

Sample drying to improve HCHO measurements by PTR-MS instruments: laboratory and field measurements

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Received: 21 August 2009 – Published in Atmos. Chem. Phys. Discuss.: 23 September 2009

Revised: 31 December 2009 – Accepted: 13 January 2010 – Published: 17 February 2010

Abstract. A significant improvement in the PTR-MS instrument sensitivity to formaldehyde was obtained by drying the air sample to a dew point of -30°C using a cold trap to condense and freeze water vapour. At warmer trap temperatures there was significant uptake of formaldehyde and other water soluble organics, suggesting the presence of a quasi-liquid layer on the ice surface. By removing water vapour to a low constant dew point, the PTR-MS can be operated at low E/N ratios, significantly increasing normalized sensitivities for all organics and removing their humidity dependence due to reactions with $\text{H}^+(\text{H}_2\text{O})_2$. At an E/N ratio of 80 Td, the formaldehyde normalized sensitivity was 25 Hz/ppbv per MHz H_3O^+ with an estimated detection limit of 78 pptv. Field testing demonstrated good agreement between HCHO measurements made at ambient humidity and corrected for water vapour effects compared to dehumidified sampling at -30°C . Field testing also revealed that at an E/N ratio of 100 Td or lower there was a significant ion signal at $m/z=49$, likely CH_3OOH . Sampling drying and operation at low E/N ratios enables sensitive measurements of HCHO and potentially CH_3OOH , both important tropospheric photoproducts.

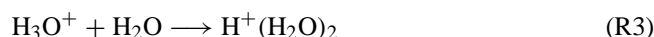
1 Introduction

The proton transfer reaction mass spectrometer (PTR-MS) is a commercial instrument sold by Ionicon Analytik (Austria). Since its initial description (Hansel et al., 1995; Lindinger et al., 1998), it has found wide use in atmospheric chemistry field and laboratory studies for measuring trace organic

gases. A description of its operating principle and performance can be found in the recent review article of deGouw and Warneke (2007). One advantage of this instrument over gas-chromatography based determinations of volatile organic compounds (VOCs) is the time resolution of the measurements and limited sample handling which makes determination of oxygenated species easier. However, formaldehyde has been proven difficult to measure with the PTR-MS. The sensitivity to formaldehyde is much less than other organics, due to a water vapour interference. The proton affinity of formaldehyde (713 kJ/mol) is only slightly greater than that of water (697 kJ/mol), so the exothermicity of the protonation Reaction (R1) is small. Within the drift tube, where the protonation reaction takes place, the kinetic energies of the reacting reagent ions are large enough that the endothermicity of the back Reaction (R2) can be overcome, and the rate of the back reaction can be significant (Hansel et al., 1997).

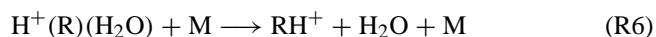
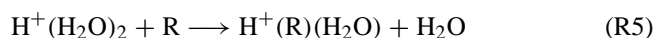
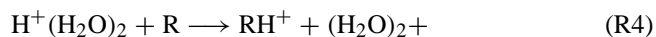


The rate of the back reaction is dependent on the water vapor concentration, the electric field E (V/cm), and the number density N (molecules/cm³) of the PTR-MS drift tube (Hansel et al., 1997). High drift pressures and low electric fields generally provide the best overall conditions to improve PTR-MS sensitivity. For HCHO measurements, the adverse consequence of operating the PTR-MS at higher drift pressures is the corresponding increase in water vapour concentration in the drift tube which increases the rate of the back reaction. Operating at a lower electric field strength can reduce the back reaction rate constant but low ratios of E/N cause excessive formation of water cluster ions (Reaction R3)



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The formation of water cluster ions complicates the determination of PTR-MS instrument sensitivities to VOCs because water clusters can also react with organics to produce protonated product ions. This can occur in 2 ways, by direct protonation of the organic by the water cluster ion (Reaction R4), or through ligand switching reactions (Reaction R5) and subsequent collision induced dissociation of the organic water cluster (Reaction R6)



Species with proton affinities (PA) greater than the 808 ± 6 kJ/mol proton affinity of the water dimer (Goebert and Wenthold, 2004; Midey et al., 2002) can be directly protonated. For species of atmospheric importance, these include isoprene (PA=826 kJ/mol), methyl vinyl ketone (835 kJ/mol) styrene (840 kJ/mol), 1,3,5-trimethylbenzene (836 kJ/mol), and acetone (812 kJ/mol) where the listed proton affinities were taken from Hunter and Lias (1998). Monoterpenes also appear to have PA's greater than the water dimer. The PA of α -pinene and camphene was calculated to be 879 kJ/mol by Solouki and Szulejko (2007), and the PA of limonene was measured to be 875 ± 5 kJ/mol by Fernandez et al. (1998). The ligand switching reactions are probably a more general mechanism for causing a water vapour dependent sensitivity (de Gouw and Warneke, 2007). Switching reactions are thought to be fast if exothermic and possibly need the organic molecule to have a significant dipole moment (Spanel and Smith, 1995). Midey et al. (2000) have measured the rate constants for formaldehyde and acetaldehyde reacting with H_3O^+ and $\text{H}^+(\text{H}_2\text{O})_2$ using a SIFT. They observed that both species reacted with $\text{H}^+(\text{H}_2\text{O})_2$ in ligand switching reactions, with the $\text{HCHO} + \text{H}^+(\text{H}_2\text{O})_2$ reaction rate being 75% of the calculated collision rate. This suggests that ligand switching reactions between HCHO and $\text{H}^+(\text{H}_2\text{O})_2$ could also occur in the drift tube of the PTR-MS.

Despite these complications, there are several reports of PTR-MS based measurements of HCHO which try to account for the humidity dependence of the sensitivity (Holzinger et al., 1999; Karl et al., 2003, 2007; Steinbacher et al., 2004; Inomata et al., 2008; Seco et al., 2008; Taipale et al., 2008). In this paper, we will show that the PTR-MS sensitivity to HCHO can be dramatically improved by removing water vapour from the sample stream. This has the advantages of decreasing the rate of the back reaction and keeping the water vapour concentration in the drift tube constant so that HCHO and other VOC sensitivities are independent of ambient water vapour concentration. Sample drying also has the added advantage of allowing the drift tube to be operated at much lower E/N ratios. Reducing the ratio increases reaction times, further increasing sensitivities. The reduction in the E/N ratio also reduces the degree of fragmentation from

dissociative protonation reactions and may allow for the determination of CH_3OOH .

2 Experimental

2.1 Impact of water vapour on HCHO normalized sensitivity

Following Warneke et al. (2001), the normalized sensitivity is defined here as the RH^+ count rate per ppbv per MHz H_3O^+ count rates (1 ppbv=1 nmol mol⁻¹). This definition of sensitivity follows from the ion-molecule reaction kinetics in the drift tube where the RH^+ count rate is given as

$$\text{RH}^+ = k t [R] \text{H}_3\text{O}^+ \varepsilon \quad (1)$$

with k being the ion molecule rate coefficient, t the reaction time, $[R]$ is the number density of the organic species in the drift tube, RH^+ and H_3O^+ are the ion count rates (Hz), and ε is the ion transmission efficiency of RH^+ relative to H_3O^+ . The reaction time is determined by the H_3O^+ drift velocity and length of drift tube, so Eq. (1) can be re-written to account for the drift tube parameters on the reaction time:

$$\text{RH}^+ = k \left(\frac{LN}{\mu_o N_o E} \right) [R] \text{H}_3\text{O}^+ \varepsilon \quad (2)$$

where L is the drift length, N is the number density of air in the drift tube, N_o is the number density at 273 K and 1 atmosphere, μ_o is the reduced mobility of H_3O^+ (2.76 cm² V⁻¹ s⁻¹), and E is the electric field. Assuming $\varepsilon=1$, and a calculated thermal reaction rate coefficient of 2.0×10^{-9} cm³ molecule⁻¹ s⁻¹ (Su and Chesnavich, 1982; Su, 1988), a normalized sensitivity of 9.6 Hz/ppbv can be calculated for benzene using the typical experimental conditions described in this paper: 120 Td drift field intensity, 2.1 mbar pressure, 50 °C drift temperature, 9.2 cm drift tube length. To increase the inherent sensitivity of the PTR-MS there are three strategies employed: increase the H_3O^+ count rate, increase the concentration of the analyte by increasing the drift pressure, or increase the reaction time by decreasing the E/N ratio. The ratio E/N is given the unit of Townsends (1 Td=1 × 10⁻¹⁷ V/cm²).

