

胃癌微卫星不稳定与错配修复蛋白表达的缺失

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Microsatellite instability and loss of mismatch-repair protein expression in gastric carcinoma

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Abstract

AIM: To detect the microsatellite instability (MSI) and expression of mismatch repair (MMR) gene in gastric carcinoma, and to explore the molecular biological mechanism underlying the carcinogenesis of gastric carcinoma.

METHODS: A total of 56 cases of gastric carcinomas and surrounding non-cancerous tissues from surgical excision samples were collected, among which 22 cases were well and 34 cases were poorly differentiated adenocarcinoma, 20 cases were in early stage and 36 cases were in advanced stage of the disease. The microsatellite locus of BAT-26, D17S261, D3S1283, D2S123 and D3S1611 were amplified by PCR after DNA abstraction. Then PCR products were mixed together with GeneScan 500 size standard followed by heat denaturation. Microsatellites were analyzed by capillary electrophoresis with 6% SLPA and 8 mol/L⁻¹ urea as sieving medium. Carcinoma were characterized as high MSI (MSI-H) if they manifested instability at two or more

markers, low MSI (MSI-L) if unstable at only one marker, and microsatellite stable (MSS) if they showed no instability at any marker. Expression of MMR gene hMLH1 and hMSH2 were detected by immunohistochemical staining using the streptavidin-biotin-peroxidase complex method with 3, 3'-diaminobenzidine as chromogen.

RESULTS: Of the 56 cases of gastric carcinomas, 14 cases (25%) showed MSI-H and 14 cases (25%) showed loss of MMR. In the 14 cases of the MSI, 11 cases (79%) were accompanied by loss of hMLH1/hMSH2 expression, whereas in the 42 cases of the MSI-L/MSS, only 3 cases (7%) were accompanied by loss of hMLH1/hMSH2 expression. MSI was significantly related with mismatch repair deficiency ($P < 0.01$). Of the 22 cases of well-differentiated carcinomas, 7 cases (32%) manifested MSI-H and 6 (27%) cases showed protein defection of MMR. Comparatively, 7 cases (21%) manifested MSI-H and 8 cases (24%) showed protein defection of MMR in 34 cases poorly-differentiated carcinomas. Of the 20 cases early stage carcinomas, only 1 cases (5%) manifested MSI-H and 3 (15%) cases showed protein defection of MMR, whereas 13 cases (36%) manifested MSI-H and 11 cases (31%) showed protein defection of MMR in 36 cases advanced carcinomas. The MSI frequency was higher in advanced stage (36%) than that in early stage (5%) of gastric carcinoma and the difference was significant ($P < 0.05$), but no difference between well and poorly differentiated gastric carcinoma. The difference of loss frequency of hMLH1/hMSH2 expression was not significant in different stage and different differentiation of gastric carcinoma.

CONCLUSION: The defect of mismatch repair may be involved in the carcinogenesis of a subset of gastric cancer but not in the biologic behavior. MSI frequency increases with the progression of gastric carcinoma.

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摘要

目的: 分析胃癌组织微卫星不稳定性、错配修复蛋白表达以及二者的关系, 探讨胃癌发生的分子生物学机制。

方法: 胃腺癌及癌旁组织标本 56 例, 高分化癌 22 例, 中低分化癌 34 例, 早期癌 20 例, 中晚期癌 36 例, 常规酚-氯仿法提取 DNA, 选取基因组上的五个微卫星位点 BAT-26、D17S261、D3S1283、D2S123 和 D3S1611, 进行 PCR 扩增, 扩增产物加入 GeneScan 500 size standard 共同热变性后, 用 60 g/L 的 SLPA 添加 8 mol/L 尿素做筛分递质的毛细管电泳进行分析。被检测的五个微卫星位点如出现两个或两个以上位点的不稳定, 定为微卫星的高

度不稳定(MSI-H), 一个位点出现不稳定、定为微卫星的低度不稳定(MSI-L), 没有位点出现不稳定、定为微卫星稳定(MSS). 石蜡切片常规免疫组织化学SP方法检测hMLH1和hMSH2蛋白的表达, 肿瘤组织上皮细胞核不着色, 而周围组织上皮细胞核着色判定为hMLH1或hMSH2蛋白表达的缺失.

