

离舌橐吾中一新三萜皂甙——离舌橐吾甙 A赵 昱¹ 田 军¹ 贾忠建² 孙汉董^{1*}(¹ 中国科学院昆明植物研究所植物化学开放实验室, 昆明 650204)(² 兰州大学有机化学研究所, 兰州 730000)**LIGUVEITOSIDE A, A NEW TRITERPENOID SAPONIN FROM LIGULARIA VEITCHIANA**ZHAO Yu¹, TIAN Jun¹, JIA Zhong-Jian², SUN Han-Dong^{1*}(¹ Laborotary of Phytochemistry, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204)(² Institute of Organic Chemistry, Lanzhou University, Lanzhou 730000)**关键词** 离舌橐吾, 菊科, 三萜皂甙, 离舌橐吾甙 A, 离舌橐吾醇**Key words** *Ligularia veitchiana*, Compositae, Triterpenoid saponin, Liguveitoside A, Liguveitol

In our previous paper, we reported two new eremophilane derivatives from *Ligularia veitchiana*^[1]. Further research to the polar section of this plant led to the isolation of a new triterpenoid saponin, named Liguveitoside A. Its structure was determined on the basis of the spectral and chemical methods.

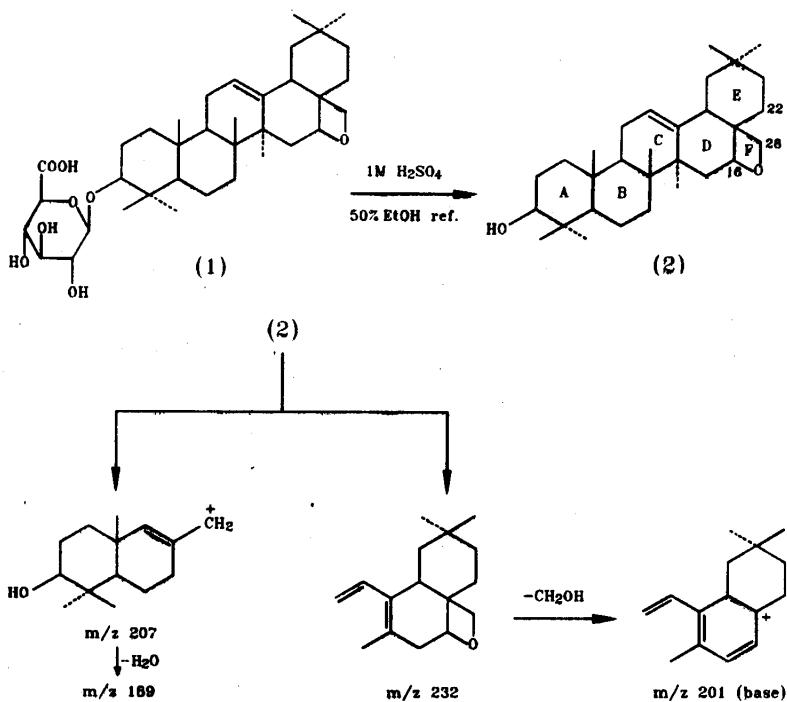
Liguveitoside A (1) was obtained as colorless needles, with a m. p. at 220—221°C, from the n-BuOH soluble part of the EtOH extract of the plant. Comparison of the ¹³C NMR spectrum of 1 with those of the reported saponins^[2] revealed that Liguveitoside A was an oleanane-type monodesmoside. The molecular formula of 1 ($C_{36}H_{56}O_8$) was concluded from the peak at m/z 617 [$M+H$]⁺ in the positive FAB mass spectrum, while its fragment ion peaks were exhibited at m/z 439 [$M-GlcUA$]⁺ and 423 [$M-OGlcUA$]⁺, suggested the presence of a glucuronic acid piece in the molecule. Acid hydrolysis of 1 gave glucuronic acid (detected by PC), and the aglycone (2).

Compound 2 was analysed for seven tertiary methyls (δ 0.73—1.16, 21H, $7 \times$ Me), one $-CH_2O-$ group (an AB system at δ 3.13 and 4.11, 1H each, a pair of doublets, $J_{AB} = 10.8$ Hz), one olefinic proton (δ 5.13, t, $J = 3.5$ Hz) and two oxygen-bearing methine protons (δ 3.17, dd, $J = 10.0, 3.7$ Hz; δ 4.24, ddd, $J = 9.9, 2.4, 1.7$ Hz). The EI mass spectrum of 2 showed a series of diagnostically important mass peaks at m/z 207, 189(207-H₂O), 232 and the base peak at m/z 201 (232-CH₂OH), which were typically attributed to a retro Diels-Alder fragmentation of an olean-12-en derivative bearing one hydroxy group in rings A / B^[3].

