

## Influence of dietary selenium level on the concentration of conjugated linoleic acid isomers, other fatty acids and amino acids in the liver and femoral muscles of rats

M. CZAUDERNA, J. KOWALCZYK, K.A. KRAJEWSKA

Kielanowski Institute of Animal Physiology and Nutrition, Polish Academy of Sciences, Jabłonna, Poland

**ABSTRACT:** The purpose of the present study was to determine the influence of diets containing conjugated linoleic acid isomers (CLAmix) with or without low ( $_{L}$ Se) or high ( $_{H}$ Se) concentration of selenised yeast (SeY) on body weight gain (BWG), feed conversion efficiency (FCE), and concentrations of CLA isomers and other fatty acids (FA) in the liver and femoral muscle of rats. The investigation was performed on 48 female rats (Wistar, Hsd Brl Han: WIST), aged 8 weeks with an initial body weight of  $195.7 \pm 0.8$  g. After one week of submaintenance feeding, for 6 weeks the rats were fed *ad libitum* the Labofeed H diet supplemented with 1.5% CLAmix with or without low ( $0.2 \mu\text{g Se/g diet}$ ) or high ( $0.5 \mu\text{g Se/g diet}$ ) concentration of Se as SeY. The diet enriched with  $_{L}$ Se or  $_{H}$ Se elevated the BWG of rats compared with the control group, while the  $_{L}$ Se diet resulted in the most efficient increase in BWG. The diet containing CLAmix, irrespective of the presence of SeY, stimulated the accumulation of Se in the liver and muscles in comparison with the diet enriched with SeY. The addition of SeY to the diet enriched with CLAmix stimulated the accumulation of *cis9trans11* (*c9t11*), *t10c12* and *cc* isomers of CLA in the liver. The diet containing CLAmix and  $_{L}$ Se most efficiently increased the concentration of these CLA isomers in muscles. The diets enriched with SeY increased the concentration of essential, endogenous and total amino acids (AA) in the liver, whereas the diets enriched with SeY or CLAmix reduced the concentration of these AA in muscles. The diets enriched with CLAmix and/or SeY decreased the  $\Delta 9$ -desaturase index in the liver and muscles compared with the control group. These diets increased  $\Delta 4$ -,  $\Delta 5$ - and  $\Delta 6$ -desaturase indexes in muscles, but significantly reduced the elongase index compared with the control group. Our study shows that dietary CLAmix increased the concentration of C14:0, C18:0 and the sum of saturated FA (SFA) in the liver, whereas the diets enriched with SeY, irrespective of the presence of CLAmix, slightly reduced the concentration of C14:0, C18:0, and SFA in muscles compared with the control group. The diets containing SeY and/or CLAmix increased the accumulation of arachidonic acid (ArA), linolenic acid ( $\alpha$ LNA) and *c4c7c10c13c16c19C22:6* in the liver compared with the control rats. Negative effects of dietary SeY and/or CLAmix on the accumulation of  $\alpha$ LNA, *c5c8c11c14c17C20:5*, *c7c10c13c16c19C22:5*, *c4c7c10c13c16c19C22:6*, linoleic acid, ArA and *c8c11c14C20:3* in muscles were found out. The CLAmix diet increased the  $\Delta 4$ -,  $\Delta 5$ -desaturase and elongase indexes in the liver. It also increased the  $\Delta 4$ -,  $\Delta 5$ - and  $\Delta 6$ -desaturase indexes in muscles, but significantly reduced the elongase index compared with the control group. The finding that the diet with CLAmix and  $_{H}$ Se fed to rats decreased total FA and most efficiently increased the content of Se and essential AA in muscles is valuable information for nutritionists carrying out research on farm animals to improve the nutritive value of food from the aspect of human health.

**Keywords:** selenised yeast; conjugated linoleic acid isomers; rats; liver; femoral muscles; fatty acids; amino acids; minerals

Partly supported by the Ministry of Science and Higher Education, Poland (Grant No. N311 3364 33).

Selenium (Se) in tissues of mammals reflects the chemical form of Se and its level in consumed diets. Therefore, it is important to know its abundance or deficiency in food and diets and to determine the correct balance of Se and its effect on the level of other substances in farm animals and in humans consuming livestock products. Se supplementation of livestock diets increases the nutritional quality of animal products. Se is usually dosed in two forms: inorganic compounds (mostly selenite or selenate) or organic forms such as selenised yeast (SeY), in which selenomethionine (Se-Met) is the prevalent form of Se (Rayman, 2004). Endogenous Se is present in fluids and tissues of mammals as selenocysteine (Se-Cys), which is the functional core of ~25 Se-proteins (like isozymes of the glutathione-peroxidase (GPx) family or thioredoxin reductases) or Se-Met, which can be bound non-specifically to proteins (Se-Met-proteins) (Suzuki, 2005; Juniper et al., 2008). Se incorporated into Se-Met-proteins can be utilised in mammals during periods of suboptimal Se dosages. Despite the different metabolic pathways for Se-containing compounds, all Se compounds seem to ultimately enter the Sec-loaded tRNA pool to be used for Se-Cys-protein synthesis (Schomburga et al., 2004). These Se-proteins (i.e. Se-enzymes) are involved in diverse metabolic processes and display unique qualities. Numerous studies have shown that nearly half of these Se-Cys-proteins (like the GPx family or selenoprotein P) protect against oxidative stress. Indeed, the GPx enzyme recycles glutathione, reducing lipid peroxidation by catalysing the reduction of peroxides, including hydrogen peroxide. All Se-enzymes in their reduced state catalyse the breakdown of lipid hydroperoxides and hydrogen peroxides in mammalian cells (Navarro-Alarcon and Cabrera-Vique, 2008). Furthermore, the antioxidant properties of Se in these enzymes can help to alleviate the damage caused by ultraviolet- $\beta$  radiation in humans and animals.

It was found out in numerous studies on laboratory animals that the concentration of polyunsaturated fatty acids (PUFA) was positively correlated with the level of Se in diets (Crespo et al., 1995). Considering the above and the potential importance of conjugated linoleic acid (CLA) isomers and Se in living organisms, essentially related to protection against oxidative stress, it was desirable to study the extent to which various dietary levels of SeY may contribute to the

concentration of CLA isomers and other fatty acids (FA), especially PUFAn-3, in the liver and femoral muscles of rats. Indeed, CLA isomers are believed to play an important role in many physiological functions and may act to convey body composition or to modulate immune function (Hur et al., 2007). CLA isomers are also believed to inhibit tumorigenesis by interfering in the metabolism of some carcinogens. Moreover, some CLA isomers are antioxidative, antiatherosclerotic and hypolipidemic.

Therefore, the main purpose of the present study was to establish the effect of different amounts of Se (as SeY) added to a diet enriched with CLA isomers (CLAMix) on the concentrations of selected amino acids and fatty acids in the liver and femoral muscles of rats. The second objective of the present study was to investigate the influence of these dietary additives on the growth of rats and concentrations of Se, Zn, Fe, Cu, Ca, and Mg in the liver and muscles.

## MATERIAL AND METHODS

### Animals, housing, diets, experimental design and sampling

The experiment was conducted with 48 female rats (Wistar, Hsd Brl Han: WIST) aged 8 weeks, of an initial body weight about  $195.7 \pm 0.8$  g. The animals were housed and handled in accordance with protocols approved by the Local Animal Care and Use Committee (Agricultural University of Warsaw, Poland). The animals were housed individually in metabolic cages at a temperature of  $22 \pm 1^\circ\text{C}$  with a 12 h light-dark cycle and relative humidity of 50–60%. Each group comprised eight rats. The rats were fed a commercial diet for laboratory rats and mice (the trade name Labofeed H diet). This diet was purchased from the Feeds and Concentrates Production Plant in Kcynia, MORAWSKI Co., Poland ([www.wp-morawski.com.pl/Labofeed/labofeed](http://www.wp-morawski.com.pl/Labofeed/labofeed)). Since 1998 the Labofeed H diet has been produced according to ISO 9001 Standards (Pastuszewska et al., 2000). For our studies, this commercial diet containing  $0.13 \mu\text{g Se}/(\text{g diet})$  (the natural content of Se) was enriched with only  $0.2 \mu\text{g Se}/(\text{g diet})$  as selenite (Table 1). Therefore, the total concentration of Se in this commercial rat diet was  $0.33 \mu\text{g Se}/(\text{g diet})$ .