Figure 1 shows the impact of sample humidity on the reagent ion and first water cluster count rates, and the influence on normalized sensitivity for HCHO, acetaldehyde and benzene for drift field intensities of 120 and 108 Td. The normalized sensitivities were determined from a 14 component VOC standard (Apel-Reimer Environmental) diluted to 19.8 ppbv with humidified zero air. The diluent air was humidified to a range of water vapour concentrations by blending a dry zero air flow with a flow of humid zero air. Humid air was generated by passing zero air over purified water (de-ionized and filtered for organics). Air flows were metered with mass flow controllers and the resulting humidity of the

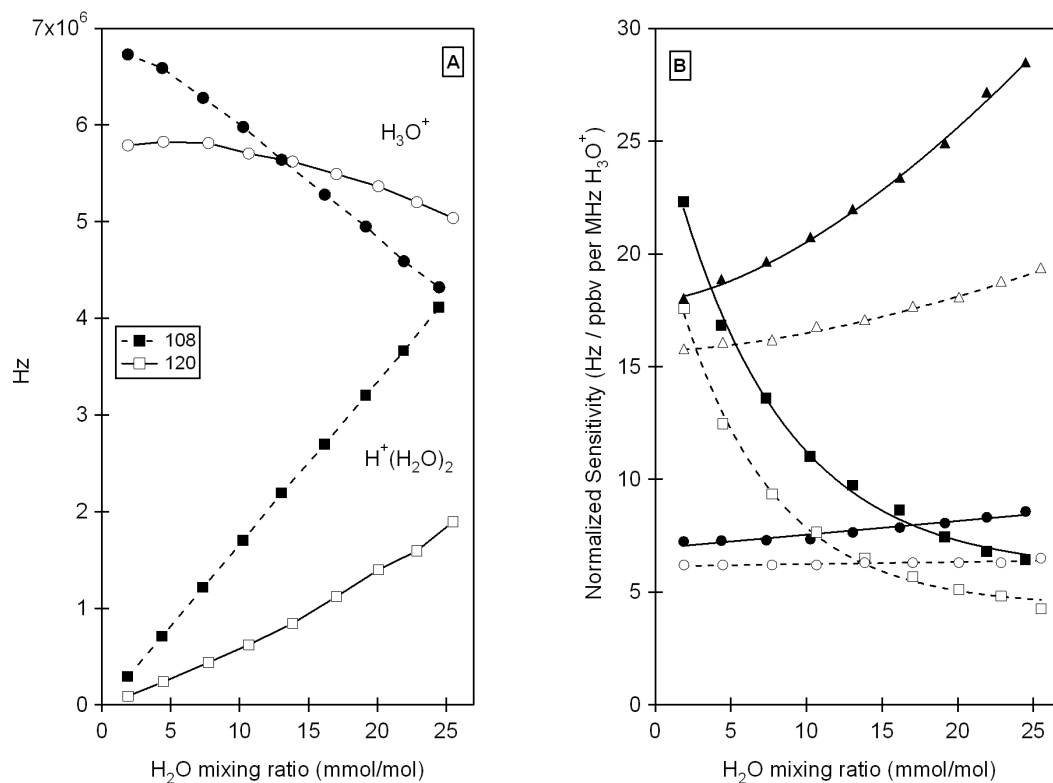


Fig. 1. Humidity dependence of ion count rates and normalized sensitivity for 120 and 108 Td drift conditions at 2.1 mbar pressure. The Td=120 data are given by open symbols, Td=108 data by filled symbols. Panel (A) count rates for H₃O⁺ (circles) and H⁺(H₂O)₂ (squares). Panel (B) shows corresponding normalized sensitivity and fits to data for benzene (circles with linear fit), acetaldehyde (triangles with power law fit), and formaldehyde (squares with exponential fit).

diluent air measured by a relative humidity sensor (Humi-Cap, Vaisala). HCHO was added to this stream of air from a high emitting HCHO permeation source (KinTek, USA).

Figure 1 clearly shows the problem with HCHO measurements by PTR-MS. HCHO normalized sensitivities decreased exponentially with water vapour mixing ratios, decreasing by about a factor of 5 from dry conditions at 1.9 mmol mol⁻¹ (0.19%) water vapour to humid conditions of 25 mmol mol⁻¹ (2.5%) water vapour for the 120 Td drift condition. A greater decrease was observed for the 108 Td drift condition. In contrast to HCHO, acetaldehyde normalized sensitivities increased with water vapour mixing ratios. If acetaldehyde was only protonated by reaction with H₃O⁺ the normalized sensitivity would not change. The increase implies that acetaldehyde was reacting with protonated water clusters such as H⁺(H₂O)₂. At the highest water vapour mixing ratios for the 108 Td drift condition, the measured H⁺(H₂O)₂ signal was nearly equal to that of H₃O⁺. Since the proton affinity of acetaldehyde is lower than that of the water dimer, the likely mechanism is ligand switching (Midey et al., 2000) and subsequent dissociation to produce protonated acetaldehyde. The normalized sensitivities of other polar organics tested but not shown in the figure,

such as acetonitrile, acetone, 2-butanone and methacrolein, also displayed significant water vapour dependence. Benzene, on the other hand, displayed only a very weak increase in normalized sensitivity, increasing by only 4.4% at Td=120 and 15.5% at Td=108. Benzene does not have a permanent dipole moment and its ligand switching reaction rate has been measured to be very slow if it occurs at all (Spanel and Smith, 1995). The increase in benzene normalized sensitivity may be due to changes in the reaction time due to a decrease in H₃O⁺ mobility with higher water vapour concentration in the drift. At a lower *E/N* ratio (\sim Td<100), Warneke et al. (2001) observed lower H₃O⁺ mobility with increasing humidity, but at higher *E/N* ratios the humidity influence was not clear. Another explanation for our observed increase in benzene normalized sensitivity is the potential for loss of H₃O⁺ to clustering in the vacuum interface after the drift (Hanson et al., 2009). The observed benzene normalized sensitivity of 6.2 Hz/ppbv at 120 Td for dry conditions was 35% less than the sensitivity calculated from Eq. (2). At 108 Td the difference was 31%. In these experiments and in the field test described later, we used a ceramic based electron multiplier detector (ETP/SGE Analytical).

Table 1. Percent difference in normalized sensitivities between dry air (2 mmol mol⁻¹ water vapour) and humid air (>24 mmol mol⁻¹ water vapour).

<i>m/z</i>	Species	Proton affinity (kJ/mol) ^a	Permanent dipole moment (Debye) ^b	% Difference in normalized sensitivity ^c	
				Td=120	Td=108
31	formaldehyde	712.9	2.18	-75.6	-246
33	methanol	754.3	1.66	17.4	25.5
42	acetonitrile	779.2	3.83	18.7	36.1
45	acetaldehyde	786.5	2.65	23.6	36.8
59	acetone	812	2.29	25.1	38.6
69	isoprene	826.4	0.250	24.5	36.5
71	methacrolein	808.7	2.80	22.8	37.9
73	2-butanone	827.3	2.76	30.2	45.4
79	benzene	750.4	0.00	4.4	15.5
93	toluene	784.0	0.343	11.2	28.9
105	styrene	839.5	0.186	17.0	35.1
107	p-xylene	794.4	0.081	19.3	35.3
121	1,2,4-trimethylbenzene		0.291	18.3	36.0
135	1,2,3,5-tetramethylbenzene			14.9	37.3
137	α -pinene	879 ^d	0.149	20.3	41.0

^a Hunter and Lias, 1998.^b Zhao and Zang, 2004.^c normalized sensitivity defined as Hz/ppbv per MHz H₃O⁺.^d Solouki and Szulejko, 2007.

Other aromatics examined, toluene, p-xylene, 1,2,4-trimethylbenzene, styrene, and 1,2,3,5-tetramethylbenzene displayed a slightly stronger dependency on humidity than benzene. Midey et al. (2002) have observed proton transfer reactions between H⁺(H₂O)₂ and alkylbenzenes such as toluene in SIFT experiments. These reactions were fast, about half the collision rate, and were attributed to ligand switching reactions. It seems likely that toluene and the alkylbenzenes in our calibration mixture also undergo proton transfer reactions via ligand switching in the PTR-MS, resulting in the observed humidity dependent normalized sensitivities. Based on proton affinities, styrene could also be directly protonated by H⁺(H₂O)₂. Table 1 lists the percentage difference in VOC normalized sensitivity between the driest condition (1.9 mmol mol⁻¹) and the most humid condition used in these experiments (24.5 mmol mol⁻¹ at Td=108 and 25.5 at Td=120) together with corresponding proton affinities and permanent dipole moments. In all cases, except for formaldehyde, there was a significant increase in normalized sensitivity for the humid conditions.