Furthermore, a double doublet at δ 3.17 in the ¹H NMR spectrum of 2 with coupling constants at $J = 1.0$ Hz (axial-axial coupling) and $J = 3.7$ Hz (axial-equatorial coupling) corresponded to 3 α -H(axial

• 通迅联系人 Author to whom correspondence should be addressed.

proton), showed the equitorial orientation of the hydroxy group at C-3. According to the unsaturated value of **2**, another ring(named F ring) was most likely to exist in compound **2**. This ring was finally deduced to be a four-membered ring, formed by connection of C-28 and C-16 through an ether link. This could be evidenced by the ^{13}C NMR spectrum of **2**, for the chemical shifts of C-28 and C-16 was somewhat upfield shifted (by γ -effects each other) when comparing with the ^{13}C NMR spectra of the 16-hydroxy derivative and the 28-CH₂OH derivatives [4].



Moreover, one of the AB system protons (H-28) was found to be a doublet ($J = 10.8, 2.4\text{Hz}$), while H-16 at $\delta 4.24$ exhibited as a ddd peak ($J = 9.9, 3.7, 2.4\text{Hz}$), suggested an long distance W-type coupling was present between H-16 and H-28. Shown by the demonstrating model, only an axial orientation of H-16 ($16\alpha-\text{H}$) permits this W-type coupling.

In addition, no variation of C-14 and C-20 in the ^{13}C NMR spectrum of **2** was found when comparing with oleanane-type triterpenoids, also supported the above-mentioned structure.

Liguveitioside A (1): $C_{36}H_{56}O_8$, colorless needles, mp 220–221°C (MeOH). FAB-MS(positive) (m/z): 617 [$M+H^+$]⁺(36), 439(25), 423(87), 286(75), 273(100), 201(65), 176(84). ^{13}C NMR (δ , C5D5N): 38.90(C-1), 26.31(C-2), 89.11(C-3), 40.26(C-4), 55.83(C-5), 18.53(C-6), 34.37(C-7), 41.11(C-8), 47.19(C-9), 36.86(C-10), 23.90(C-11), 122.66(C-12), 144.01(C-13), 43.96(C-14), 36.86(C-15), 66.80(C-16), 39.60(C-17), 44.62(C-18), 47.19(C-19), 31.10(C-20), 33.05(C-21), 26.67(C-22), 28.25(C-23), 16.99(C-24), 15.70(C-25), 16.99(C-26), 27.17(C-27), 69.08(C-28), 33.41(C-29), 24.14(C-30); 107.20(C-1'), 77.80(C-2'), 78.22(C-3'), 73.44(C-4'), 75.59(C-5'), 172.75(C-6').

Liguveitol (2): C₃₀H₄₈O₂, colorless gum, EIMS (*m/z*): 440[M]⁺(92), 423(55), 232(57), 201(100). ¹³C NMR (CDCl₃, δ, ppm): 174.0, 173.0, 172.0, 171.0, 170.0, 169.0, 168.0, 167.0, 166.0, 165.0, 164.0, 163.0, 162.0, 161.0, 160.0, 159.0, 158.0, 157.0, 156.0, 155.0, 154.0, 153.0, 152.0, 151.0, 150.0, 149.0, 148.0, 147.0, 146.0, 145.0, 144.0, 143.0, 142.0, 141.0, 140.0, 139.0, 138.0, 137.0, 136.0, 135.0, 134.0, 133.0, 132.0, 131.0, 130.0, 129.0, 128.0, 127.0, 126.0, 125.0, 124.0, 123.0, 122.0, 121.0, 120.0, 119.0, 118.0, 117.0, 116.0, 115.0, 114.0, 113.0, 112.0, 111.0, 110.0, 109.0, 108.0, 107.0, 106.0, 105.0, 104.0, 103.0, 102.0, 101.0, 100.0, 99.0, 98.0, 97.0, 96.0, 95.0, 94.0, 93.0, 92.0, 91.0, 90.0, 89.0, 88.0, 87.0, 86.0, 85.0, 84.0, 83.0, 82.0, 81.0, 80.0, 79.0, 78.0, 77.0, 76.0, 75.0, 74.0, 73.0, 72.0, 71.0, 70.0, 69.0, 68.0, 67.0, 66.0, 65.0, 64.0, 63.0, 62.0, 61.0, 60.0, 59.0, 58.0, 57.0, 56.0, 55.0, 54.0, 53.0, 52.0, 51.0, 50.0, 49.0, 48.0, 47.0, 46.0, 45.0, 44.0, 43.0, 42.0, 41.0, 40.0, 39.0, 38.0, 37.0, 36.0, 35.0, 34.0, 33.0, 32.0, 31.0, 30.0, 29.0, 28.0, 27.0, 26.0, 25.0, 24.0, 23.0, 22.0, 21.0, 20.0, 19.0, 18.0, 17.0, 16.0, 15.0, 14.0, 13.0, 12.0, 11.0, 10.0, 9.0, 8.0, 7.0, 6.0, 5.0, 4.0, 3.0, 2.0, 1.0.