Table 1. Chemical composition and energy content of the basal diet<sup>#</sup> (the Labofeed H diet manufactured by MORAWSKI Co., Poland) and selenised yeast (SeY)

Item	Content	Item	Content
<b>Composition of the basal diet*</b>		<b>Composition of selenized yeast (SeY)</b>	
Dry matter <sup>d</sup> (g/100 g diet)	88.2 ± 0.9	Se (mg/g DM)	1.8
Crude protein (g/100 g diet)	21.8 ± 1.3	Sum of identified Se species (%)	88.3
Lysine (g/100 g diet)	1.31	Seleno-methionine (%)	83.0
Methionine and cysteine (g/100 g diet)	0.76	Seleno-cysteine (%)	5.0
Tryptophan (g/100 g diet)	0.28	Selenite (%)	0.3
Threonine (g/100 g diet)	0.87	Fatty acids <sup>§</sup> (mg/g DM)	
Crude fibre (g/100 g diet)	21.8 ± 1.3	C16:0	9.0
Crude fat (g/100 g diet)	3.0 ± 0.8	<i>cis</i> 9C16:1	4.1
Ash (g/100 g diet)	5.9 ± 0.6	C18:0	13.6
N total	3.77	<i>cis</i> 9C18:1	11.3
N protein	3.17	<i>cis</i> 11C18:1	0.8
P total	0.75	<i>cis</i> 9 <i>cis</i> 12C18:2 (LA)	14.7
Mono-carbohydrates	5.75	<i>cis</i> 9 <i>cis</i> 12 <i>cis</i> 15C18:3 (αLNA)	0.16
Starch	30.3		
Metabolize energy <sup>†</sup> (MJ/kg)	13.9		

<sup>#</sup>Means of 9 samples

\*Ingredients: maize, wheat, oat flakes, green meal, soya-bean oil-meal, fish meal, soya oil, vitamins (per kg diet: vit. A 10 096 U, vit. D<sub>3</sub> 2000 IU, vit. E 86.1 mg, vit. K 1.3 mg, vit. B<sub>1</sub> 15.7 mg, vit. B<sub>2</sub> 16.0 mg, vit. B<sub>6</sub> 5.24 mg, vit. B<sub>12</sub> 81 mg, biotin 0.2 mg, folic acid 3.03 mg, nicotinic acid 79.3 mg, pantothenic acid 25.5 mg; choline 2.02 g); minerals (contained per g DM diet: Na – 3.60 mg; K – 8.30 mg; Ca – 10.68 mg; P – 7.60 mg) and trace elements (contained per kg DM diet: Se as Na<sub>2</sub>SeO<sub>3</sub> – 0.2 mg; Cu – 13.9 mg; Zn – 98 mg; Mn – 112 mg; Fe – 698 mg; Mg – 1653 mg)

<sup>d</sup>The concentrations of main fatty acids (mg per kg DM diet): C8:0 – 37; C10:0 – 6; C12:0 – 11; *cis*9*cis*12*cis*15 C18:3 (αLNA) – 8; *cis*6*cis*9*cis*12C18:3 (γLNA) – 715; C14:0 – 11; *cis*9*cis*12C18:2 (LA) – 429; C16:0 – 250; *cis*9C18:1 – 187; *cis*6C18:1 – 112; C18:0 – 89; C20 – 11; C22 – 2.4; Σ saturated fatty acids (SFA) – 417; Σ polyunsaturated fatty acids (PUFA) – 1499; Σ assayed fatty acids (ΣFA) – 1915

<sup>†</sup>The mean from 3 samples

<sup>§</sup>Main fatty acid peaks (i.e. 95% of peak areas of all fatty acids in a GC-MS chromatogram)

During a one-week preliminary period the rats were fed the Labofeed H diet offered at a sub-maintenance level to reduce the rats' body fat (Table 2). Next, for 6 weeks the rats were fed *ad libitum* the experimental diets supplemented with 1.5% CLA isomer mixture (CLAmix), 0.2 µg Se/(g diet) (<sub>L</sub>Se) or 0.5 µg Se/(g diet) (<sub>H</sub>Se), Se as selenised yeast (SeY) (Tables 1 and 2). Feed intake and body weight of rats were measured weekly. At the end of the six-week experiment the rats were euthanized with CO<sub>2</sub>. The liver and femoral muscles were removed, weighed, and frozen until analysed. The concentrations of fatty acids (FA), selenium (Se), zinc (Zn),

iron (Fe), copper (Cu), calcium (Ca) and magnesium (Mg) were determined in the liver and muscles. All liver and muscle samples were analysed individually. The concentrations of FA, Zn, Fe, Cu, Ca, and Mg were calculated based on freeze-dried liver and muscle samples.

## Reagents

All chemicals were of analytical grade and organic solvents were of HPLC grade. Dichloromethane (DCM), KOH, NaOH, Na<sub>2</sub>SO<sub>4</sub>, and

conc. HCl were purchased from POCH (Gliwice, Poland). Acetonitrile, methanol, and n-heptane (99%, GC) were supplied by Lab-Scan (Ireland), while the CLA isomer mixture (2.1% *tt*CLA, 7.1% *c11t13*CLA, 40.8% *c9t11*CLA, 41.3% *t10c12*CLA, 6.7% *c8t10*CLA, and 2.0% *cc*CLA) by the Industrial Chemistry Research Institute (Warsaw, Poland). The concentration ratio ( $R_{c9t11CLA/t10c12CLA}$ ) of *c9t11*CLA to *t10c12*CLA in the dietary CLA isomer mixture was 0.9879. Fatty acid methyl ester (FAME) standards and 50% BF<sub>3</sub> in methanol were purchased from Supelco and Sigma (Bellefonte, Pennsylvania, USA).

The selenised yeast (*Se-Saccharomyces cerevisiae*) was donated by Sel-Plex (a non-commercial yeast sample; Alltech Inc., Nicholasville, USA). About 83% of the total Se concentration of selenised yeast (SeY) is represented by Se in the form of Se-Met incorporated into proteins of *Saccharomyces cerevisiae* (Table 1) (Rayman, 2004; Weiss and Hogan, 2005).

Water used for the preparation of mobile phases and chemical reagents was prepared using an Elix™ water purification system (Millipore). The mobile phases were filtered through a 0.45 µm membrane filter (Millipore).

### Analytical methods

The Se concentration in the liver and muscles was analysed by the fluorimetric method of Rodriguez et al. (1994) using fresh samples of these tissues. The concentrations of Se were calculated per dry mass (DM) using the relation between fresh mass and mass of freeze-dried liver and femoral muscle samples. The concentrations of Zn, Fe, Cu, Mg, and Ca in freeze-dried liver and muscle samples were determined by flame (air-acetylene) atomic absorption spectrometry (Korniluk et al., 2006).

The methods of alkaline hydrolysis and base- and acid-catalyzed methylation of fatty acids (i.e. the derivatization) were used as previously described (Czauderna et al., 2007a). The methylated CLA isomers (CLA-ME) were determined using silver ion liquid chromatography with photodiode array detection (Ag<sup>+</sup>-HPLC-DAD) according to Czauderna et al. (2005).

The concentrations of methylated FA (i.e. FAME and CLA-ME) and Zn, Fe, Cu, Mg, and Ca were calculated based on freeze-dried liver and muscle samples.

### Analytical equipment

The analyses of methylated fatty acids (FAME) were performed on a SHIMADZU GC-MS-QP2010 Plus EI equipped with a BPX70 fused silica capillary column (120 m × 0.25 mm i.d. × 0.25 µm film thickness); SHIM-POL, a quadrupole mass selective (MS) detector (Model 5973N) and an injection port. Helium as the carrier gas operated at a constant pressure (223.4 kPa) and flow rate of 1 ml per min. Injector and MS detector temperatures were maintained at 200 and 240°C, respectively. The total FAME profile in a 1-µl sample at the split ratio of 10:1 was determined using the column temperature gradient programme, which was described previously (Czauderna et al., 2005).

FAME identification was validated based on electron impact ionisation spectra of FAME and compared with authentic FAMS standards and the NIST 2007 reference mass spectra library.

Methylated CLA isomers (CLA-ME) were determined using a Waters 625 LC system that included a controller for gradient elution and two pumps (Waters Model 515). The apparatus consisted of a Waters 712 WISP autosampler, two ion-exchange columns loaded with silver ions (250 × 4.6 mm Chrompack ChromSper 5 µm Lipids, the Netherlands) and a Waters 996 photodiode array detector (Czauderna et al., 2005).

The concentrations of Zn, Fe, Cu, Mg, and Ca in lyophilised liver and muscle samples were determined using a PU9100X Atomic Absorption Spectrometer (UNICAM, Philips) (Korniluk et al., 2006).

### Statistical analyses

Results are presented as means ± SD of 8 individually analysed samples of the liver and muscles. The one-factorial statistical analyses of the effects of SeY or the CLAmix in the experimental diets were conducted using the non-parametric Mann-Whitney U test for comparing independent experimental groups. Statistical analyses of the interactions between CLAmix and  ${}_L\text{Se} ({}_L\text{Se}_{CLA})$  and CLAmix and  ${}_H\text{Se} ({}_H\text{Se}_{CLA})$  were performed using two-factorial ANOVA analyses. Differences were considered significant at the  $P < 0.05$  (\*) and  $P < 0.01$  (\*\*) levels, while at  $P < 0.1$  (°) differences were indicated as tendencies. Statistical analyses were performed using the Statistica v. 6 software package (Statistica, 2002).