结果: 在56例胃癌中, 有14例(25%)表现为MSI-H, 14例(25%)有错配修复蛋白表达的缺失. 在14例MSI-H的胃癌组织中, 11例(79%)有hMLH1或hMSH2表达的缺失, 42例MSI-L/MSS的胃癌组织仅3例(7%)有hMLH1或hMSH2表达的缺失. 胃癌的MSI-H与错配修复蛋白表达的缺失高度相关($P < 0.01$). 其中的22例高分化腺癌有7例(32%)表现为MSI-H, 6例(27%)有错配修复蛋白表达的缺失, 34例中低分化腺癌7例(21%)表现为MSI-H, 8例(24%)有错配修复蛋白表达的缺失, 20例早期腺癌有1例(5%)表现为MSI-H, 3例(15%)有错配修复蛋白表达的缺失, 36例中晚期腺癌13例(36%)表现为MSI-H, 11例(31%)有错配修复蛋白表达的缺失. 微卫星不稳定性在中晚期胃癌明显高于早期胃癌($P < 0.05$), 但在不同分化程度的胃癌差异不明显; MMR蛋白表达缺失在高分化与低分化以及早期与中晚期的胃癌均无显著性差异.

结论: 细胞错配修复功能缺陷与部分胃癌的发生有关, 而与胃癌的生物学行为无关; 胃癌的微卫星不稳定性随着肿瘤的演进而增加.

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0 引言

胃癌是一种发病率和死亡率都很高的一种恶性肿瘤, 其发病的分子机制目前尚不清楚. 人体细胞错配修复(mismatch repair MMR)系统的主要功能是修复DNA复制时形成的错误, 维持人体基因组的稳定性^[1-3]. 当MMR的功能障碍时DNA复制时形成的错误因得不到修复而在细胞内积累^[4-5]. 基因突变的积累则是多种肿瘤发生的分子基础. 细胞错配修复功能的丧失是欧美多见的遗传性非息肉性大肠癌(hereditary nonpolyposis colorectal carcinoma, HNPCC)发病的主要原因之一^[6-7]. 我国常见的胃癌的发生是否也与错配修复功能缺陷有关? 我们分析胃癌微卫星不稳定性与错配修复蛋白hMLH1和hMSH2表达的关系, 对这一问题进行探讨.

1 材料和方法

1.1 材料 胃癌及癌旁组织标本56例取自中国东北地区临床外科手术患者, 年龄28-71(平均57岁), 男32例, 女24例, 其中早期癌20例, 中晚期癌36例, 高分化癌22例, 中低分化癌34例, 手术标本部分用

石蜡包埋, 部分于-80℃保存, 备用. 用常规酚-氯仿法提取组织DNA. Perkin-Elmer Model 2700 PCR扩增仪、ABI Prism 310基因分析仪、商品化分离胶(美国ABI公司); P/ACE MDQ型毛细管电泳(美国Beckman公司); 丙烯酰胺、四甲基乙二胺(Temed)、过硫酸胺(APS)、 γ -甲基丙烯酰基-三(甲氧基)硅烷与TBE电泳缓冲液(Sigma), PCR反应试剂、引物(宝生物工程公司), 抗hMSH2小鼠mAb(美国Calbio Chem公司), 抗hMLH1小鼠mAb(美国Pharm Ingen公司), 免疫组化试剂盒(北京中山生物技术有限公司).

1.2 方法 选取五个微卫星位点, 引物设计参考文献^[8], 序列(见表1). PCR反应体积12.5 μ L, 其中模板DNA 50 ng, 4 \times dNTPs终浓度各0.2 mmol/L, MgCl₂的浓度1.5 mmol/L, Taq酶0.5 U. 循环条件: 94℃预变性5 min后, 94℃ 30 s, 58-60℃ 30 s, 72℃ 30 s, 35个循环周期, 最后72℃延长5 min. PCR产物各取0.5 μ L, 加入1 μ L GeneScan 500 size standard和12 μ L水, 然后95℃变性5 min后迅速冰浴冷却, 待测. 电泳时由毛细管电泳仪压力系统自动填充筛分递质到毛细管中(总长47 cm, 有效长度36 cm, 内径50 μ m), 60 g/L的SLPA添加8 mol/L/L尿素做筛分递质, 分析温度60℃, 分离电压15 kV, 激光诱导荧光检测器检测DNA片段, GeneScan数据处理软件采集数据. 被检测的五个微卫星位点如出现两个或两个以上位点的不稳定, 定为微卫星的高度不稳定(MSI-H), 一个位点出现不稳定定为微卫星的低度不稳定(MSI-L), 没有位点出现不稳定定为微卫星稳定(MS-S). 石蜡切片常规免疫组织化学SP方法检测hMLH1和hMSH2蛋白的表达, 厚度为4 μ m, hMLH1mAb稀释度为1:60, hMSH2mAb稀释度为1:80, 37℃温育1 h. 以后步骤均按试剂盒说明书进行, 用DAB呈色. 肿瘤组织上皮细胞核不着色, 而周围组织上皮细胞核着色判定为hMLH1或hMSH2蛋白表达的缺失.