The humidity dependent sensitivities for benzene and toluene have been examined in some detail by Warneke et al. (2001). In that study, conducted at 2.5 mbar drift pressure and a drift field intensity of ~108 Td, the normalized sensitivity for benzene and toluene decreased nearly by a factor of 2 when the water vapour was decreased from

5 mmol mol⁻¹ to 27 mmol mol⁻¹. Their results are in direct contrast to the results shown here. Warneke et al. (2001) explained their decrease in sensitivity to be a consequence of dissociation of H⁺(H₂O)₂ in the vacuum interface between the drift tube and quadrupole causing higher measured H₃O⁺ than actually present in the drift tube. The higher drift pressure in that study resulted in a significantly greater relative abundance of H⁺(H₂O)₂ compared to H₃O⁺. Steinbacher et al. (2004) also reported that benzene normalized sensitivity decreased by 16% over a similar humidity range. This suggests to us that the humidity dependence of benzene normalized sensitivity could be used as an instrument performance test to determine the fidelity of the ion sampling from the drift by the mass spectrometer interface. If benzene does not react with H⁺(H₂O)₂ in the PTR-MS, any change in its normalized sensitivity would indicate ion sampling artifacts. The caveat is the potential influence of water vapour on H₃O⁺ mobility. The expected normalized benzene sensitivity can also be readily calculated from the drift operating parameters, since benzene is only reacting with H₃O⁺. The normalized sensitivity for benzene and its humidity dependence is probably the best metric for comparing different PTR-MS instruments and new designs sold by Ionicon Analytik. We note that different versions of the normalized sensitivity metric are used in the PTR-MS literature to account for various instrument factors that influence ion count

rates per ppbv of analyte (de Gouw and Warneke, 2007; Taipale et al., 2008) but we prefer the simpler metric originally proposed by Warneke et al. (2001).

2.2 Sample drying

The technical approach to a water-removal system was based on cold trapping as is often done for VOC measurements by gas chromatography. The design of the dehumidifier was based on that used by Goldan et al. (1995). The dehumidifier assembly consisted of two 150 mm × 6.4 mm OD stainless steel tubes coated with amorphous silicon (Silonite coating, Restek Inc.) mounted inside a block of aluminum that was cooled by an immersion cooler probe (FTP Systems). The tubes were in loose thermal contact with the block, with an air gap of 1.5 mm between the tube wall and the block. The immersion cooler probe cooled the block to about -50°C , cooling the tubes as well. The temperature of the tubes were controlled by resistive heating using a polyamide coated nichrome wire with a resistance of ~ 95 ohms (California Fine Wire Corp.) that was coiled around the tubes. The temperature of the individual tubes was controlled by temperature controllers (Watlow, USA) and tube temperature was monitored using an ungrounded thermocouple probe in contact with the inner tube wall. The thermocouple was inserted through a septum mounted on a Silonite coated tee on the upstream side of the tube. The tubes could, therefore, be held at a constant temperature ranging from the lower limit of the immersion cooler ($\sim -50^{\circ}\text{C}$) to an upper limit of about 100°C . The tubes were thermally isolated from each other so that they could be operated independently; the temperature set-point of one did not impact the ability to control the temperature of the other. Water vapour was removed by pulling the sample air through a cold tube with the PTR-MS inlet flow. The water vapour condenses and freezes onto the tube surface, effectively lowering the dew point temperature of the sample air flow to that of the tube surface temperature.

The dehumidifier was constructed with two tubes to ensure continuous measurements over long time periods. Each tube of the dehumidifier was operated in one of three states: sample, condition and back flush. While one tube was removing water from the sample stream, the other tube was heated ($+70^{\circ}\text{C}$) and back flushed with dry nitrogen to remove ice accumulated in the tube during the last sample cycle. It was then cooled and conditioned with ambient air prior to sampling to ensure a seamless transition between tube measurement cycles. The conditioning state ensures that the VOC's have come to equilibrium with the ice-covered tube wall before being switched over to the sample state as described below. All flows through the tubes in the dehumidifier were directed by six 3-way PFA solenoid valves. The valves were arranged so that one tube can be in the sample mode, while the other tube can be operated in the back flush or conditioning state. While a tube is in the sample state, flow through the tube is determined by the PTR-MS inlet flow, set

to approximately 250 ml min^{-1} . Back flush and conditioning flows were regulated by two separate rotameters. Data acquisition and control of the dehumidifier apparatus was done using a LabJack U3-LV USB data acquisition device (LabJack, USA) and DaqFactory Standard software (Azeotech, USA). The LabJack U3-LV provided control voltages for all the valves and solenoids and inputs for digital and analog signals. DaqFactory provided the user interface and drivers to communicate with the LabJack U3-LV and temperature controllers. DaqFactory communicated with the temperature controllers through a serial 485 interface, allowing the high and low temperature set-points for each tube to be set by a software programme that automated functioning of the water trap.

The water trap efficiently removed water vapour from the sample at flow rates less than 300 ml min^{-1} . This could be monitored by the PTR-MS by measuring the abundance of the water cluster ion $\text{H}^+(\text{H}_2\text{O})_2$. At flows greater than 300 ml min^{-1} , the trap was less efficient as evidenced by an increase in $\text{H}^+(\text{H}_2\text{O})_2$ with air flow. The use of a cold tube to remove water by condensation could potentially also remove organics by physical adsorption to the cold surfaces. We conducted a series of experiments testing the transmission of HCHO and a suite of VOCs through the dehumidifier at different temperatures which are described below.

2.3 Field test

The water trap was field tested in the city of Houston, Texas as part of the Study of Houston Atmospheric Radical Precursor (SHARP) campaign from April–May 2009. The study was conducted on the roof of an 18-story building on the University of Houston campus. For the first two weeks of the campaign, the PTR-MS alternated between sampling through the water trap at -30°C and through PFA tubing at ambient room temperatures ($\sim 25^{\circ}\text{C}$). The sampling duration was 25 min in each state. This allowed verification of potential losses of less volatile aromatic VOCs to the water trap and to verify the impact of ambient humidity on PTR-MS calibrations. Drift field intensity was 120 Td with 2.1 mbar drift pressure. After this test period, the PTR-MS sampled continuously through the water trap at -30°C using drift field intensities of 120, 100 and 80 Td. The PTR-MS was calibrated and zeroed on an automated, regular schedule using a 14 VOC component compressed gas standard (Scott-Marrin, USA). The standard was diluted to 19.8 ppbv with humid zero air produced by scrubbing ambient air with a Pt catalyst (1% Pt on alumina spheres) at 260°C . HCHO was calibrated using a permeation tube with a stated permeation rate accuracy of 2%. The permeation tube was placed in a permeation oven heated to 80°C and the permeation flow (100 ml min^{-1} zero grade air) added to the diluted multicomponent standard flow to yield 43 ppbv HCHO. The permeation oven was not as well thermostated as we would have liked and observed that its temperature varied $\pm 2^{\circ}\text{C}$ as room

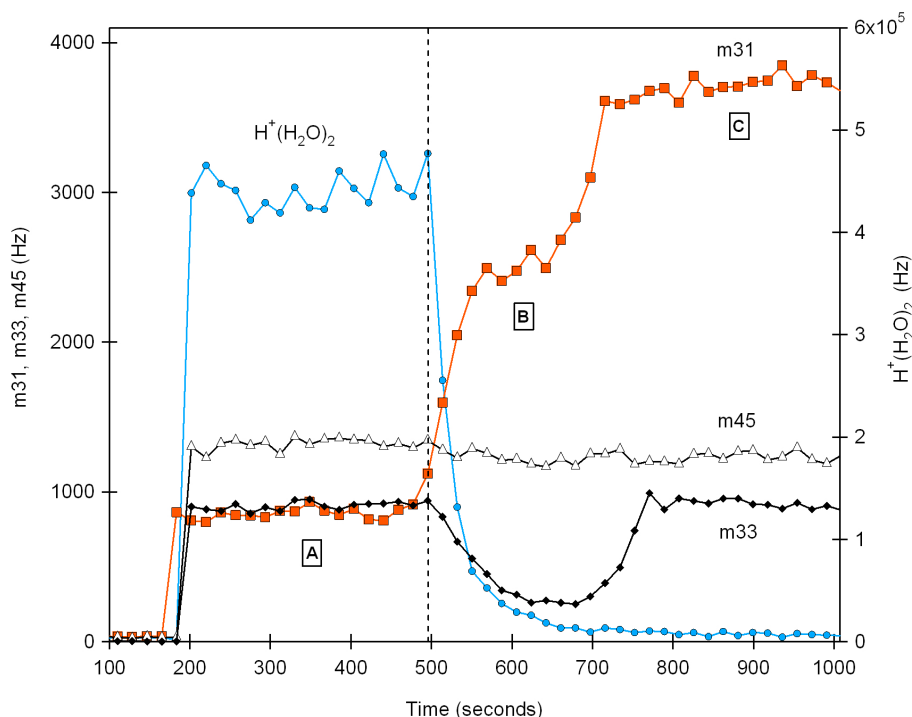


Fig. 2. Impact of water trap on formaldehyde (m31), methanol (m33), and acetaldehyde (m45) ion signals. PTR-MS switches from sampling at 50% RH at 23 °C (region A) at about the 500 s mark to sampling through the water trap at -40 °C as indicated by dashed line and abrupt decrease in $\text{H}^+(\text{H}_2\text{O})_2$ count rates. Formaldehyde initially increases to a level indicated by region B as trap accumulates ice and PTR-MS dries (conditioning period), then increases to a final value indicated by region C. By contrast, methanol decreases then recovers to original count rate.

temperature varied. The automated calibrations and zeroes were performed for both the dehumidified and ambient humidity sampling.