Table 2. Dietary effects of 1.5% CLA isomer mixture (CLAmix) and selenised yeast (SeY) on feed conversion efficiency (FCE; g body weight gain/g feed intake)<sup>#</sup>, body weight gain (BWG)<sup>#</sup> and liver fresh mass (g) of rats and the concentrations\* (µg/g) of Se, Zn, Cu, Fe, Mg, Ca in the liver and femoral muscles of rats after 6 weeks of feeding the experimental diets

Group	Additives per g diet	Body weight (g) Initial adapted <sup>&amp;</sup>	BWG (g)	FCE (g/g)	Liver					Femoral muscles <sup>§</sup>							
					Liver mass (g)	Se <sup>t</sup>	Zn	Cu	Fe	Ca	Mg	9Se <sup>t</sup>	Zn	Fe	Ca	Mg	
Control <sup>‡</sup>	–	196.0	180.4	50.0 <sup>ab</sup>	0.068	8.718	4.3 <sup>α</sup>	120	6.3 <sup>AB</sup>	1036 <sup>α</sup>	206 <sup>abx</sup>	580 <sup>A</sup>	0.25 <sup>Aa</sup>	49	68	246 <sup>a</sup>	869
<sub>L</sub> Se <sup>‡</sup>	0.2 µg Se	196.7	179.2	58.1 <sup>a</sup>	0.076 <sup>a</sup>	8.408	4.6 <sup>a</sup>	129	10.0	1047 <sup>β</sup>	149 <sup>α</sup>	607 <sup>B</sup>	0.44 <sup>B</sup>	47	74 <sup>A</sup>	256	892 <sup>a</sup>
<sub>H</sub> Se <sup>‡</sup>	0.5 µg Se	195.0	177.3	55.6 <sup>b</sup>	0.075 <sup>c</sup>	8.315	5.1 <sup>b</sup>	125 <sup>α</sup>	8.9 <sup>AC</sup>	1067 <sup>α</sup>	134 <sup>a</sup>	651 <sup>C</sup>	0.72 <sup>ab</sup>	44	79 <sup>a</sup>	230 <sup>a</sup>	844
CLA <sup>‡</sup>	CLAmix	196.5	181.7	51.9 <sup>cd</sup>	0.069 <sup>b</sup>	8.486	4.6 <sup>cc</sup>	128	10.2 <sup>BDx</sup>	1061	144 <sup>b</sup>	682 <sup>A</sup>	0.49 <sup>A</sup>	46	78	219 <sup>b</sup>	836
<sub>L</sub> Se <sup>‡</sup> <sub>CLA</sub> <sup>§</sup>	0.2 µg Se CLAmix	195.3	178.3	56.1 <sup>d</sup>	0.073 <sup>abx</sup>	8.482	4.9 <sup>ax</sup>	129	12.1 <sup>α</sup>	984 <sup>β</sup>	217	668 <sup>B</sup>	0.64 <sup>B</sup>	47	81 <sup>A</sup>	325	829 <sup>a</sup>
<sub>H</sub> Se <sup>‡</sup> <sub>CLA</sub> <sup>§</sup>	0.5 µg Se CLAmix	194.7	177.4	54.6 <sup>c</sup>	0.076 <sup>y</sup>	8.361	5.3 <sup>bcy</sup>	135 <sup>α</sup>	13.0 <sup>CD</sup>	1045	183	665 <sup>C</sup>	0.82 <sup>b</sup>	52	93 <sup>a</sup>	247 <sup>b</sup>	859

<sup>#</sup>FCE and BWG after feeding for 6 weeks with the experimental diets enriched with CLAmix and/or Se-yeast

\*The concentrations of assayed elements in dry mass (DM); the liver and femoral muscles were freeze-dried

<sup>&</sup>The body weight (g) of individually adapted rats after 7 days of submaintenance feeding (9 g the Labofeed H diet per day per rat)

<sup>§</sup>The concentration of Cu in the muscles was below the quantification limit

<sup>‡</sup>Means in columns sharing the same letter are significantly different

<sup>a,b</sup> $P < 0.05$  and <sup>A,B</sup> $P < 0.01$ ; <sup>§</sup>Interactions of CLAmix × <sub>L</sub>Se and CLAmix × <sub>H</sub>Se are significant at <sup>x,y</sup> $P < 0.05$  and <sup>x,y</sup> $P < 0.01$ , respectively

<sup>†</sup>Fresh liver and muscle samples were used to determine Se by the fluorimetric method

Table 3. The effect of diets enriched with CLAmix and 0.2 or 0.5  $\mu\text{g Se}/(\text{g diet})$  as SeY ( ${}_{\text{L}}\text{Se}$  and  ${}_{\text{H}}\text{Se}$ ) on the concentration of CLA isomers ( $\mu\text{g/g}$ )<sup>&</sup>, essential (EAA; mg/g DM) and endogenous (NEAA; mg/g DM) amino acids, the sum of amino acids ( $\Sigma\text{AA}$ ; mg/g DM), methionine (Met; mg/g DM), *c9C18:1* (mg/g), SFA (mg/g), MUFA (mg per g), PUFA<sub>n-3</sub> (mg/g), PUFA<sub>n-6</sub> (mg/g), the sum of FA ( $\Sigma\text{FA}$ ; mg/g), values of  $\Delta 9$ -desaturase ( $\Delta 9_{\text{index}}$ )<sup>§</sup>, atherogenic ( $A_{\text{index}}$ )<sup>\*</sup>, thrombogenic ( $T_{\text{index}}$ )<sup>||</sup>indexes<sup>d</sup> in the liver and muscles of rats

Group	Control	${}_{\text{L}}\text{Se}$	${}_{\text{H}}\text{Se}$	CLA	${}_{\text{L}}\text{Se}_{\text{CLA}}$	${}_{\text{H}}\text{Se}_{\text{CLA}}$	Significance of effects analysis				
							one factorial			interactions	
							${}_{\text{L}}\text{Se}$	${}_{\text{H}}\text{Se}$	CLA	${}_{\text{L}}\text{Se}_{\text{CLA}}$	${}_{\text{H}}\text{Se}_{\text{CLA}}$
<b>Liver</b>											
<i>t</i> CLA	– <sup>h</sup>	–	–	2.1 <sup>Aa</sup>	6.5 <sup>A</sup>	3.6 <sup>a</sup>	–	–	–	–	–
$\alpha$ CLA	–	–	–	0.9 <sup>ab</sup>	1.3 <sup>a</sup>	1.4 <sup>b</sup>	–	–	–	–	–
<i>c9t11</i>	–	–	–	181 <sup>a</sup>	231	255 <sup>a</sup>	–	–	–	–	–
<i>t10c12</i>	–	–	–	87	118	122	–	–	–	–	–
$\Sigma\text{CLA}$	–	–	–	271 <sup>ab</sup>	367 <sup>a</sup>	402 <sup>b</sup>	–	–	–	–	–
EAA <sup>l</sup>	262	278	279	272	266	268	NS	NS	NS	NS	NS
NEAA <sup>l</sup>	280	310	312	282	256	306	NS	NS	NS	α	NS
$\Sigma\text{AA}$ <sup>l</sup>	542	589	591	554	522	574	NS	NS	NS	α	NS
Met <sup>l</sup>	14.5	15.4	20.2	13.0	8.6	10.5	NS	α	NS	*	*
$\Delta 9_{\text{index}}$	0.156	0.132	0.123	0.115	0.119	0.129	NS	NS	α	NS	NS
$A_{\text{index}}$	0.287	0.256	0.263	0.340	0.257	0.264	α	NS	NS	NS	NS
$T_{\text{index}}$	0.688	0.604	0.632	0.634	0.566	0.577	**	NS	NS	α	α
<i>c9C18:1</i>	2.19	1.88	1.84	1.89	1.93	2.01	NS	NS	NS	NS	NS
SFA	13.8	14.0	14.8	16.4	15.4	15.4	NS	NS	*	NS	NS
MUFA	3.07	2.63	2.50	2.49	2.50	2.67	NS	NS	NS	NS	α
PUFA <sub>n-3</sub>	3.76	4.79	4.84	6.01	5.72	5.40	**	*	**	NS	α
PUFA <sub>n-6</sub>	12.1	12.5	12.8	9.2	14.4	13.7	NS	NS	NS	α	α
$\Sigma\text{FA}$	32.7	34.0	34.9	34.3	38.2	37.4	NS	NS	NS	*	*

## RESULTS

### The effect of dietary SeY and CLAmix on body weight gain, feed conversion efficiency and levels of elements in liver and muscles

The influence of diets enriched with CLAmix and/or SeY on body weight gain (BWG), feed conversion efficiency (FCE), liver weight and concentrations of Se, Zn, Fe, Cu, Ca, and Mg in both tissues is summarized in Table 2. As can be seen from the obtained results, the diet enriched with  ${}_{\text{L}}\text{Se}$  or  ${}_{\text{H}}\text{Se}$  elevated the BWG of rats ( $P < 0.05$ ) compared with the control group, while the lower concentration of extra Se in the

diet resulted in the most efficient increase in the BWG of rats. On the other hand, the diet containing only CLAmix exerted a negligible influence on BWG in comparison with control rats, whereas the diets with CLAmix and SeY, regardless of the concentration of extra SeY, increased the value of BWG compared with rats fed the control diet or the diet enriched with CLAmix, although no statistically significant interactions (i.e.  ${}_{\text{L}}\text{Se} \times \text{CLAmix}$ ,  ${}_{\text{H}}\text{Se} \times \text{CLAmix}$ ) were determined ( $P > 0.05$ ).