表1 微卫星位点的引物序列

位点	引物序列	退火温度(℃)
BAT-26	5'-FAM-TGACTACTTTGACTTCAGCC	58
	5'-AACCAATTCACATTTTAACCC	
D17S261	5'-HEX-AGGGATACTATTACAGCCCGAGGTG	60
	5'-ACTGCCACTCCTTGCCCCATTC	
D3S1283	5'-TET-GGCAGTACCACCTGTAGAAATG	60
	5'-GAGTAACAGAGGCATCGTGATTC	
D2S123	5'-FAM-AAACAGGATGCCTGCCTTTA	60
	5'-GGACTTCCACCTATGGGAC	
D3S1611	5'-HEX-CCCCAAGGCTGCACCTT	60
	5'-AGCTGAGACTACAGGCATTG	

2 结果

胃癌56例中, 微卫星高度不稳定14例, 低度不稳定

4例, 稳定38例; 由于微卫星低度不稳定病例不足5例, 统计时与微卫星稳定病例合并. 据此, 将56例胃癌分为二组: 微卫星不稳定组(MSI)14例(25%)和微卫星稳定组(MSS)42例(75%). 胃癌组织中, 14例(25%)表现出错配修复蛋白表达缺失, 14例微卫星不稳定的胃癌组织中, 11例(79%)有hMLH1蛋白或hMSH2蛋白表达的缺失, 42例微卫星稳定的胃癌组织仅3例(7%)有hMLH1或hMSH2表达的缺失(表2), 胃癌的微卫星不稳定与错配修复蛋白表达的缺失高度相关($P < 0.01$). 图1所示5个微卫星序列在相同电泳条件下的电泳图谱; 图2所示胃癌组织hMLH1和hMSH2蛋白表达的免疫组织化学染色.

表2 胃癌微卫星不稳定性与错配修复蛋白表达缺失的比较

微卫星位点	MMR 表达缺失(n)		合计
	hMLH1	hMSH2	
MSI(14例)	10	3	79% (11/14) ^a
MSS(42例)	2	1	7% (3/42)

MSI- 微卫星不稳定, MSS- 微卫星稳定, MMR- 错配修复; ^aMSI组hMSH2表达缺失的3例胃癌有2例伴随hMLH1表达的缺失, MSI与MSS组MMR蛋白表达缺失率比较 $\chi^2=28.57$, $P < 0.01$.

根据临床分级和分期对56例胃癌的微卫星不稳定性与错配修复蛋白表达缺失进行分析(表3), 结果进展期胃癌的微卫星不稳定性明显高于早期胃癌, 而在不同分化程度的胃癌之间微卫星不稳定性无明显差异; 错配修复蛋白表达缺失在不同的分化程度和临床分期的胃癌差异不明显.

表3 胃癌微卫星不稳定和MMR表达缺失与临床分级、分期的分析

分组	MSI		MMR 表达缺失	
	n	P值	n	P值
高分化 (22)	7 (32%)	0.50	6 (27%)	0.75
中低分化 (34)	7 (21%)		8 (24%)	
早期 (20)	1 (5%)	0.01 ^a	3 (15%)	0.33
中晚期 (36)	13 (36%)		11 (31%)	

MSI- 微卫星不稳定, MMR- 错配修复; ^a微卫星不稳定性在中晚期胃癌明显高于早期胃癌($\chi^2=6.64$, $P < 0.05$); 不同分化程度的胃癌微卫星不稳定性差异不明显, 高分化与低分化以及早期与中晚期的胃癌MMR蛋白表达缺失均无显著性差异.

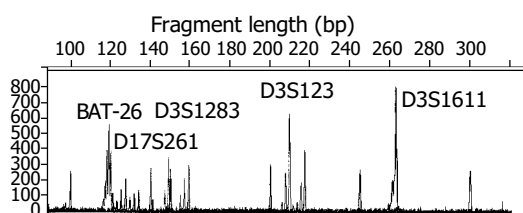


图1 5个微卫星序列的电泳图. (红色峰是 GeneScan 500 size 标准峰) 电泳条件: 毛细管 47cm \times 50 μ m 内径; 缓冲液 1 \times TBE 内含 8 mol/L 尿素; 分离速度 60 g/L SLP A+8 mol/L 尿素; 进样量 15 kV \times 5 s; 分离温度 60 $^{\circ}$ C; 分离电压 -15 kV. 在该条件下, 可以对5个微卫星位点同时进行检测.