3 Results

3.1 Trap conditioning

Tests were performed to determine the optimum temperature and flow rates for operating the trap. The lower the water trap temperature, the more water vapour is removed improving sensitivity to HCHO. The concern with the trap was that HCHO would be lost to the ice accumulating in the water trap and that other VOCs, such as larger aromatics with lower vapour pressures, would also be removed to the cold ice surface.

Figure 2 illustrates the impact of the dehumidifier on a 19.8 ppbv calibration mixture at a relative humidity of 50% at 23 °C. There are three identifiable ion signal regions that we labelled A, B, and C. Region A is the ion signal at an ambient tube temperature (23 °C) corresponding to normal air sampling to serve as a reference point to compare with dehumidified sampling. In region A, water vapour in the sample contributes to a decrease in HCHO sensitivity and

a lower observed count rate. The sample was then redirected by solenoid valves through the water trap at a temperature of -40 °C. Region B was identified as a conditioning period as the tube becomes coated with ice and adsorbed VOCs. During this period, there was a significant decrease in the methanol ion signal as it is lost to the cold tube. In contrast, the HCHO ion signal increased, due to the reduction of water vapour in the drift tube. After ~ 7 min, the HCHO signal was reasonably stable at ~ 3800 Hz. At this point, the HCHO was in equilibrium with the cold ice surface, identified as region C. In region C, the methanol ion signal has recovered to values similar to region A. The sample drying is illustrated by the large change in $\text{H}^+(\text{H}_2\text{O})_2$ count rates, from $\sim 450,000$ Hz at room temperature to ~ 3800 Hz at -40 °C. Removing water vapour by condensing it in a cold tube clearly improved the PTR-MS sensitivity to HCHO. The HCHO ion count rate at -40 °C was 5 times greater than at 23 °C.

Figure 2 also highlights a clear need for a conditioning period in which the tube becomes passivated before efficient transmission through the water trap is possible. This practice of conditioning is part of the general lore of VOC analysis, though not well quantified nor illustrated in the literature. We conducted several tests using dry and humidified test mixtures to determine losses of VOCs to cold tubing.

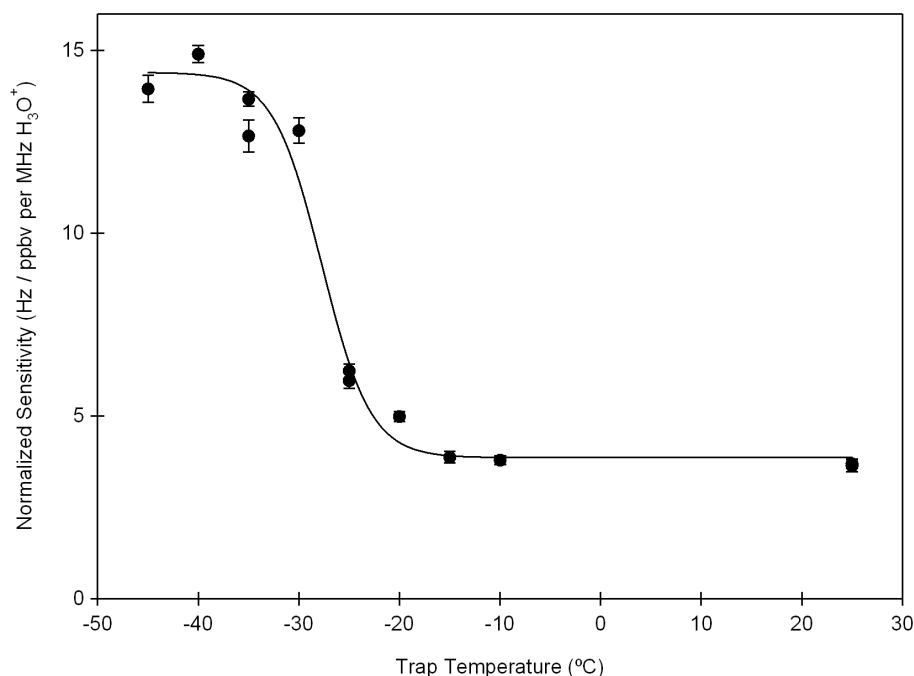


Fig. 3. Dependence of HCHO normalized sensitivity on water trap temperature for 50% RH sample flow at 23 °C. PTR-MS conditions were 120 Td, 2.4 mbar pressure, 50 °C drift temperature. Trace is a sigmoid fit through the data to illustrate the abrupt increase in sensitivity at water trap temperatures below -25 °C.

Even for relatively volatile species a conditioning effect was observed. For example, tests with our HCHO + multicomponent VOC test mixture in dry air showed that ~ 10 ng of HCHO adsorbed to the Silonite coated steel tubing at -65 °C before the tube was passivated and allowed 100% transmission of HCHO. Higher mass flows led to quicker conditioning times. Conditioning is a complex surface adsorption process dependent on the nature of the sample including the VOC composition and abundance and the amount of water vapour. For the water trap, the tubes become coated with ice so the interaction of the VOC with ice becomes an important process. For the less volatile non-polar aromatic compounds in our test mixture, 1,2,3,5-tetramethylbenzene and 1,2,4-trimethylbenzene, lower tube temperatures lead to greater losses as would be anticipated from surface losses due to physical adsorption. Interestingly, for water soluble species such as HCHO and methanol, colder trap temperatures lead to greater transmission efficiencies.

3.2 Loss of water soluble species to warm ice

We observed that the improvement in HCHO sensitivity was a nonlinear function of the trap temperature. Figure 3 shows a plot of HCHO normalized sensitivity versus water trap temperature for a series of tests conducted at 2.4 mbar drift pressure and Td=120 drift field intensity. We found a steep increase in HCHO sensitivity at temperatures lower than -25 °C. There appears to be a transition at this temperature.

We believe this transition is due to soluble VOCs being taken up into a liquid-like layer on the ice surface at temperatures greater than -25 °C (Dash et al., 1995). This transition is clearly seen in Fig. 4. The figure shows results from a test comparing the transmission of a 19.8 ppbv cal mixture through the water trap at -30 and -25 °C. The bottom panel shows the $\text{H}^+(\text{H}_2\text{O})_2$ ion count rates. Initially the PTR-MS samples dry zero air, then the calibration mixture at 50% RH through the tube at 23 °C, then the sample passes through the water trap. The cycle was then repeated for the other water trap temperature. Passage through the water trap is indicated by the drop in the water cluster count rates. It can be seen that there was no significant change in the aromatic count rates (benzene, 1,2,4-trimethylbenzene) as the sample was switched from a tube at $+25$ °C to -30 °C. There was, however, a dip in the methanol count rates which then recovered to those observed for the $+25$ °C condition. Similarly, there was a small dip in the acetonitrile count rates which then recovered. HCHO count rates initially increased (due to lower water vapour concentration), levelled out for ~ 300 s, and then increased to a plateau. A much different behaviour was observed for the trap temperature of -25 °C. While the aromatics passed through unaffected, methanol and acetonitrile were persistently lower, indicating continual loss to the trap; the trap never conditioned. HCHO count rates increased initially then slowly decreased. This behaviour is consistent with uptake into a liquid water