The diets containing SeY (as  ${}_{\text{L}}\text{Se}$  or  ${}_{\text{H}}\text{Se}$ ), regardless of the presence of CLAmix, resulted in a decrease in the liver weight in comparison with the control group, however, these differences

Table 3 to be continued

Group	Control	LSe	HSe	CLA	LSe <sub>CLA</sub>	HSe <sub>CLA</sub>	Significance of effects analysis				
							one factorial			interactions	
							LSe	HSe	CLA	LSe <sub>CLA</sub>	Se <sub>CLA</sub>
<b>Femoral muscles</b>											
ttCLA	–	–	–	26 <sup>ab</sup>	41 <sup>a</sup>	56 <sup>b</sup>	–	–	–	–	–
ccCLA	–	–	–	16	18	17	–	–	–	–	–
c9t11	–	–	–	3172 <sup>a</sup>	3586	2596 <sup>a</sup>	–	–	–	–	–
t10c12	–	–	–	1954	2260	1570	–	–	–	–	–
ΣCLA	–	–	–	5888 <sup>ab</sup>	6932 <sup>a</sup>	4857 <sup>b</sup>	–	–	–	–	–
EAA <sup>L</sup>	320	263	188	211	290	325	NS	*	NS	α	NS
NEAA <sup>L</sup>	376	345	252	275	354	369	NS	*	NS	NS	NS
ΣAA <sup>L</sup>	696	608	440	485	644	694	NS	*	NS	α	NS
Met <sup>L</sup>	19.1	19.5	12.3	15.4	20.1	21.3	α	*	NS	NS	α
Δ <sup>9</sup> <sub>index</sub>	0.474	0.454	0.457	0.458	0.458	0.459	*	α	α	NS	NS
A <sub>index</sub>	0.332	0.337	0.335	0.365	0.359	0.367	NS		α	NS	α
T <sub>index</sub>	0.320	0.329	0.336	0.389	0.380	0.408	NS	α	**	NS	α
c9C18:1	17.3	13.0	13.3	15.6	15.5	13.7	*	*	NS	NS	*
SFA	20.6	16.5	16.6	19.4	19.6	17.0	*	NS	NS	NS	NS
MUFA	20.5	15.2	15.6	18.3	18.4	15.8	*	*	NS	NS	*
PUFA <sub>n-3</sub>	13.0	10.6	10.0	9.5	9.8	7.7	α	α	*	NS	α
PUFA <sub>n-6</sub>	23.6	18.9	19.1	21.3	21.6	18.7	*	*	NS	NS	NS
ΣFA	83.4	64.5	64.8	76.6	78.6	66.1	*	*	NS	NS	NS

&Means in rows sharing the same letter are significantly different

<sup>a,b</sup> $P < 0.05$  and <sup>A,B</sup> $P < 0.01$ ; ΣCLA – a sum of CLA isomers

<sup>§</sup>Δ<sup>9</sup>-desaturase index = (c9C14:1 + c9C16:1 + c9C18:1)/(c9C14:1 + c9C18:1 + c9C16:1 + C14:0 + C16:0 + C18:0)

\*The atherogenic index = (C12:0 + 4\*C14:0 + C16:0)/(MUFA + PUFA<sub>n-6</sub> + PUFA<sub>n-3</sub>)

<sup>††</sup>The thrombogenic index = (C14:0 + C16:0 + C18:0)/0.5\*MUFA + 0.5\*PUFA<sub>n-6</sub> + 3\*PUFA<sub>n-3</sub> + PUFA<sub>n-3</sub>/PUFA<sub>n-6</sub>)

<sup>d</sup>Ulbricht and Southgate (1991)

<sup>L</sup>The content of amino acid(s) in the dry mass of the liver and muscles (i.e. mg/g DM)

<sup>h</sup>Below the quantification limit (L<sub>Q</sub>)

were not significant ( $P > 0.05$ ) (Table 2). On the other hand, the Se concentration increased in the liver and muscles of rats fed the diets enriched with SeY and with or without CLAmix compared with control rats, although this increase was relatively lower in the liver than in muscles. The diet containing CLAmix, irrespective of the presence of SeY, stimulated the accumulation of Se in both tissues in comparison with the diet enriched with SeY (as LSe or HSe). All experimental diets also caused a numerical or statistically significant increase in the concentrations of Zn, Cu, and

Mg in the liver, whereas they resulted only in an increase in the Fe concentration in muscles in comparison with control rats (Table 2).

### The effect of experimental diets on the concentration of CLA isomers, amino acids and PUFA in liver and muscles

As can be seen from the data summarized in Table 3, the addition of SeY to the diet enriched with CLAmix elevated the concentration of c9t11,

Table 4. The effect of diets enriched with CLAmix and 0.2 or 0.5  $\mu\text{g Se}/(\text{g diet})$  as SeY ( $_{\text{L}}\text{Se}$  and  $_{\text{H}}\text{Se}$ ) on the concentrations (mg/g) of C14:0, linolenic acid ( $\alpha\text{LNA}$ ), linoleic acid (LA), C18:0,  $c8c11c14C20:3$  ( $C20:3_{n-6}$ ),  $c5c8c11c14C20:4$  (ArA),  $c5c8c11c14c17C20:5$  ( $C20:5_{n-3}$ ),  $c7c10c13c16c19C22:5$  ( $C22:5_{n-3}$ ),  $c4c7c10c13c16c19C22:6$  ( $C22:6_{n-3}$ ), values of the PUFA-to-SFA, PUFA-to- $\Sigma\text{FA}$ , UFA-to- $\Sigma\text{FA}$ , PUFA $n-6$ -to-PUFA $n-3$  (n-6/n-3) ratios, values of the  $\Delta 4$ -( $\Delta 4_{\text{index}}$ )<sup>&</sup>,  $\Delta 5$ -( $\Delta 5_{\text{index}}$ )<sup>&</sup> and  $\Delta 6$ -( $\Delta 6_{\text{index}}$ )<sup>#</sup> desaturase, elongase ( $\text{Elong}_{\text{index}}$ )<sup>\*</sup> indexes in the liver and muscles of rats

Group	Control	$_{\text{L}}\text{Se}$	$_{\text{H}}\text{Se}$	CLA	$_{\text{L}}\text{Se}_{\text{CLA}}$	$_{\text{H}}\text{Se}_{\text{CLA}}$	Significance of effects analysis				
							one factorial			interactions	
							$_{\text{L}}\text{Se}$	$_{\text{H}}\text{Se}$	CLA	$_{\text{L}}\text{Se}_{\text{CLA}}$	$_{\text{H}}\text{Se}_{\text{CLA}}$
<b>Liver</b>											
C14:0	0.053	0.051	0.052	0.066	0.051	0.052	NS	NS	NS	NS	NS
$\alpha\text{LNA}$	0.366	0.562	1.088	1.365	0.699	0.490	**	**	**	$\alpha$	*
LA	7.16	6.79	7.23	7.05	8.00	7.72	NS	NS	NS	NS	NS
C18:0	7.69	8.30	8.86	9.33	8.83	8.47	*	**	**	NS	NS
C20:3n-6	0.134	0.209	0.139	0.073	0.175	0.223	**	NS	NS	*	*
ArA	4.79	5.54	5.40	5.21	6.05	5.61	NS	NS	NS	$\alpha$	NS
C20:5n-3	1.05	1.36	1.16	0.939	0.974	1.12	**	$\alpha$	NS	NS	*
C22:5n-3	0.701	0.906	0.793	0.477	0.952	0.973	*	NS	NS	NS	*
C22:6n-3	1.65	1.96	1.79	3.19	3.05	2.77	*	NS	**	*	$\alpha$
<u>PUFA</u> SFA	1.146	1.234	1.188	1.134	1.322	1.261	*	NS	NS	*	NS
<u>PUFA</u> $\Sigma\text{FA}$	0.484	0.510	0.504	0.496	0.532	0.518	**	NS	NS	$\alpha$	NS
<u>UFA</u> $\Sigma\text{FA}$	0.578	0.587	0.576	0.562	0.598	0.589	NS	NS	NS	NS	NS
n-6/n-3	3.21	2.61	2.64	2.06	2.51	2.54	**	**	**	$\alpha$	NS
$\Delta 4_{\text{index}}$	0.701	0.684	0.693	0.870	0.762	0.740	NS	NS	*	$\alpha$	$\alpha$
$\Delta 5_{\text{index}}$	0.973	0.964	0.975	0.986	0.972	0.962	*	NS	$\alpha$	NS	NS
$\Delta 6_{\text{index}}$	0.982	0.970	0.981	0.990	0.979	0.972	**	NS	NS	NS	NS
$\text{Elong}_{\text{index}}$	0.599	0.601	0.594	0.663	0.506	0.535	NS	NS	*	*	$\alpha$

$t10c12$  and  $cc$  isomers of CLA in the liver in comparison with the concentrations of these isomers in the liver of rats fed the diet with CLAmix only; the increase in the concentration of these isomers in the liver was observed with the increase in the SeY concentration in the diet with CLAmix. On the other hand, the lower concentration of extra SeY in the diet containing CLAmix most efficiently increased the concentration of these CLA isomers in muscles. The higher concentration of SeY in the diet with CLAmix resulted in a decrease in the concentrations of  $c9t11\text{CLA}$  and  $t10c12\text{CLA}$  in muscles in comparison with the control group.

The diets enriched with SeY (as  $_{\text{L}}\text{Se}$  and  $_{\text{H}}\text{Se}$ ) increased the concentration of methionine (Met), essential (EAA), endogenous amino acids (NEAA) and the sum of all amino acids ( $\Sigma\text{AA}$ ) in the liver,

although no significant differences ( $P > 0.05$ ) in these values were found out (Table 3). On the other hand, these diets usually resulted in a decrease in the concentration of these amino acids in muscles, however, significant differences ( $P < 0.05$ ) in these values were determined in muscles of rats fed the diet enriched with the higher concentration of SeY (i.e.  $_{\text{H}}\text{Se}$ ). The results show that the addition of CLAmix to the diet containing  $_{\text{L}}\text{Se}$  or  $_{\text{H}}\text{Se}$  reduced the effect of dietary  $_{\text{L}}\text{Se}$  or  $_{\text{H}}\text{Se}$  on the concentrations of EAA, NEAA,  $\Sigma\text{AA}$  and Met in the liver and muscles compared with these values in the liver and muscles of rats fed the diets with  $_{\text{L}}\text{Se}$  or  $_{\text{H}}\text{Se}$ .