NMR (δ , C₅D₅N): 38.63(C-1), 26.21(C-2), 78.73(C-3), 39.78(C-4), 55.22(C-5), 18.32(C-6), 33.58(C-7), 40.11(C-8), 46.76(C-9), 38.77(C-10), 23.46(C-11), 122.30(C-12), 142.98(C-13), 43.67(C-14), 36.04(C-15), 67.52(C-16), 36.84(C-17), 44.72(C-18), 46.64(C-19), 30.73(C-20), 32.56(C-21), 27.18(C-22), 28.08(C-23), 15.52(C-24, 25), 16.65(C-26), 26.76(C-27), 70.88(C-28), 33.06(C-29), 23.87(C-30).

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$\Delta\epsilon_{253}-1.02$ (MeOH; c 0.164)表示。

10. NMR 表示为 ¹H NMR 或 ¹³C NMR, 须注明仪器的频率, 溶剂及内标物。化学位移以 δ 值(对 TMS)表示, 注明峰形, 如: 单峰(s), 宽单峰(brs), 双峰(d), 双二重峰(dd), 复峰(m)等。¹³C NMR 及 ¹H NMR 数所须注明所对应的碳和氢的位置, 采用 IUPAC 定位, 标为 C-1, C-2; H-1, H-2。例如: ¹³C NMR(21.15MHz, CDCl₃): δ 30.1(t, C-5), 74.1(d, C-6), 121.3(d, C-3), 144.2(s, C-4)。 ¹H NMR(100MHz, CDCl₃): δ 0.681(3H, s, H-18), 0.884(6H, d, J=6.0Hz, H-26 and H-27), 0.901(3H, d, J=5.0Hz, H-21), 4.342(1H, q, J_{6 α} =4.5Hz, J_{6 α} , 7 α =2.0Hz, H-6), 4.211(1H, m, W_{1/2}=18.0Hz, H-3 α)。所用仪器频率及溶剂若在实验部分的总论中已注明, 则以下皆可省略。

11. 质谱须注明所用的方法, 如(EIMS, CIMS, GC-MS, FABMS 等)及离解能, 只须给出分子离子峰及重要的特征碎片峰(相对强度), 如:EIMS(70eV m/z(%)): 386[M⁺](36), 368[M-H₂O]⁺(100), 275[M-111]⁺(35)等。高分辨质谱(HRMS)若有必要可多给一些信息。

12. 紫外光谱表示法, 如 UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm(lg ϵ): 203(4.17)。

13. 红外光谱表示法, 如 IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1740。官能团的指定放在圆括号内, 如: 1740(>C=O)。若要标明吸收带的强度, 则采用以下缩写符号: w(弱), m(中等), v(可变), s(强), vs(很强)。

14. 有机化合物和无机化合物及有关的缩写符号须规范化(参考 CA), 如氘代溶剂 CDCl₃, DMSO-d₆, D₂O, pyridine-d₅ 等。常见化学试剂在文中均以化学符号表示, 如: MeOH, EtOH, n-BuOH, PrOH, iso-PrOH, PhOH(苯酚), petrol(石油醚), CHCl₃, CCl₄, C₆C₆, Et₂O, Me₂CO, HOAc, EtOAc, THF, Ac₂O, NaOMe, CH₂N₂, HCO₂H(甲酸), TCA(三氯乙酸), TFA(三氟乙酸), NaOAc, NaOH, HCl, H₂SO₄, CO₂, H₃BO₃, NH₃, N₂ 等。

15. 制备薄层析须注明(1)薄层厚度; (2)样品的量; (3)确定带的方法; (4)从吸附剂上洗脱下化合物所用的溶剂。特殊 TLC 的吸附剂须注明, 如: AgNO₃-硅胶(1:9)。

16. 气相色谱(GC)须注明检测器(FID, EC 等), 载气及流速, 操作温度, 柱子情况等。

17. 高压液相(HPLC)须注明(1)柱子情况, 如大小、型号; (2)压力及溶剂; (3)检测方法, 如 UV 或折光率。

18. X-衍射只须给出立体结构图(最好有键长)及必要的数据, 详细记录可指明在什么地方储存。