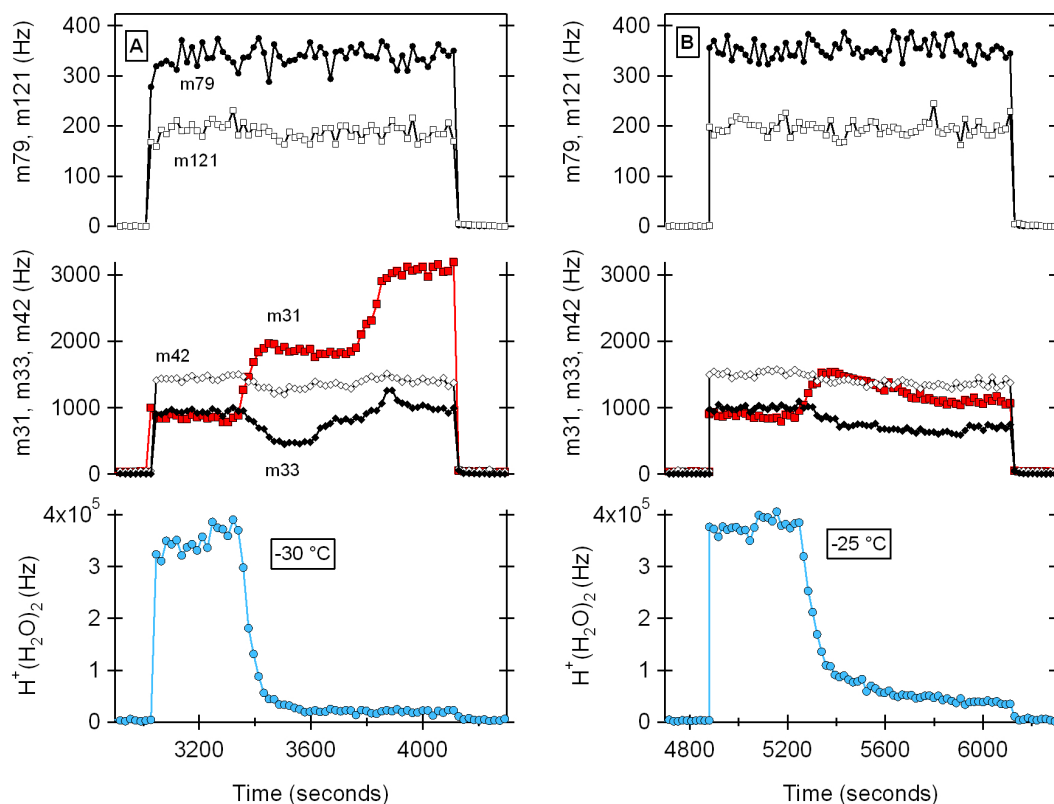


Fig. 4. Illustration of HCHO (m31), methanol (m33), and acetonitrile (m42) loss to warm ice. Panel (A) shows change in ion signal as PTR-MS switches from sampling at 50% RH at 23 °C to sampling through water trap at -30°C . An abrupt decrease in the $\text{H}^+(\text{H}_2\text{O})_2$ ion signal indicates when the PTR-MS samples through the water trap. Methanol displayed an initial decrease in ion signal then recovered to the ambient RH level. Panel (B) shows the change in ion signal for sampling through a -25°C water trap temperature. In this case, the methanol and acetonitrile count rates did not recover to those observed for ambient RH sampling. HCHO initially increased then decreased to a count rate slightly greater than ambient RH conditions. Benzene (m79) and 1,2,4-trimethylbenzene (m121) displayed no losses to water trap at either temperature.

surface that is constantly being replenished. Formaldehyde is very soluble with a Henry's law coefficient at -25°C of $3.1 \times 10^5 \text{ M atm}^{-1}$ (Staudinger and Roberts, 1996). Partitioning into the liquid water layer would qualitatively explain loss of formaldehyde from the sample flow. At a trap temperature of -25°C , less formaldehyde makes it through the tube than the ambient RH condition, despite the lower water vapour concentration in the drift tube and the inherently greater normalized sensitivity. This absorption effect was also observed for methanol but not for other gases in the test mixture such as acetaldehyde, acetone, and methacrolein. At -25°C these species have a Henry's Law coefficient of at least a factor of 9 less than that of methanol and partitioning into the liquid phase would not be as significant, consistent with our observations. Thus, two effects determine the transmission efficiency through the water trap: physical adsorption of VOC's to ice or the tubing surface and absorption of soluble VOC's into a quasi liquid layer of water on the ice surface. From these tests we concluded that the optimum trap temperature was -30°C . At trap temperatures colder

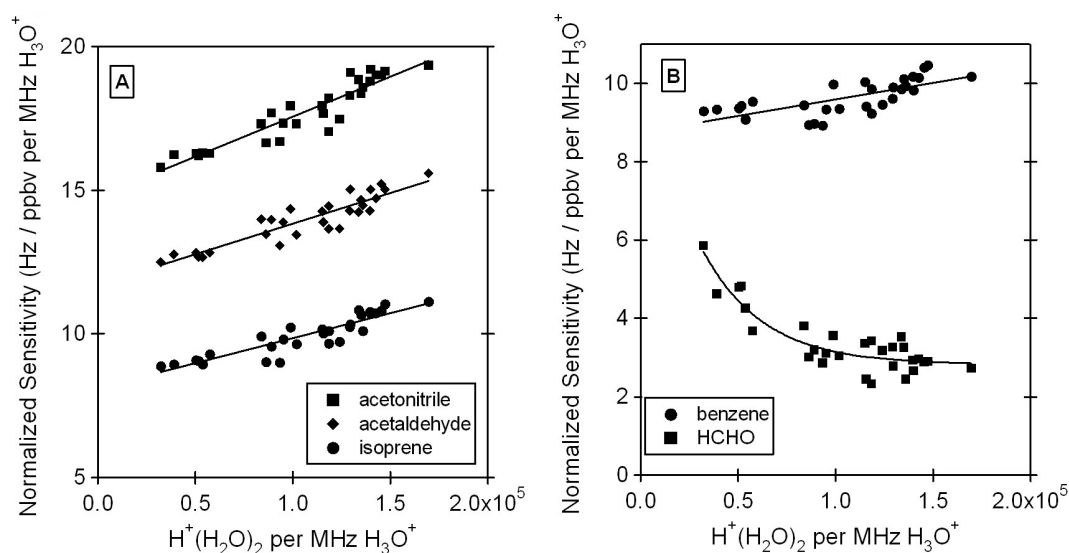
than this, we observed losses of 1,2,3,5-tetramethylbenzene. At trap temperatures warmer than this, there were significant losses of soluble species such as methanol. A conditioning flow rate of $200\text{--}300 \text{ ml min}^{-1}$ for 6 min was adequate for conditioning the trap for HCHO and other VOCs.

3.3 Field calibrations

Figure 5 shows the normalized sensitivities for HCHO, benzene, acetaldehyde, isoprene and acetonitrile as a function of the normalized water cluster count rates for the ambient humidity calibrations. The relative humidity of the sample air was measured with a sensor and was well correlated with $\text{H}^+(\text{H}_2\text{O})_2$ count rates. HCHO normalized sensitivity displayed an exponential dependence on the water cluster count rates. At the highest water cluster count rates, corresponding to a dew point of 22°C , the normalized sensitivity for HCHO was 2.7 Hz/ppbv. At the lowest water cluster count rates, corresponding to a dew point of 1°C , the normalized sensitivity was 5.9 Hz/ppbv. Most VOC species displayed a

Table 2. Averaged normalized sensitivities (Hz/ppbv per MHz H_3O^+).

<i>m/z</i>	Species	Ambient	Dehumidified at -30°C		
		RH	Td=120	Td=100	Td=80
31	formaldehyde	3.4 ± 0.8	11.3 ± 1.1	16.9 ± 1.1	25.0 ± 1.5
33	methanol	10.5 ± 0.6	10.4 ± 0.6	13.4 ± 0.5	19.1 ± 1.2
42	acetonitrile	17.9 ± 1.1	17.3 ± 0.6	23.7 ± 0.3	35.4 ± 0.7
45	acetaldehyde	14.2 ± 0.9	13.5 ± 0.5	18.5 ± 0.2	26.4 ± 0.4
59	acetone	15.2 ± 0.9	14.6 ± 0.5	19.9 ± 0.5	29.7 ± 0.6
69	isoprene	10.0 ± 0.6	9.5 ± 0.5	14.0 ± 0.3	20.3 ± 0.4
71	methacrolein	8.6 ± 0.5	8.1 ± 0.4	11.9 ± 0.3	17.6 ± 0.3
73	2-butanone	19.1 ± 1.1	18.5 ± 0.7	25.5 ± 0.6	37.8 ± 0.7
79	benzene	9.7 ± 0.4	10.2 ± 0.6	13.3 ± 0.3	18.4 ± 0.3
93	toluene	10.0 ± 0.5	10.1 ± 0.5	13.2 ± 0.3	18.5 ± 0.3
105	styrene	10.5 ± 0.5	10.6 ± 0.6	13.6 ± 0.4	19.4 ± 0.3
107	p-xylene	9.2 ± 0.5	9.2 ± 0.4	11.8 ± 0.3	17.1 ± 0.3
121	1,2,4-trimethylbenzene	8.5 ± 0.5	8.6 ± 0.5	10.8 ± 0.4	15.7 ± 0.3
135	1,2,3,5-tetramethylbenzene	8.1 ± 0.8	8.0 ± 0.7	9.5 ± 0.4	13.7 ± 0.3
137	α -pinene	3.2 ± 0.2	3.1 ± 0.2	4.5 ± 0.2	7.5 ± 0.2

**Fig. 5.** Dependence of normalized sensitivities on water cluster count rates. Water cluster count rates were normalized to MHz of H_3O^+ . Panel (A) shows acetonitrile, acetaldehyde and isoprene as examples of the positive linear dependence most species displayed. Panel (B) illustrates a weaker dependence for benzene and a negative exponential dependence for HCHO.

positive linear dependence on the cluster count rates, indicating humidity dependent sensitivities likely due to reactions with $\text{H}^+(\text{H}_2\text{O})_2$. Benzene displayed the weakest dependence as expected, since it is not thought to react with the water dimer. The fits to these calibrations were used together with measured water cluster count rates to determine mixing ratios from the ambient ion signals. The averaged normalized sensitivities for ambient humidity calibrations compared to the dehumidified calibrations are shown in Table 2. On average,

reducing water vapour concentration to -30°C dew point increased the HCHO normalized sensitivity by a factor of 3.