The diets enriched with CLAmix and/or SeY (as  $_{\text{L}}\text{Se}$  or  $_{\text{H}}\text{Se}$ ) reduced the values of  $\Delta 9$ -desaturase index ( $\Delta 9_{\text{index}}$ ) in the liver and muscles of rats



Table 4 to be continued

Group	Control	LSe	HSe	CLA	LSe <sub>CLA</sub>	HSe <sub>CLA</sub>	Significance of effects analysis				
							one factorial			interactions	
							LSe	HSe	CLA	LSe <sub>CLA</sub>	HSe <sub>CLA</sub>
<b>Femoral muscles</b>											
C14:0	1.090	0.822	0.766	0.908	0.933	0.702	NS	α	NS	NS	α
αLNA	10.44	8.09	8.06	7.98	8.10	6.15	αα	α	α	NS	*
LA	22.1	17.7	17.9	20.3	20.2	17.6	*	α	NS	NS	NS
C18:0	4.18	3.51	3.52	3.80	3.84	3.20	NS	NS	NS	NS	NS
C20:3n-6	0.054	0.040	0.034	– <sup>π</sup>	0.038	0.019	NS	α	**	NS	NS
ArA	1.44	1.14	1.09	0.67	0.91	0.80	**	**	**	α	α
C20:5n-3	0.150	0.118	0.098	0.023	0.065	0.043	NS	α	**	*	*
C22:5n-3	0.818	0.710	0.643	0.360	0.354	0.339	NS	*	**	NS	NS
C22:6n-3	1.53	1.34	1.23	0.92	0.97	0.95	*	**	**	NS	NS
PUFA SFA	2.05	1.99	1.96	2.01	2.07	1.96	NS	NS	NS	NS	NS
PUFA ΣFA	0.507	0.508	0.503	0.508	0.517	0.504	NS	NS	NS	NS	NS
UFA ΣFA	0.753	0.744	0.743	0.747	0.751	0.743	NS	NS	NS	NS	NS
n-6/n-3	1.823	1.845	1.900	2.247	2.209	2.439	NS	NS	**	NS	α
Δ <sub>4</sub> <sub>index</sub>	0.652	0.654	0.657	0.720	0.732	0.737	NS	NS	**	NS	α
Δ <sub>5</sub> <sub>index</sub>	0.963	0.966	0.970	1.000	0.960	0.977	NS	NS	**	NS	NS
Δ <sub>6</sub> <sub>index</sub>	0.997	0.998	0.998	1.000	0.998	0.999	NS	*	**	NS	NS
Elong <sub>index</sub>	0.155	0.143	0.132	0.977	0.156	0.113	NS	NS	**	NS	α

<sup>&</sup>The Δ<sub>4</sub>-desaturase index =  $c4c7c10c13c16c19C22:6 / (c4c7c10c13c16c19C22:6 + c7c10c13c16c19C22:5)$

<sup>§</sup>The Δ<sub>5</sub>-desaturase index =  $c5c8c11c14C20:4 / (c8c11c14C20:3 + c5c8c11c14C20:4)$

<sup>#</sup>The Δ<sub>6</sub>-desaturase index =  $c9c12C18:2 / (c9c12C18:2 + c11c14c17C20:3)$

<sup>\*</sup>The elongase index =  $c5c8c11c14c17C20:5 / (c5c8c11c14c17C20:5 + c7c10c13c16c19C22:5)$

compared with control animals (Table 3), however, in the present trials differences in Δ<sub>9</sub><sub>index</sub> values were usually non-significant ( $P > 0.05$ ). Moreover, changes in concentrations of c9C18:1 and MUFA in the liver and muscles were also consistent with changes in Δ<sub>9</sub><sub>index</sub> values in both tissues of the examined rats.

The present study documented the positive influence of SeY (as LSe or HSe) in the diets with or without CLAmix on values of atherogenic (A<sub>index</sub>) and thrombogenic (T<sub>index</sub>) indexes in the liver (Table 3), although differences in A<sub>index</sub> and T<sub>index</sub> values were usually non-significant ( $P > 0.05$ ). On the other hand, there was a negligible effect of dietary SeY on A<sub>index</sub> and T<sub>index</sub> values in muscles, whereas the simultaneous addition of CLAmix and SeY to the diet resulted in an increase in A<sub>index</sub> and T<sub>index</sub> val-

ues in muscles. The diet containing CLAmix and the higher concentration of SeY (i.e. HSe) however showed a tendency ( $P < 0.1$ ) to increase A<sub>index</sub> and T<sub>index</sub> values in muscles. Rats fed the diet containing CLAmix tended to increase the A<sub>index</sub> ( $P = 0.07$ ) and significantly ( $P < 0.01$ ) increased the T<sub>index</sub> in muscles, whereas the dietary CLAmix elevated only the A<sub>index</sub> in the liver, although no significant differences ( $P > 0.05$ ) between these values were found out (Table 3).

The results show (Tables 3 and 4) that dietary CLAmix increased the concentration of C14:0, C18:0 as well as the sum of saturated fatty acids (SFA) in the liver compared with the control group ( $P = 0.12$ ,  $P < 0.01$  and  $P < 0.05$ , respectively) and non-significantly the concentration of C14:0, C18:0 and SFA in the liver in comparison with rats fed the

diet containing only SeY (as  $_L$ Se or  $_H$ Se). The addition of  $_L$ Se or  $_H$ Se to the diet with CLAmix slightly reduced the concentration of C14:0, C18:0, and SFA in the liver compared with this value in the liver of rats fed the diet containing only CLAmix. On the other hand, the diets enriched with SeY (as  $_L$ Se or  $_H$ Se), regardless of the presence of CLAmix, slightly reduced the concentration of C14:0, C18:0, and SFA in muscles compared with the control group, although no significant differences ( $P > 0.05$ ) in these values were usually observed.

In the present study there was usually a significant positive influence of dietary SeY (as  $_L$ Se or  $_H$ Se) and/or CLAmix on the accumulation of arachidonic acid (ArA), linolenic acid ( $\alpha$ LNA),  $c4c7c10c13c16c19C22:6$  (C22:6n-3) and PUFA-3 in the liver compared with control rats (Tables 3 and 4). On the other hand, negative effects of dietary SeY and/or CLAmix on the accumulation of PUFA-3 ( $\alpha$ LNA,  $c5c8c11c14c17C20:5$ ,  $c7c10c13c16c19C22:5$  and C22:6n-3) and PUFA-6 (linoleic acid, ArA and  $c8c11c14C20:3$ ) in muscles were found out. Increased values of the PUFA/SFA and PUFA/ $\Sigma$ F A ratios were observed in the liver of rats fed the diets containing SeY (as  $_L$ Se or  $_H$ Se) or both CLAmix and SeY (as  $_L$ Se or  $_H$ Se) compared with control rats, although statistically significant effects or a tendency were noticed only in the livers of rats fed the diet containing  $_L$ Se or  $_L$ Se and CLAmix ( $P < 0.05$ ,  $P < 0.1$ , for the interactions, respectively). On the other hand, no statistically significant differences in the values of the PUFA/SFA and PUFA/ $\Sigma$ F A ratios in muscles were observed among all rat groups. Similarly, dietary treatments did not affect the values of the UFA/ $\Sigma$ F A ratios in the liver and muscles.

The simultaneous addition of CLAmix and SeY (as  $_L$ Se or  $_H$ Se) to the diet significantly increased the concentration of the sum of all assayed fatty acids ( $\Sigma$ F A) in the liver (the interaction:  $P < 0.05$ ), while the diet containing only  $_L$ Se,  $_H$ Se or CLAmix non-significantly elevated the concentration of  $\Sigma$ F A in this organ. Dietary  $_L$ Se or  $_H$ Se treatments reduced ( $P < 0.05$ ) the concentration of  $\Sigma$ F A in muscles in comparison with the control group, whereas the addition of CLAmix to the diet, regardless of the presence of SeY, non-significantly decreased the concentration of  $\Sigma$ F A in muscles.

The values of the PUFA-6/PUFA-3 ratio in the liver were significantly decreased ( $P < 0.01$ ) by dietary SeY (as  $_L$ Se or  $_H$ Se) or CLAmix, whereas no statistically significant interactions of CLAmix  $\times$  SeY (as  $_L$ Se or  $_H$ Se) were observed ( $P < 0.1$  and

$P > 0.1$ , respectively). The diet enriched with CLAmix resulted in a statistically significant increase ( $P < 0.01$ ) in the PUFA-6/PUFA-3 ratio in muscles. Similarly, interactions of CLAmix  $\times$  SeY (as  $_L$ Se or  $_H$ Se) caused an increase in the ratio of PUFA-6/PUFA-3 in muscles, although no significant differences were observed ( $P < 0.1$  and  $P > 0.1$ , respectively).

The CLAmix diet resulted in higher values of  $\Delta 4$ -,  $\Delta 5$ -desaturase (i.e.  $\Delta 4_{\text{index}}$ ,  $\Delta 5_{\text{index}}$ ) and elongase ( $\text{Elong}_{\text{index}}$ ) indexes in the liver ( $P < 0.05$ ,  $P < 0.1$ ,  $P < 0.05$ , respectively) compared with the control group (Table 4). Similarly, this diet caused an increase in the  $\Delta 4_{\text{index}}$ ,  $\Delta 5_{\text{index}}$  and  $\Delta 6_{\text{index}}$  in muscles ( $P < 0.01$ ) and significantly reduced ( $P < 0.01$ ) the elongase index compared with the control group. The addition of SeY (as  $_L$ Se or  $_H$ Se) to the diet, regardless of the presence of CLAmix, exerted a small influence on the values of desaturase indexes in the liver and muscles, with the exception of  $\Delta 4_{\text{index}}$  values in the liver and muscles of rats fed the diet with CLAmix and SeY (as  $_L$ Se or  $_H$ Se). The diet enriched with  $_L$ Se or  $_H$ Se resulted in small changes in the values of the  $\text{Elong}_{\text{index}}$  in the liver and muscles. Surprisingly, the diet containing only CLAmix considerably decreased ( $P < 0.01$ ) the value of the  $\text{Elong}_{\text{index}}$  in muscles.