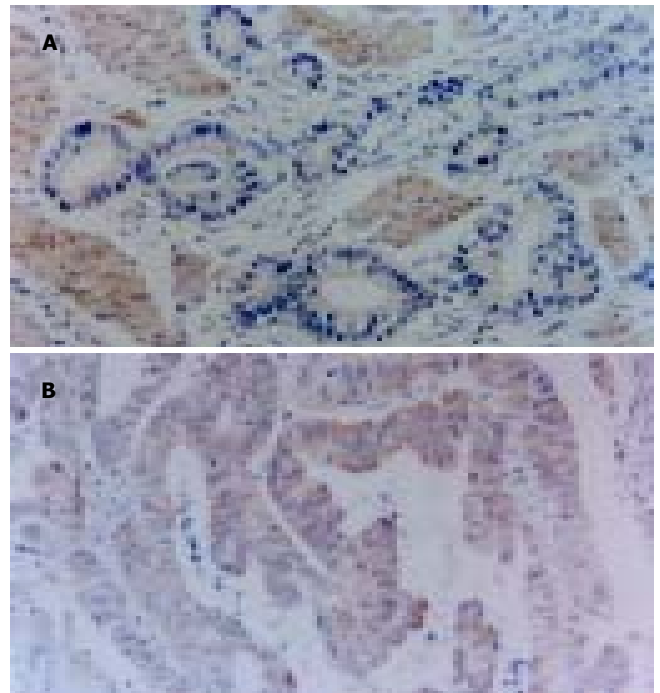


图2 胃癌组织hMLH1和hMSH2蛋白表达的免疫组化染色. A: 浸润到肌层的胃癌组织hMLH1蛋白表达的缺失: 癌组织细胞核不着色, 而癌周围组织细胞核着色; B: hMSH2表达阳性的胃癌细胞核的着色(放大倍数 \times 200).

3 讨论

微卫星是由1-6个碱基组成的简单重复序列, 广泛分布于人类基因组内^[9-10]. DNA复制时, 微卫星序列很容易形成错配或重组错误. 当细胞的错配修复系统功能障碍时, 这些错误得不到修复, 微卫星表现为重复单位的增加或减少, 称为微卫星不稳定性(microsatellite instability, MSI)^[11-12]. 15%的大肠癌具有遗传背景, HNPCC的发生率占全部结肠癌的0.5-13%^[13-15]. 有报告90%的HNPCC的肿瘤细胞中都能检测到MSI, 且影响MSI的主要是单个和二个碱基的重复序列^[16-19]. 我们在本实验中选取的微卫星位点也是由单个和二个碱基的重复序列, 运用聚丙烯酰胺毛细管电泳在25%的胃癌组织中检测到了MSI, 这与大肠癌相似, 可以认为这些胃癌的发生与胃黏膜细胞的错配修复功能缺陷有关. 而另一部分胃癌的发生可能与错配修复功能缺陷无关. 这提示多种不同机制可能参与胃癌发生的过程.

HMLH1和hMSH2基因是错配修复基因家族中的重要成员, 其编码的蛋白质作为错配修复体系中的起连接和识别作用的亚基, 在错配修复过程中起重要作用. 在HNPCC, 由于这两种基因的突变或蛋白表达的异常而造成的细胞错配修复功能障碍的占80%以上^[20-24]. 到目前为止, 已经检测到400多种错配修复基因的突变, MLH1约占50%, MSH2约占40%(国际合作组织关于HNPCC网站: <http://www.nfdht.nl>)^[25-28]. 我们在25%的胃癌组织中检测到错配修复基因hMLH1或hMSH2蛋白表达的缺失, 这与MSI的出现率恰好吻合. MSI的胃癌中有79%出现MMR蛋白表达的缺失, 我们可以推测由于hMLH1和hMSH2

(主要是 hMLH1) 蛋白缺失而导致的细胞错配修复功能缺陷在 MSI 的胃癌的发生过程中起重要作用。

对 MSI 和 MMR 表达缺失的胃癌根据不同分化程度和不同临床分期进行进一步的分析发现: MSI 随着胃癌的演进而增加, 而与胃癌的分化程度关系不明显; MMR 的表达缺失与胃癌的生物学行为无关。这从另一个侧面说明, 与结肠癌一样, 细胞 MMR 缺陷是胃癌发生的早期事件, 也就是说, 胃黏膜细胞 MMR 缺陷导致了 MSI 以及基因组的不稳定, 而基因组的不稳定使胃黏膜细胞恶变的危险性增高。反过来, 胃癌在演进过程中也促进了基因组的不稳定性的累积。

典型的 HNPCC 体细胞内存在 MLH1 和 MSH2 的胚系突变, 当携带一条等位基因已发生突变的体细胞受到第二次打击时, 错配修复功能即丧失。至于胃癌的 MMR 缺失是否同 HNPCC 一样来源于遗传, 或者说存在着遗传性胃癌, 还有待于探讨。

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