For the dehumidified calibrations, the average normalized water cluster count rates were 4700 ± 800 . It is noted from Table 2 that the dehumidified calibration values were consistently and significantly greater than the trend line extrapolation to low humidity in Fig. 5. The reason for the higher than expected sensitivities is that H_3O^+ count rates were consistently lower in the dehumidified mode. On average,

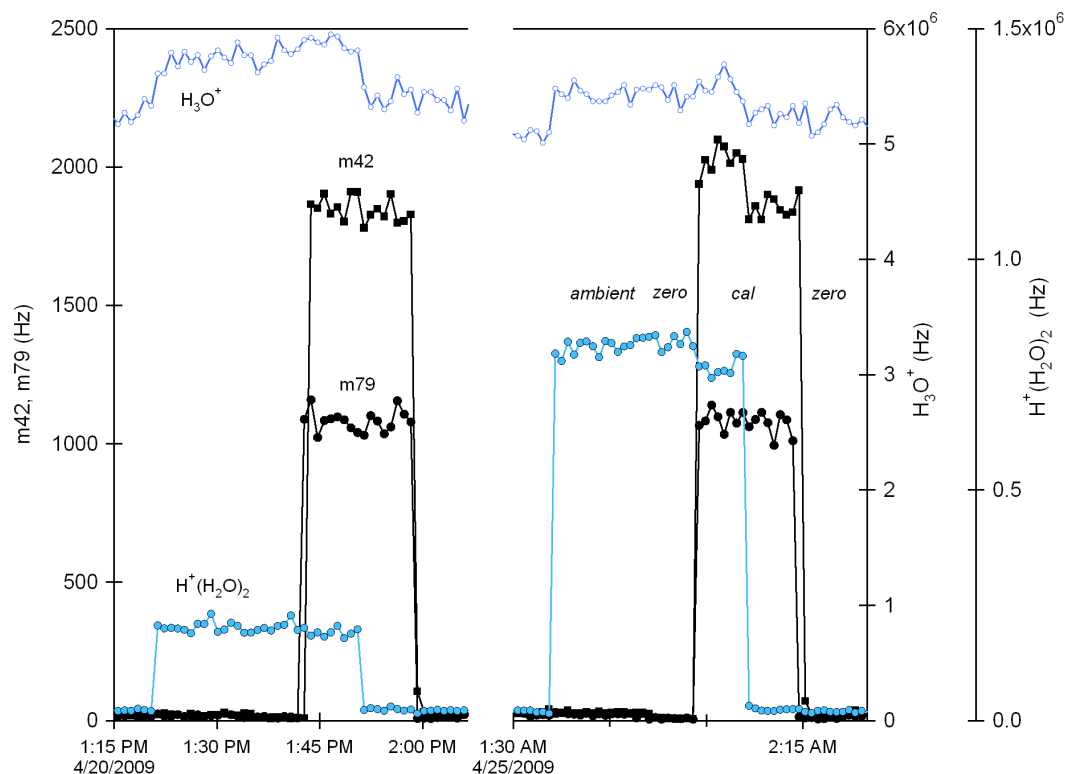


Fig. 6. Acetonitrile (m42, squares), benzene (m79, circles), H_3O^+ (open blue circles) and $\text{H}^+(\text{H}_2\text{O})_2$ (filled blue circles) ion signals during calibration. Shown, are two different days representing relatively dry (20 April) and humid (25 April) sampling conditions. For both time periods the data show a sequence of ambient air sampling (ambient), followed by zero air sampling (zero), addition of calibration gases (cal) and a final zero air period. The decrease in $\text{H}^+(\text{H}_2\text{O})_2$ signal during calibration was due to the addition of dry calibration gas to the humid zero air flow. Sampling through the water trap is apparent by the decrease in $\text{H}^+(\text{H}_2\text{O})_2$ count rates. A significant decrease in H_3O^+ was always observed when sampling through the water trap.

the H_3O^+ count rates were $6.7 \pm 2.3\%$ less when sampling through the water trap during calibration periods. For many VOCs there was little difference in the ion signal during calibration between ambient RH and dehumidified sampling for dry conditions. This is shown in Fig. 6 where 2 field calibration periods are depicted: 20 April representing relatively dry conditions and 25 April representing relative humid conditions as indicated by the $\text{H}^+(\text{H}_2\text{O})_2$ count rates. For benzene ($m/z=79$), there was no discernable change in ion signals when the PTR-MS sampled through the water trap for either calibration period. However, the decrease in H_3O^+ count rates when sampling through the water trap, -7.6% for the 20 April calibration and -4.3% for the 25 April calibration, causes the normalized sensitivity to be greater for dehumidified sampling. For acetonitrile ($m/z=42$), there was no significant difference in ion signals between ambient humidity and water trap calibration for the April 20 period, thus, the dehumidified normalized sensitivity is 7.6% larger because of the difference in H_3O^+ count rates. For 25 April, the ambient humidity calibration displayed a significantly higher count rate than on 20 April and a significant drop in the count rate through the water trap. During this period, the $\text{H}^+(\text{H}_2\text{O})_2$ ion

count rate was 15% of the H_3O^+ count rate compared to 3.4% for the 20 April ambient humidity calibration. The difference in acetonitrile behaviour between the 2 calibration periods is qualitatively consistent with $\text{H}^+(\text{H}_2\text{O})_2$ acting a reagent ion. It was clear from the calibrations that the PTR-MS sensitivity in dehumidified mode could not be extrapolated from the ambient humidity trend lines given in Fig. 5.

Figure 7 illustrates the impact of the water trap on $\text{H}^+(\text{H}_2\text{O})_2$ and formaldehyde ($m/z=31$) count rates. When the trap switched to sampling through the -30°C water trap, count rates for $\text{H}^+(\text{H}_2\text{O})_2$ decreased to $\sim 21\,000$ Hz, while count rates for formaldehyde increased, producing a crenulated pattern. The HCHO mixing ratio could be determined from the $m/z=31$ count rate by applying the respective humidity dependent normalized sensitivity for the ambient humidity data, and the 11.3 Hz/ppbv sensitivity factor for the data collected at -30°C . Count rates for H_3O^+ were reasonably steady at 5×10^6 Hz during the entire field campaign. The resulting HCHO mixing ratios from the 2 sampling conditions were well matched, indicating the humidity dependent sensitivity was accounted for correctly and there were no significant losses of HCHO to the water trap. It is apparent in the

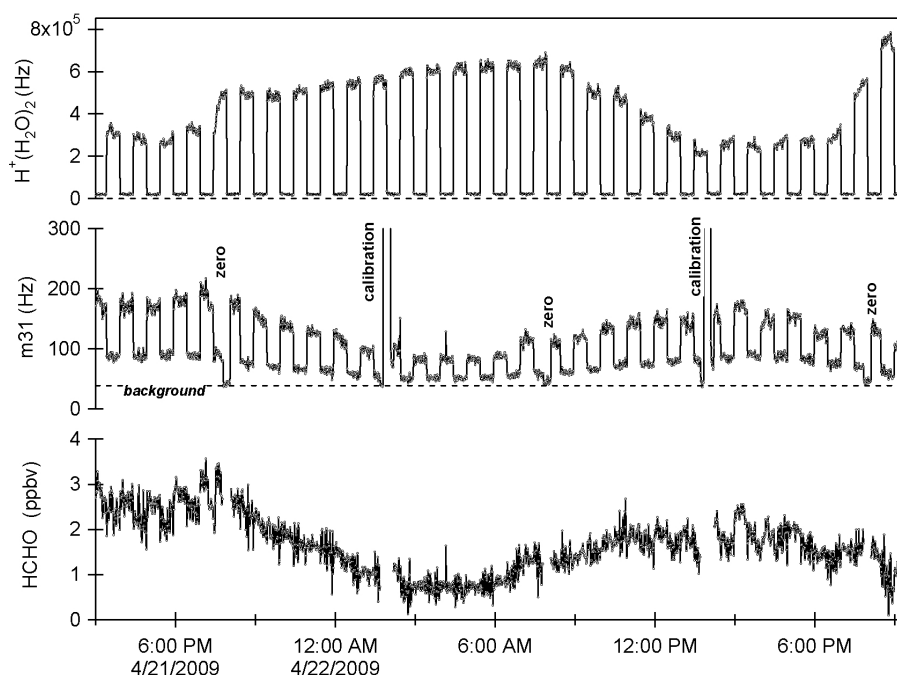


Fig. 7. Time series of $\text{H}^+(\text{H}_2\text{O})_2$ count rates in upper panel, HCHO count rates in middle panel (m31), and HCHO mixing ratios in lower panel. The crenulations in the $\text{H}^+(\text{H}_2\text{O})_2$ and m31 count rates are due to switching between ambient humidity sampling and dehumidified sampling conditions. Calibration and zero periods are indicated. HCHO mixing ratios were determined from the measured m31 count rates using separate calibration factors for the 2 sampling conditions.

figure that the worst discrepancy between -30°C sampling and ambient humidity sampling occurred when the air was dry, as indicated by a crenulated pattern in the HCHO mixing ratio. HCHO normalized sensitivity displayed a strong dependence on water vapour at low water vapour concentrations and the dependence may not have been adequately captured by the calibration curve in Fig. 5.