## DISCUSSION

The present studies demonstrate the ability of supplemented Se-yeast, regardless of the presence of CLAmix in the diet, to increase the body weight gain of the examined animals. Fortunately, neither macroscopic lesions nor toxic symptoms of SeY and CLA isomers were observed in rats fed experimental diets. Indeed, diets containing up to 2 mg Se/kg would not be toxic to rodents (like rats or mice) (McDowell et al., 2005; Korniluk et al., 2006).

The results of this study indicate that dietary 0.5  $\mu\text{g}$  Se/g diet as SeY exerts an inhibitory effect on the BWG in comparison with the diet enriched with 0.2 ppm Se. Thus, we argue that the diet containing 0.5  $\mu\text{g}$  Se/g diet catalyses hydrosulphide oxidation, which inhibits protein synthesis in rats (Navarro-Alarcon and Cabrera-Vique, 2008). Indeed, the diet containing the higher content of Se (i.e. 0.5  $\mu\text{g}$  Se/g diet) most efficiently decreased the concentrations of Met, EAA and NAA in muscles.

Considering our previous studies (Czuderna et al., 2004a,b; Korniluk et al., 2006) and the present

study, we argue that only the lower concentration of Se (i.e. 0.2–0.5  $\mu\text{g Se/g diet}$ ) in the diet, regardless of the presence of CLA isomers, elevated body weight gain and improved feed conversion efficiency. Our previous and present studies support the concept that a Se pro-oxidative effect of diets containing 1.2–2  $\mu\text{g Se/g diet}$  (Czauderna et al., 2004a,b; Korniluk et al., 2006) is due to the catalysis of hydrosulphide oxidation that results in an inhibitory effect on the yield of protein synthesis (Navarro-Alarcon and Cabrera-Vique, 2008). Moreover, the supranutritional levels of Se in diets may act as a pro-oxidant, which can result in peroxidative damage of unsaturated fatty acids (particularly PUFA) as well as in stimulation of oxidative stress in living organisms (Czauderna et al., 2004a,b; Juniper et al., 2008). Consequently, a diet containing 2  $\mu\text{g Se/(g diet)}$  as selenate reduced body weight gain and feed conversion efficiency in rats (Czauderna et al., 2003, 2004a,b), while the diet enriched with 0.2  $\mu\text{g Se/(g diet)}$  as SeY most effectively increased body weight gain and feed conversion efficiency (Table 2) as well as considerably increased protein synthesis in the liver, as the concentration of amino acids (AA) in this organ was higher than in control rats and practically similar to the concentration of AA in the liver of rats fed the diet with 0.5  $\mu\text{g Se/g diet}$  as SeY.

The present study demonstrated that the addition of CLAmix to a diet enriched with  ${}_{\text{L}}\text{Se}$  or  ${}_{\text{H}}\text{Se}$  positively correlated with the accumulation of Se in liver and muscles. Our present studies are thus in agreement with our previous investigations (Czauderna et al., 2007b) in which the addition of an individual isomer (1%) or an isomer mixture of CLA (2%) to a diet enriched with SeY (i.e. 1.2  $\mu\text{g Se/g diet}$ ) also stimulated the accumulation of Se in the liver and femoral muscles of rats. The possible explanation is that dietary CLAmix leads to increased oxygen consumption associated with significantly increased oxidation of fat (Belury, 2002). This higher oxygen consumption and formation of fat-oxidation products stimulated the biosynthesis of the Se-proteins that mainly protect against oxidative stress (Schomburga et al., 2004; Suzuki, 2005). So, we suggest that the stimulation of Se-protein biosynthesis by dietary CLAmix resulted in increasing the concentration of Se in the liver and muscles of rats fed the diet enriched with both CLAmix and SeY (as  ${}_{\text{L}}\text{Se}$  or  ${}_{\text{H}}\text{Se}$ ).

The present study also investigated relationships between the experimental diets and the concen-

trations of CLA isomers in the liver and muscles. The addition of 0.2 and 0.5  $\mu\text{g Se/g diet}$  to the diet enriched with CLAmix stimulated the incorporation of CLA isomers, with the exception of *t,t* isomers of CLA, in the liver; the concentrations of isomers increased as the Se content in the diet increased. Therefore, we suggest that the low level of Se in the Labofeed H diet (i.e. 0.13  $\mu\text{g Se/g}$  as the natural content of Se and extra 0.2  $\mu\text{g Se/g}$  as selenite; Table 1) is the main reason why the 7-day preliminary period followed by dietary extra Se stimulated the assimilation of Se into the rats' organisms. Considering the above, we supposed that the metabolites of Se in the liver positively interacted with CLA isomers. On the other hand, the addition of the lower amount of SeY ( ${}_{\text{L}}\text{Se}$ ) to the diet containing CLAmix resulted in the increase in concentrations of *cc*, *c9t11*, *t10c12* and in the sum of CLA isomers ( $\Sigma\text{CLA}$ ) in muscles. The chemical form of Se added to the diet is relevant to this finding; Se-Met, the main component of dietary SeY, is incorporated into muscle protein in place of methionine (Weiss and Hogan, 2005). So, it seems reasonable to suggest that formed Se-Met-proteins in muscles stimulated the incorporation of these CLA isomers into muscles. On the other hand, the addition of the higher amount of Se (0.5  $\mu\text{g Se/g diet}$ ) to the diet containing CLAmix reduced the concentrations of *cc*, *c9t11*, *t10c12* and  $\Sigma\text{CLA}$  isomers and elevated the level of Se in muscles. Considering these facts, we argue that the higher concentration of Se in muscles exerts the antagonistic and pro-oxidative effects on *cc*, *c9t11*, *t10c12* and  $\Sigma\text{CLA}$  isomers in muscles.

Our present results (Table 3) are in agreement with the observations of Alasnier et al. (2002) and our previous studies (Czauderna et al., 2003, 2004a,b, 2007b; Korniluk et al., 2006) in which *t10c12CLA* and *t10t12CLA* are more efficiently driven through  $\beta$ -oxidation in cells of liver, muscles, kidneys or adipose tissue in rodents than their 9,11 homologues. Therefore, in the present studies, the values of the ratio ( $R = c9t11\text{CLA}/t10c12\text{CLA}$ ) of these isomers in the liver (1.958–2.090) and muscles (1.587–1.654) of rats fed the diet enriched with CLAmix, regardless of the presence of SeY, were higher compared with the same ratio in the CLA isomer mixture (CLAmix) added to the Labofeed H diet ( $R = 0.988$ ). Moreover, the present studies clearly indicate that the  $\beta$ -oxidation rate of *t10c12CLA* relative to the  $\beta$ -oxidation rate of *c9t11CLA* is higher in the liver than in muscles.

Our previous experiments (Czauderna et al., 2004b) demonstrated that dietary *t,t* isomers of CLA are preferentially metabolised in comparison with the metabolism of dietary *c,c*, *c9t11* and *t10c12* CLA isomers. Considering our current results and previous studies (Czauderna et al., 2004b) it can be concluded that the addition of Se (as SeY or selenate) to the diet containing CLA isomers reduced the metabolism of *t,t* isomers of CLA in comparison with the diet containing only CLA isomers. Thus, dietary inorganic Se (as selenate) and organic Se (as SeY) exhibited a similar influence on the metabolic capacity of *tt* isomers of CLA in rats.

As expected, the diets containing CLAmix, irrespective of the presence of SeY, decreased the  $\Delta 9$ -desaturation capacity of C18:0. This is consistent with our previous results (Czauderna et al., 2003, 2004a,b, 2007b; Korniluk et al., 2006) showing that CLA isomers and/or selenate are responsible for a change in the levels of monounsaturated (MUFA) and saturated fatty acids (SFA) in tissues of animals fed CLA isomer-enriched diets due to a decrease in the  $\Delta 9$ -desaturation of FA such as C16:0 or C18:0 (Alasnier et al., 2002). Recent studies (Alasnier et al., 2002; Belury, 2002; Kang et al., 2004; Korniluk et al., 2006; Czauderna et al., 2007b) have demonstrated that CLA isomers, particularly *t10c12* CLA, caused a reduction in  $\Delta 9$ -desaturase capacity, inhibited stearoyl-CoA desaturase mRNA expression and fatty acid synthesis (e.g. compare  $\Sigma$ FAC concentrations in muscles, Table 3). Similarly, in the present study, the value of the  $\Delta 9$ -desaturase index in the liver and muscles of rats fed a diet enriched with CLAmix and/or SeY decreased compared with that in the liver and muscles of control rats (Table 3). The experimental diet enriched only with SeY also decreased the  $\Delta 9$ -desaturase capacity, and thus the concentration of *c9*C18:1 in both tissues (Table 3), while the concentration of C18:0 in the liver and muscles increased (Table 4). Similarly, the addition of inorganic Se as selenate to diets decreased also the concentration of *c9*C18:1 in the liver, femoral muscles and spleen of rats (Czauderna et al., 2004a,b; Niedźwiedzka et al., 2006).

