth and ion content of *Atriplex halimus* (Chenopodiaceae) [J]. *Canada Journal of Botany*, 2002, 80(3):297-304.
- [5] Mohammad S, Sen D N. Germination behavior of some halophytes in Indian desert [J]. *Indian Journal of Experimental Biology*, 1990, 28(5):545-549.
- [6] Pujol J A, Calvo J F, Diaz L R. Recovery of germination from different osmotic conditions by four halophytes from southeastern Spain [J]. *Annals of Botany* 2000, 85(2):279-286.
- [7] Li L, Zhang X M, Rung M, et al. Responses of germination and radicle growth of two *Populus* species to water potential and salinity [J]. *Forestry Studies in China*, 2006, 8(1):10-15. (in Chinese with English abstract)
- [8] Hymowitz T, Newell C A. Taxonomy, speciation, domestication, dissemination, germplasm resources and variation in the genus *Glycine* [M]// Summerfield R J, Bunting A H. *Advances in Legume Science*. Kew: Royal Botanic Gardens, 1980:251-264.
- [9] Hymowitz T, Newell C A. Taxonomy of the genus *Glycine*, domestication and uses of soybeans [J]. *Economic Botany*, 1981, 35:272-288.
- [10] Lee J D, Yu J K, Hwang Y H, et al. Genetic diversity of wild soybean (*Glycine soja* Sieb. and Zucc.) accessions from South Korea and other countries [J]. *Crop Science*, 2008, 48(2):606-616.
- [11] Dong Y S, Zhuang B C, Zhao L M, et al. The genetic diversity of annual wild soybeans grown in China [J]. *Theoretical and Applied Genetics*, 2001, 103(1):98-103.
- [12] Chen X Q, Liu H P, Luo Q Y, et al. Response of seed and seedling of *Glycine soja* with different salt tolerance to isoosmotic water and NaCl stresses [J]. *Journal of Nanjing Agricultural University*, 2006, 29(4):28-32. (in Chinese with English abstract)
- [13] Shaikh F, Gul B, Li W Q, et al. Effect of calcium and light on the germination of *urochondra setulosa* under different salts [J]. *Journal of Zhejiang University Science B*, 2007, 8(1):20-26.
- [14] Gulzar S, Khan M A. Seed germination of a halophytic grass *aelurus lagopoides* [J]. *Annals of Botany*, 2001, 87(3):319-324.
- [15] Zhao K F, Fan H. Halophytes and their physiological adaptation to saline habitats [M]. Beijing: Science Press, 2005. (in Chinese)
- [16] Huang J, Redmann R E. Salt tolerance of hordeum and brassica species during germination Zidan and early seedling growth [J]. *Canadian Journal of Plant Science*, 1995, 75(6):815-819.
- [17] Bewley J D, Black M. *Seeds: Physiology of development and germination* [M]. 2nd Edn. Plenum Press, New York and London, 1994:147.
- [18] Liu X J, Duan D Y, Li W Q, et al. A comparative study on responses of growth and solute composition in halophytes *suaeda salsa* and *limonium bicolor* to salinity [M]//Khan M A, Weber D J. *Ecophysiology of High Salinity Tolerant Plants*. Springer, Netherlands, 2006:135-143.
- [19] Subbarao G V, Itoa O, Berryb W L, et al. Sodium- a functional plant nutrient [J]. *Critical Reviews in Plant Sciences*, 2003, 22:391-416.