The reconstruction of the mixing ratio time series for several ions was complicated by significant differences in background count rates between dehumidified and ambient humidity sampling. The most notable differences were for $m/z=45$ (acetaldehyde) and $m/z=47$. In both instances, the dehumidified backgrounds were much higher, by factors of 2.0 and 4.2, respectively. Contributions to the $m/z=45$ ion from CO_2H^+ and at $m/z=47$ from the $\text{N}_2\text{H}_3\text{O}^+$ cluster ion have been noted (Hanson et al., 2009) as well as humidity dependent background count rates for $m/z=45$ (de Gouw and Warneke, 2007). Why more $\text{N}_2\text{H}_3\text{O}^+$ would form with less water vapour in the drift is somewhat puzzling, but suggests that it may be coming from the ion source rather than formed in the vacuum interface of the mass spectrometer.

Table 2 shows a significant increase in normalized sensitivity was achieved when the PTR-MS was operated at lower E/N ratios. For the 100 Td sampling period, the HCHO normalized sensitivity was 16.9 Hz/ppbv and for the 80 Td sampling period it was 25.0 Hz/ppbv. The H_3O^+ count rates were 5.0×10^6 Hz for 100 Td sampling and 4.6×10^6

Hz for 80 Td sampling, resulting in absolute sensitivities of 84.5 Hz/ppbv and 115 Hz/ppbv, respectively. The measured ion signal of $\text{H}^+(\text{H}_2\text{O})_2$ relative to H_3O^+ was 4% at 100 Td and 17% at 80 Td. Following the procedure described by de Gouw and Warneke (2007), the HCHO detection limit for the 100 Td sampling period was calculated from the average 36 Hz background signal and 5 s dwell time to be 95 pptv. At 80 Td, the detection limit was calculated to be 78 pptv. Karl et al. (2003) reported formaldehyde detection limits ($S/N=2$) of 1–2 ppbv using a PTR-MS operated at 123 Td, 2.5 mbar drift pressure and 4 MHz H_3O^+ for a field study conducted in Houston during summer. Inomata et al. (2008) estimated a detection limit ($S/N=2$) of 0.2–0.5 ppbv depending on ambient humidity. Removing water vapour clearly improved the lower limit of formaldehyde detection by PTR-MS instruments. However, the detection limits calculated are based on detector noise and are not method detection limits that account for potential sampling losses of HCHO at low mixing ratios to the water trap and other tubing.

3.4 Loss of compounds to water trap

Loss of compounds to the water trap was evaluated for low vapour pressure compounds that would be expected to be lost to cold surfaces such as C_3 -alkylbenzenes and C_4 -alkylbenzenes. No losses were apparent during calibrations at 19.8 and 5 ppbv, respectively for 1,2,4-trimethylbenzene

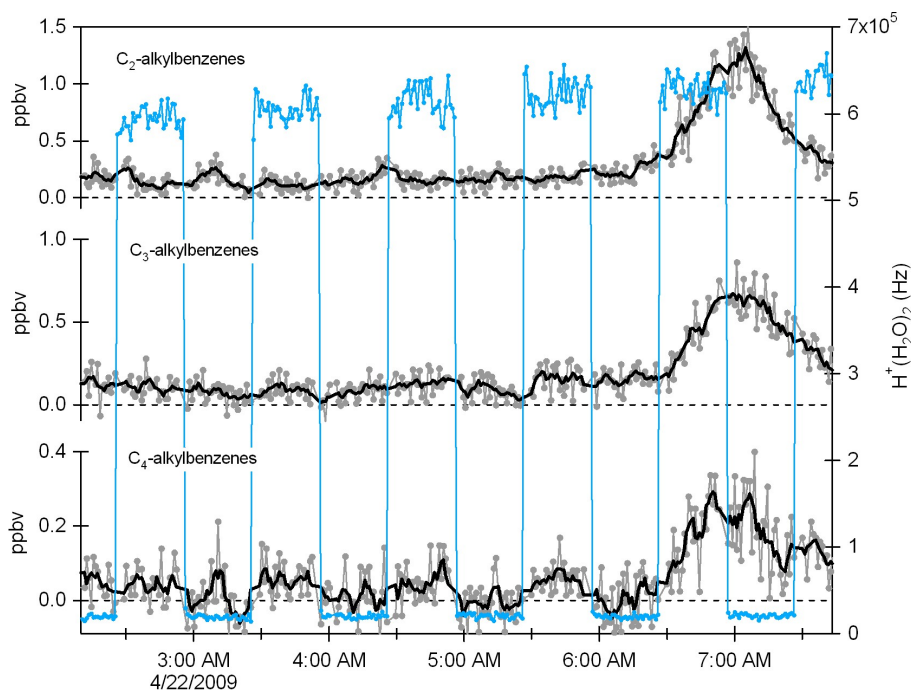


Fig. 8. Time series of alkylbenzene mixing ratios on left axes and $\text{H}^+(\text{H}_2\text{O})_2$ ion count rates (blue trace) on the right axis, illustrating loss of C_4 -alkylbenzenes through the water trap at low mixing ratios. Alkylbenzene data are given by grey circles. Solid black line through the data is a 5 point box smooth to better display trends.

and 1,2,3,5-tetramethylbenzene. The ambient data were also evaluated and a selected time series of C_2 , C_3 , and C_4 -alkylbenzenes mixing ratios is shown in Fig. 8. The clear square wave oscillation in the $\text{H}^+(\text{H}_2\text{O})_2$ ion signal indicates when the PTR-MS was sampling dehumidified air from the water trap at -30°C . The aromatics are a good test of water trap losses because their normalized sensitivity displayed a weak dependence on ambient humidity and there was no difference in their backgrounds between the 2 sampling conditions. No losses for C_2 and C_3 -alkylbenzenes were observed in the data, as illustrated by the selected time series in Fig. 8. Losses of C_4 -alkylbenzenes to the water trap were evident at low mixing ratios (<100 pptv), as indicated by ion signals that were not significantly different from background count rates when sampling through the water trap. The C_4 -alkylbenzenes were the only compounds from Table 1 that displayed any observable loss. We note that for the ions $m/z=47$ and $m/z=61$, attributed to formic and acetic acid, significant loss of ion signal was observed through the water trap.

3.5 HCHO interferences

Inomata et al. (2008) reported a positive interference on the formaldehyde ion due to methanol. This can be caused by the O_2^+ + methanol reaction to create an ion at $m/z=31$ (CH_3O^+). O_2^+ is produced in the PTR-MS ion source and

its presence can be controlled by adjusting water vapour flow rates through the source and extraction voltages into the drift. For the SHARP experiment O_2^+ ion signal was kept at $\sim 1.2\%$ of H_3O^+ ion signal. The level of the interference was established in the field by determining the $m/z=31$ ion signal during automated instrument calibrations from 14–16 April. During this period, the HCHO permeation tube was not yet integrated into the calibration system. The methanol calibration level was 19.8 ppbv. We found no significant difference in the normalized $m/z=31$ ion signal between the zero air period immediately before the calibration (7.0 ± 0.6 Hz per MHz H_3O^+) and the calibration period (7.2 ± 0.9 Hz per MHz H_3O^+) for the seven zero-cal period pairs. The average difference (cal – zero) was 0.33 ± 0.49 Hz per MHz H_3O^+ . We concluded that methanol interference was not significant for our HCHO data.

Another probable interference noted by Inomata et al. (2008) and Steinbacher et al. (2004) is methyl hydroperoxide (CH_3OOH) which likely undergoes dissociative proton transfer reaction with H_3O^+ to produce CH_3O^+ ions. Inomata et al. (2008) reported significant fragmentation of CH_3OOH producing $m/z=31$ and $m/z=49$ ion signals. In our experience with our instrument operating at 120 Td, there is no significant ion signal at $m/z=49$, the CH_3OOHH^+ ion. For the SHARP field experiment, the drift field intensity was reduced during the month of May to 100 Td, and then to 80 Td for the last week of May. Under these conditions a significant