The diet enriched with CLAmix resulted in increased values of the atherogenic index ( $A_{\text{index}}$ ) in the liver and muscles of rats, likewise, the addition of CLAmix to the diet with SeY also caused an increase in the values of the  $A_{\text{index}}$  and thrombogenic index ( $T_{\text{index}}$ ) in muscles. Considering the above results, it seems reasonable to suggest that

dietary CLAmix can be responsible for accelerating atherogenesis and increasing platelet aggregation (Ulbricht and Southgate, 1991). Fortunately, the diets enriched with 1.5% CLAmix, irrespective of the presence of SeY, increased the concentrations of  $\alpha$ LNA, C22:6n-3, the sum of polyunsaturated fatty acids n-3 (PUFAn-3) and the ratio PUFAn-6 to PUFAn-3 (PUFAn-6/PUFAn-3) in the liver. On the other hand, the diets enriched with CLAmix with or without SeY decreased the accumulation concentrations of  $\alpha$ LNA, C22:6n-3 and the value of PUFAn-6/PUFAn-3 in muscles. So, the present results are in agreement with our previous studies (Korniluk et al., 2006) showing that a diet containing less than 2% of CLA mixture increased the concentration of  $\alpha$ LNA, C22:6n-3 and PUFAn-3 in the liver, whereas diets containing 1 or 2% of CLA mixture decreased the concentration of these fatty acids in femoral muscles of rats (Czauderna et al., 2007b). The possible mechanism through which the liver accumulation of long-chain PUFAn-3 (especially C22:6n-3) was increased by CLA isomers, particularly *t10c12*, is by changes in the capacity of  $\Delta 6$ -,  $\Delta 4$ -desaturations and elongation of these long-chain PUFAn-3 to very-long chain PUFAn-3 (Alasnier et al., 2002; Heird and Lapillonne, 2005). Moreover, these dietary CLA isomers inhibited enzymes involved in the synthesis of eicosanoids from long-chain PUFAn-3 (Belury, 2002; Heird and Lapillonne, 2005). Another reason may be that elongated and desaturated metabolites of CLA isomer(s) compete for the same enzymes as long-chain PUFAn-3. The above explanations are consistent with our results regarding the sum of concentrations of all assayed fatty acids ( $\Sigma$ FAC) in the liver of rats fed the diet containing CLAmix with or without SeY. Indeed, dietary CLAmix and/or SeY resulted in an increase in the content of  $\Sigma$ FAC in the liver (i.e. stimulated symptoms of fatty liver).

The diet containing CLAmix with or without SeY reduced the concentrations of assayed fatty acids in muscles; indeed, dietary CLA isomers reduced the adiposity of animals (Belury, 2002). The ability of CLA isomers to reduce adipose tissue was linked with the stimulation of adipocyte apoptosis and/or differentiation. Moreover, dietary CLA isomers elevated oxygen consumption concomitantly with increased oxidation of fat in animals (Belury, 2002). Considering the above results, we suggest that dietary CLAmix, irrespective of the presence of SeY, decreased

the nutritive value of meat of monogastric animals, as the value of the PUFA<sub>n</sub>-6-to-PUFA<sub>n</sub>-3 ratio increased in femoral muscles of rats fed the diet containing CLAmix with or without SeY. Fortunately, no noticeable harmful influence on the values of the PUFA/SFA, PUFA/ΣFA and UFA/ΣFA ratios in muscles of rats fed the experimental diets was found out.

The present study documented that the influence of diets containing CLAmix with or without SeY on the capacity of Δ4-, Δ5-, Δ6-desaturases and elongase was tissue-specific. In truth, the experimental diets usually elevated the capacity of these desaturases and decreased the yield of elongase only in muscles. On the other hand, the diet enriched with CLAmix elevated the elongase index (Elong<sub>index</sub>) in the liver with the result that the concentration of C22:6n-3 in this organ increased (Table 4).

## CONCLUSIONS

Feeding CLA isomers, irrespective of the addition of dietary SeY, increased the concentration of CLA isomers in the liver and muscles of rats. Our results documented that the addition of CLAmix to the diet with SeY stimulated the accumulation of Se in the liver and muscles. The finding that CLAmix and 0.5 μg Se/g diet as SeY fed to animals considerably decreased the content of the sum of all assayed fatty acids and most efficiently increased the concentration of Se in the muscles is valuable information for nutritionists carrying out research on farm animals to improve the nutritive value of food from the aspect of human health.

## REFERENCES

- Alasnier C., Berdeaux O., Chardigny J.M., Sébédio J.L. (2002): Fatty acid composition and conjugated linoleic acid content of different tissues in rats fed individual conjugated linoleic acid isomers given as triacylglycerols. *The Journal of Nutrition Biochemistry*, 13, 337–345.
- Belury M.A. (2002): Dietary conjugated linoleic acid in health: physiological effects and mechanisms of action. *Annual Review of Nutrition*, 22, 505–531.
- Crespo A.M., Reis M.A., Lanca M.J. (1995): Effect of selenium supplementation on polyunsaturated fatty acids in rats. *Biological Trace Element Research*, 47, 335–341.
- Czauderna M., Kowalczyk J., Wąsowska I., Niedźwiedzka K.M., Pastuszewska B. (2003): The effects of selenium and conjugated linoleic acid (CLA) isomers on fatty acid composition, CLA isomer content in tissues, and growth of rats. *Journal of Animal Feed Sciences*, 12, 865–881.
- Czauderna M., Kowalczyk J., Niedźwiedzka K.M., Wąsowska I., Pastuszewska B. (2004a): Conjugated linoleic acid (CLA) content and fatty acids composition of muscle in rats fed isomers of CLA and selenium. *Journal of Animal Feed Sciences*, 13, 183–196.
- Czauderna M., Kowalczyk J., Niedźwiedzka K.M., Wąsowska I., Pastuszewska B., Bulska E., Ruszczyńska A. (2004b): Liver and body mass gain, content of conjugated linoleic acid (CLA) isomers and other fatty acids in the liver of rats fed CLA isomers and selenium. *Journal of Animal Feed Science*, 13, 353–369.
- Czauderna M., Kowalczyk J., Korniluk K., Wąsowska I. (2005): Improving the analysis of fatty acids using combination of gas chromatography and Ag<sup>+</sup>-liquid chromatography. *Journal of Animal Feed Sciences*, 14, Suppl. 1, 563–566.
- Czauderna M., Kowalczyk J., Korniluk K., Wąsowska I. (2007a): Improved saponification then mild base and acid-catalyzed methylation is a useful method for quantifying fatty acids, with special emphasis on conjugated dienes. *Acta Chromatographica*, 18, 59–71.
- Czauderna M., Kowalczyk J., Korniluk K. (2007b): Effect of dietary conjugated linoleic acid mixture and selenized yeast on concentrations of selected fatty acids and mineral elements in rats. *Archives of Animal Nutrition*, 61, 135–150.
- Heird W.C., Lapillonne A. (2005): The role of essential fatty acids in development. *Annual Review of Nutrition*, 25, 1–23.
- Hur S.J., Park G.B., Joo S.T. (2007): Biological activities of conjugated linoleic acid (CLA) and effects of CLA on animal products. *Livestock Science*, 110, 221–229.
- Juniper D.T., Phipps R.H., Ramos-Morales E., Bertin G. (2008): Effect of dietary supplementation with selenium-enriched yeast or sodium selenite on selenium tissue distribution and meat quality in beef cattle. *Journal of Animal Science*, 86, 3100–3109.
- Kang K., Miyazaki M., James M., Ntambi J.M., Pariza M.W. (2004): Evidence that the anti-obesity effect of conjugated linoleic acid is independent of effects on stearoyl-CoA desaturase1 expression and enzyme activity. *Biochemical Biophysical Research Communications*, 315, 532–537.
- Korniluk K., Czauderna M., Kowalczyk J., Mieczkowska A., Taciak M., Leng L. (2006): Influence of dietary conjugated linoleic acid isomers and selenium on growth,

- feed efficiency, and liver fatty acid profile in rats. *Journal of Animal Feed Sciences*, 15, 131–146.
- McDowell L.R., Davis P.A., Cristaldi L.A., Wilkinson N.S., Buergelt C.D., Van Alstyne R. (2005): Toxicity of selenium: fear or precaution? *Feedstuffs*, 30, 12–13.
- Navarro-Alarcon M., Cabrera-Vique C. (2008): Selenium in food and the human body: A review. *Science of the Total Environment*, 400, 115–141.
- Niedźwiedzka K.M., Wąsowska I., Czauderna M., Kowalczyk J., Pastuszewska B. (2006). Influence of dietary conjugated linoleic acid isomers and Se on fatty acids profile in blood plasma and some tissues of rats. *Journal of Animal Feed Sciences*, 5, 471–489.
- Pastuszewska B., Ochtabińska A., Morawski A. (2000): A note on the nutritional adequacy of stock diets for laboratory rats and mice. *Journal of Animal Feed Sciences*, 9, 533–542.
- Phipps R.H., Grandison A.S., Jones A.K., Juniper D.T., Ramos-Morales E., Bertin G. (2008): Selenium supplementation of lactating dairy cows: effects on milk production and total selenium content and speciation in blood, milk and cheese. *Animal*, 2, 1610–1618.
- Rayman M.P. (2004): The use of high-selenium yeast to raise selenium status: How does it measure up? *British Journal of Nutrition*, 92, 557–573.
- Rodriguez E.M., Sanz M.T., Romero C.D. (1994): Critical study of fluorometric determination of selenium in urine. *Talanta*, 41, 2025–2031.
- Schomburga L., Schweizera U., Köhrle J. (2004): Selenium and selenoproteins in mammals: extraordinary, essential, enigmatic. *Cellular and Molecular Life Sciences*, 61, 1988–1995.
- Statistica by StatSoft (2002): Available at [www.statsoft.pl](http://www.statsoft.pl)
- Suzuki K.T. (2005): Metabolomics of selenium: Se metabolites based on speciation studies. *Journal of Health Science*, 51, 107–14.
- Weiss W.P., Hogan J.S. (2005): Effect of selenium source on selenium status, neutrophil function, and response to intramammary endotoxin challenge of dairy cows. *Journal of Dairy Science*, 88, 4366–4374.
- Ulbricht T.L.V., Southgate D.A.T. (1991): Coronary heart disease: seven dietary factors. *Lancet*, 338, 985–992.