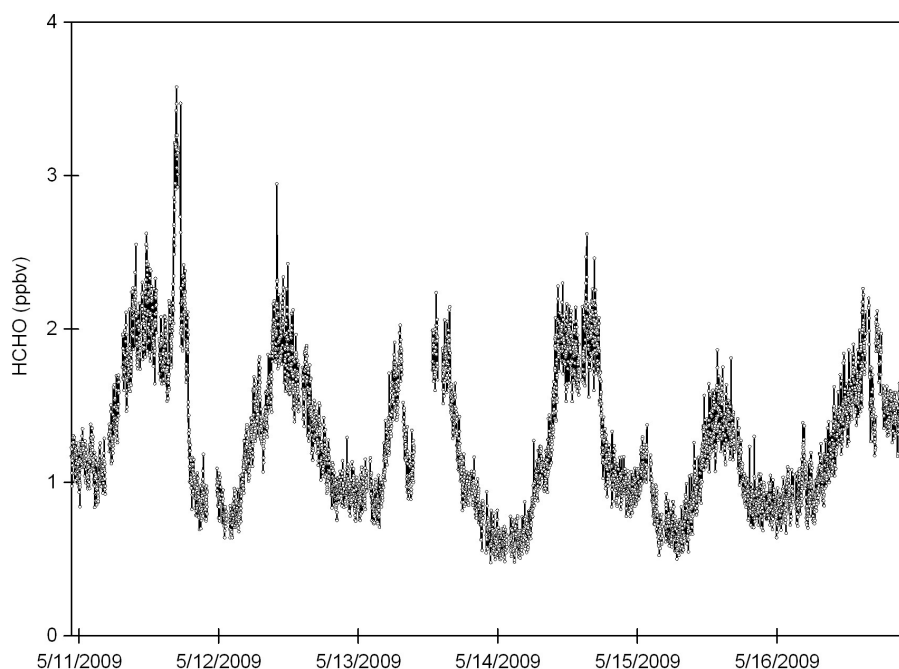


Fig. 9. Formaldehyde data illustrating signal-to-noise compared to amplitude of diel variation. There is a data point every 73 s and the formaldehyde dwell time was 5 s. The sample was dried to -30°C dew point and the PTR-MS operated at 100 Td and a pressure of 2.1 mbar.

ion signal at $m/z=49$ was observed. A small fraction of this ion signal was due to the protonated formaldehyde water cluster ion. The estimated interference was 0.09 Hz per ppbv of formaldehyde at 100 Td. Assuming CH_3OOH has a similar normalized sensitivity as methanol, this would amount to only ~ 8 pptv CH_3OOH per ppbv of formaldehyde. It seems possible that operating the PTR-MS at lower field intensities would reduce interferences from CH_3OOH and allow CH_3OOH to be measured. The level of interference from CH_3OOH on the $m/z=31$ ion signal could not be quantified in the field study. This issue will be the subject of a future study.

3.6 Dehumidified field sampling

Figure 9 shows a time series plot for a 2 day period for the 100 Td operating condition to illustrate the signal-to-noise of the HCHO data. Clear diel variations with amplitude of ~ 1 ppbv could be observed with nighttime/early morning minima and maxima during the day. During this time, air flow was from the south and relatively unpolluted. Clear diel variations in isoprene ($m/z=69$) and its photoproducts methacrolein and methyl vinyl ketone ($m/z=71$) were observed so a likely mechanism of diel HCHO variation was in its production rates from isoprene oxidation (Sumner et al., 2001). Lowest observed HCHO mixing ratios were 480 pptv, well above the calculated detection limits. The minimum mixing ratios were consistent with minimums reported for

Houston (Wert et al., 2003; Karl et al., 2003; Dasgupta et al., 2005; Eom et al., 2008). A small plume of HCHO was observed on 11 May that may indicate industrial emissions. A separate analysis of the HCHO and VOC data collected in this study will be presented elsewhere.

Figure 10 illustrates the correspondence between HCHO mixing ratios and the ion signal at $m/z=49$ which we interpret as CH_3OOH . The figure displays 5 min average data. Mixing ratios of CH_3OOH were determined assuming it had the same normalized sensitivity as methanol. This is an assumption made only to evaluate the temporal behaviour and potential merit of CH_3OOH determination by PTR-MS instruments. CH_3OOH displayed a diel cycle similar to that of HCHO. The CH_3OOH maxima were sometimes displaced later in the afternoon, reflecting perhaps lower NO concentrations and, hence, greater production rates of CH_3OOH from the $\text{CH}_3\text{OO}+\text{HO}_2$ reaction. On 17 May, a cold front passed through bringing rainy weather and a change in wind direction from a relatively clean southerly flow to polluted north easterly flow impacted by the heavily industrialized area known as the Houston Ship Channel. CH_3OOH decreased by a factor of 3 during the cold front passage and remained relatively low during the period of polluted northeasterly flow, whereas HCHO mixing ratios were much higher. Plumes of HCHO were observed during this period with the highest mixing ratio of 11.2 ppbv. The $m/z=49$ ion signal data are interesting and suggest the behaviour of peroxide product. This warrants further investigation to determine

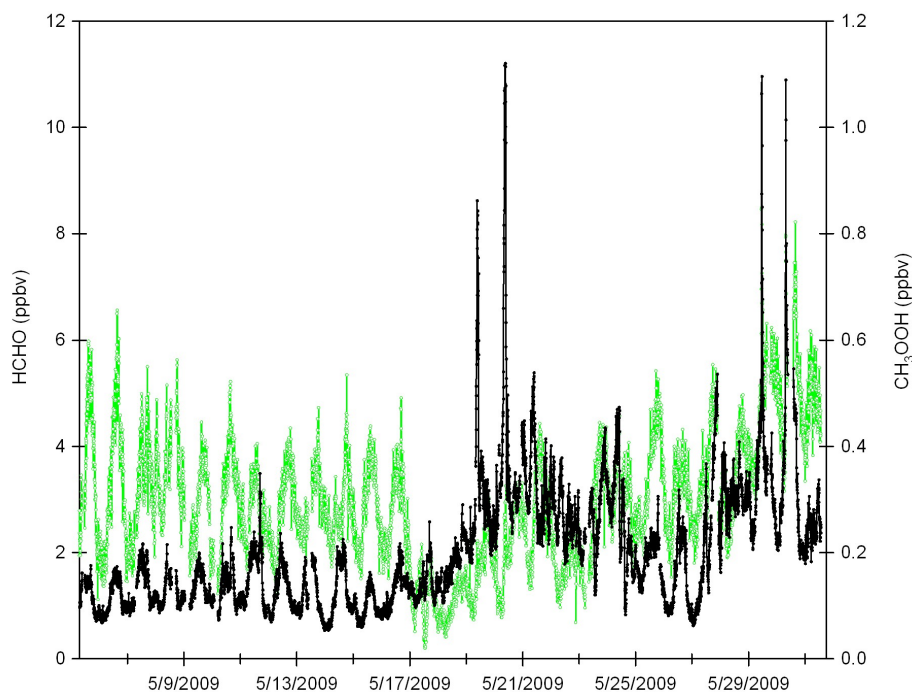


Fig. 10. Time series of 5 min averages for HCHO (black) and the $m/z=49$ ion signal interpreted to be CH_3OOH (green). Both species displayed a clear diel variation with nighttime minimums during a period with persistent southern air flow (5 May–17 May). The decrease in CH_3OOH on 17 May corresponds to the passage of a cold front and rainy weather. The cold front passage also marked a change in air flow from southerly to northeasterly, bringing polluted air from a heavily industrialized region of Houston and higher HCHO mixing ratios.

CH_3OOH transmission efficiency through the water trap and the degree of fragmentation upon reaction with H_3O^+ as a function of E/N .

4 Conclusions

The humidity dependence of the normalized sensitivity (Hz/ppbv per MHz H_3O^+) for formaldehyde and other VOC was investigated in both laboratory experiments and in a field study. Formaldehyde normalized sensitivity displayed an exponential dependence on water vapour concentration. Benzene displayed a weak positive dependence. In field tests, a factor of 7 increase in formaldehyde normalized sensitivity was demonstrated by drying the air sample to a dew point of -30°C . Laboratory tests showed that at temperatures warmer than -30°C there was significant uptake of formaldehyde and other soluble organics to the ice surface, consistent with the presence of a liquid water layer on the ice surface. Benzene, toluene, C_2 -alkylbenzenes, and C_3 -alkylbenzenes passed quantitatively through the trap at -30°C . Evidence of C_4 -alkylbenzene losses were observed at mixing ratios less than 100 pptv. Drying the sample has four benefits: (1) elimination of water vapour dependent sensitivity for all species; (2) significant increase in HCHO normalized sensitivity (3) ability to operate drift tube at low E/N ratios to further increase sensitivity and reduce dissociative

proton transfer reactions, and (4) reduced clustering due to low water vapour in the drift tube. Field testing at an E/N ratio of 100 Td demonstrated that 1 ppbv diel variation in HCHO mixing ratios could be easily resolved. Detection limits at 100 Td were calculated to be 95 pptv, and 78 pptv at an E/N ratio of 80 Td. The formaldehyde normalized sensitivity at 80 Td was 25 Hz/ppbv, yielding an absolute sensitivity of 115 Hz/ppbv. During field testing, a significant ion signal at $m/z=49$ was observed which was attributed to CH_3OOH . This species is potentially quantifiable by PTR-MS instruments if they are operated at low E/N and low water vapour concentrations and warrants further investigation.

Acknowledgements. This work was funded by the Houston Advanced Research Center (HARC). The authors would like to thank colleagues at the University of Houston for organizing the SHARP field campaign.

Edited by: J. Williams

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