Received: 2010–01–21

Accepted after corrections: 2010–05–20

---

#### Corresponding Author

Prof. dr. hab. Marian Czauderna, The Kielanowski Institute of Animal Physiology and Nutrition,  
Polish Academy of Sciences, Jabłonna, Poland  
Tel. +48 227 824 422, fax +48 227 742 038, e-mail: [m.czauderna@ifzz.pan.pl](mailto:m.czauderna@ifzz.pan.pl)

---

## INSTRUCTIONS TO AUTHORS

## Czech Journal of Animal Science

The journal publishes original scientific papers and board-invited review articles in English. The author is fully responsible for the originality of the paper and its subject and formal correctness. The author's declaration that the paper has not been published anywhere else should be enclosed. The Board of Editors decides on the publication of papers, taking into account peer reviews, scientific importance, and manuscript quality. Good laboratory practice and ethical rules must be followed. The SI international system of measurement units should be used. Manuscripts must be grammatically and linguistically correct and authors whose native language is not English are advised to seek the help of native English-speaker. Manuscript containing language errors are prejudiced in the reviewing process and may be returned to the author for rewriting before peer review and/or before acceptance.

Only manuscripts assessed by leading experts in the field will be published. If such reviews are not available within four months after registration of the manuscript, the peer review process is ceased, and the authors are notified. They can resubmit the manuscript, after its thorough revision and/or update, either to Czech Journal of Animal Science for a new assessment or to another journal. This should eliminate a long waiting period, and likely rejection of the manuscript.

If a revision of manuscript following the recommendation of the reviewers is requested, the modified manuscript must be re-submitted within six weeks. The authors can, however request an extension of the re-submission deadline, if necessary. All parts of the manuscript, including tables and figures (even unchanged) must be re-submitted. A detailed reply by the authors to every point of reviewer's recommendations must be attached to the revision manuscript (cover letter). It is not necessary to accept all requests of the reviewers, but a clear explanation of why reviewers' comments were not accepted, has to be provided. If the deadline for re-submission is missed, the paper will be removed from the reviewing process.

The proof reading must be returned within three days. Only errors originating during preparation of the document for printing can be corrected. Standard proof marks will be used. No changes in the manuscript after acceptance for publication can be allowed.

Czech Journal of Animal Science **do not accept hard-copy paper manuscripts**; all manuscripts must be submitted electronically to the journal website. This method of submission results in much faster handling of your manuscript, fewer handling errors, and allows you to track the handling progress of your manuscript at any time. Manuscripts must be submitted as a Microsoft Word or other word processing document (filetype ".doc" or ".rtf").

**Copyright.** The journal is protected by copyright held by the publisher after the manuscript has been accepted for publication. As concerns the transfer of rights, the corresponding author takes over responsibility for all authors. No part of this publication may be reproduced, stored, or transmitted in any form or by any means, without the written permission of the publisher.

**Manuscript layout.** The Microsoft (MS) Word for Windows word-processing software should be used for creating the text in non-formatted style strictly following the journal layout. The manuscript must be typed (Times New Roman, font 12) with double spacing throughout. Lines must be numbered in a consecutive manner starting on the first page, in the left-hand margin. All pages of the manuscript must also be numbered consecutively.

If any abbreviations are used in the paper, they have to be explained appropriately when they are used in the text for the first time. It is not advisable to use any abbreviation in the paper title or in the abstract. Tables, graphs and other MS Word documents are to be submitted on separate pages appended to the article. The document must not be formatted in columns, heading styles, etc. This unique MS Word file must be saved under the first author's surname only. Graphs should be provided in MS Excel and they should be stored with original data. Photographs and autotypes should be submitted in high resolution (min. 300 dpi) JPG or TIFF format. All tables, graphs and photos should be numbered in the order in which they are included in the text, using Arabic numerals.

**Manuscript length.** Originals papers, including figures, tables and references, should not exceed 25 typed or computer-written pages. The number of figures and tables should be kept to a minimum.

**The Title of the Paper** should be short and informative, no subtitles or numbering the "serial" articles (Part I, Part II etc.) should be used.

**The Abstract** should not have more than 2,500 keystrokes (characters plus spaces). It should contain important information on methods used to solve the problem, clear description of results and their statistical significance, and brief and unambiguous conclusions drawn from the results. References and discussion of results should not be included in the abstract.

**Keywords** should not repeat nouns used in the title and should describe the studied problem as best as possible.

**The Introduction** section should provide information on the present state of research in the field concerned. It briefly justifies the research, specifies the hypotheses to be tested and gives the objective (s). References to literary sources document such

present findings that are used by the authors, not all that have been published until now. References in the text should agree with those in the list of references. It is recommended to include references to papers from peer periodicals only. Citations from non-available sources (reports, national journals, proceedings etc.) should be omitted. Papers published by one or two authors are to be cited by their names, those published by three or more authors by the name of the first one et al. If more than one paper by the same author(s) published in the same year and are cited, they should be differentiated by YEARa,b,c both in the text and the list of References. Names and year of publication are to be cited by including them in the text directly, e.g. "... as published by Brown (1995)" or indirectly – citing authors and year of publication in parenthesis (Green and Grey, 1996), (Jakl et al., 2002). Several papers cited together should be arranged according to the year of publication starting with the oldest one.

**Material and Methods.** All preliminary material, conducted experiments, their extent, conditions and course (experimental design) should be described in detail in this section. All original procedures that were used for the processing of experimental material and all analytical methods used for evaluation should also be detailed. Data verifying the quality of acquired data should be indicated for the used methods. The whole methodology is to be described only if it is an original one, in other cases it is sufficient to cite the author of the method and to mention any particular differences. Methods of statistical processing including the software used should also be listed in this section.

**Results and Discussion.** The results obtained from the experiments including their statistical evaluation and any commentary should be presented graphically or in tables in this section. The author should confront partial results with data published by other authors, whose names and year of publication are to be cited by including them in the text directly or indirectly.

**References** should be arranged in alphabetical order according to the surname and initials of authors. The year of publication cited in parenthesis, the full title of the paper in English with the language of publication in parenthesis, e.g. (in Czech) should follow. The title of the periodical should be typed in full.

Only papers cited in the text should be included in the list of references. All names of the authors must be printed in English transcription without national letters. Authors are responsible for the accuracy of their references.

#### **Examples of references in the list:**

- **Journal article**

Brown J. (1995): Estradiol determination in post-partum sows. *Journal of Endocrinology*, 198, 155–169.

- **Electronic journal article:**

Steinkraus K.H. (2002). Fermentation in world food processing. *Comprehensive Reviews in Food Science and Food Safety* [serial online]. 1, 23–32. Available from [www.ift.org](http://www.ift.org) (accessed Apr 1, 2002).

- **Books and article within edited books, proceedings**

Spally M.R., Morgan S.S. (1989): *Methods of food analysis*. 2<sup>nd</sup> Ed. Elsevier, New York, 682 pp.

Kalab J. (1995): Changes in milk production during the sexual cycle. Hekel K. (ed.): *Lactation in Cattle*. Academic Press, London, UK, 876–888.

Janson L., Ahlin K.A. (1992): Postpartum reproductive performance in cattle selected for high and low fat content. In: *Proc. 43<sup>rd</sup> Annu. Mtg. European Association for Animal Production (EAAP)*. Madrid, Spain, 93–95.

- **Patent:**

Harred J.F., Knight A.R., McIntyre J.S., inventors; Dow Chemical Co., assignee. 1972 Apr 4. Epoxidation process. U.S. patent 3, 654, 317.

- **Dissertation:**

Smith D.E. (1988): Lipid oxidation at very low water activities. PhD Diss. Ithaca, NY: Cornell University. Available from: University Microfilms, Ann Arbor, MI: ABD62-83. 210 p.

**Corresponding Author** should include his or her full name including all academic, scientific and pedagogic titles and detailed address of the institution with postal code, phone and fax numbers, e-mail address. The author who is responsible for any correspondence with the journal should be indicated clearly.

**Declaration of the authors** must be carefully completed and signed by the first author, and then scanned in .pdf format to be uploaded in the submission of your manuscript, along side with the other manuscript files.

**Offprints:** Free reprint in Portable Document Format (PDF) sent via e-mail as an attachment.

**Compliance with these instructions is obligatory for all authors. If a manuscript does not comply exactly with the above requirements, the editorial office will not accept it for a consideration and will return it to the authors without reviewing.**

Revised February 2010