# 溶剂萃取法从褐藻浸提液中分离提取褐藻糖胶

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摘 要:用溶剂萃取法分离提取了实际海藻浸提液中的褐藻糖胶.考察了接触时间、溶剂加入量及萃取剂浓度等对萃取的影响,并与从多糖配制液中的萃取情况进行比较.考察了无机盐水溶液反萃褐藻糖胶的性能及在溶剂中加入 TOA(三正辛胺)对萃取和反萃的影响,结果表明,季铵盐从实际鼠尾藻浸提液中萃取褐藻糖胶受溶剂加入量的影响很大,溶剂加入量越大,萃取率越低;而萃取时间对萃取的影响不大.海带浸提液的萃取优于鼠尾藻浸提液,而超声破碎法有利于萃取.TOA的加入既有利于褐藻糖胶的萃取也有利于无机盐水溶液反萃褐藻糖胶.采用溶剂萃取法制备的固体褐藻糖胶的纯度优于乙醇分步沉淀法制备的固体褐藻糖胶的纯度.

关键词:褐藻浸提液;褐藻糖胶;萃取;反萃取

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## 1 前言

褐藻属海洋藻类 ,其中所含的硫酸酯多糖—褐藻糖胶(Fucoidan)具有抗病毒、抗肿瘤、抗凝血、抗溃疡及增强和调节免疫等作用. 例如 ,从鼠尾藻中提取的硫酸酯多糖对人食管癌细胞株 Ec109、小鼠肉瘤 S180 均有抑制作用[1] ,从海带中提取的褐藻糖胶在体外可诱导白细胞介素-1(IL-1)和丙型干扰素 $(IFN-\gamma)$ 产生 ,在体内可增强 T-细胞、B-细胞和巨噬细胞等的功能[2].

传统的从褐藻中提取分离褐藻糖胶的方法一般是原料经预处理后用热水或稀酸水浸提,提取液用乙醇沉淀出粗褐藻糖胶,粗多糖再经乙醇沉淀法重结晶,从而得到产品多糖<sup>[3,4]</sup>. 乙醇沉淀法对多糖的分离选择性较差,要提高多糖的纯度需经反复多次沉淀,从而导致多糖损失大,降低了多糖的回收率.

作者用溶剂萃取法对从水溶液中提取分离褐藻糖胶进行了相关研究<sup>[5-7]</sup>. 本文报道溶剂萃取法从鼠尾藻和海带的浸提液中提取褐藻糖胶的研究结果.

## 2 实验

实验材料:鼠尾藻(*Sargassum thunbergii*)由中国科学院海洋研究所提供,采自山东青岛海域;海带(*Laminaria japonica*)由福建罗源闽港水产公司生产. 以上原料预处理后经热酸水或超声强化浸提、过滤,浸提滤液备用.

实验试剂:褐藻糖胶由中国科学院海洋研究所提供(作配制液及测定用);萃取剂 TOA(三正辛胺)为试剂级,  $N263(R_3CH_3N^+Cl^-,R=C_7\sim C_9)$ 和  $7401(R_3CH_3N^+Cl^-,R=C_8\sim C_{10})$ 为工业品,纯度皆大于95%;岩藻糖(Fucose)为 Feinbiochemica Heidelberg 产品;其它试剂和药品为市售分析纯或化学纯.

实验设备:主要有医用电动振荡器,医用离心机,电磁搅拌器,超级恒温槽,PHS-3型酸度计,722型分光光度计等.

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实验方法: 取一定体积浸提滤液和有机试剂于离心杯中,通过电动振荡或电磁搅拌使两相混合,混合时间一般为 10 min. 离心分相后,取平衡水相分析褐藻糖胶或岩藻糖.

分析方法:褐藻糖胶或岩藻糖的分析测定采用苯酚-半胱氨酸-硫酸法[8].

## 3 结果和讨论

#### 3.1 萃取时间的影响

与配制液相比,海藻浸提液的成份是非常复杂和多样的,为此首先考察了接触时间对萃取的影响,实验结果示于图 1. 由图可知,从海藻浸提液中萃取褐藻糖胶非常迅速,一般 5 min 即可达萃取平衡,与从配制液中萃取区别不大.

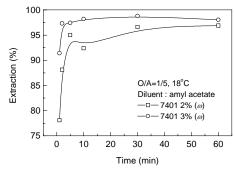


图 1 时间对从实际浸提液中萃取褐藻糖胶的影响 Fig.1 Effect of contact time on liquid—liquid extraction of fucoidan leached from *Laminaria japonica* 

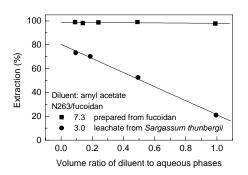


图 2 配制液与鼠尾藻浸提液中萃取的比较 Fig.2 Extraction of fucoidan as a function of volume ratio of diluent to aqueous phases

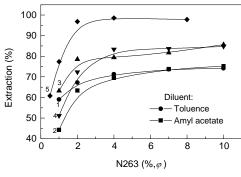
### 3.2 从浸提液和配制液中萃取褐藻糖胶的比较

研究表明<sup>[5]</sup>,从水溶液中萃取褐藻糖胶,萃取率主要取决于萃取剂对多糖的质量比,而与稀释剂的加入量基本无关. 考虑到实际浸提液与配制液的不同性质 ,考察了稀释剂加入量对实际浸提液中多糖萃取的影响 ,并与配制液的结果进行比较 .结果示于图 2. 图中配制液的多糖浓度为 1.6~g/L,并加入  $0.5~mol/L~CaCl_2$ 除去褐藻胶. 从图 2 可知,在固定萃取剂与多糖质量比的情况下,实际浸提液中多糖的萃取率随稀释剂加入量的增加几乎直线下降. N263 是强碱性萃取剂,对褐藻糖胶的萃取是通过离子间相互作用以离子交换或形成复盐的形式进行的,配制液中只要有足够的萃取剂,就能把褐藻糖胶萃取入有机相而与稀释剂的多寡关系不大;而从实际浸提液中萃取褐藻糖胶的萃取率低于从配制液中的萃取率,这首先是因为海藻浸提液中酸性杂质与褐藻糖胶竞争 N263,其次是实际浸提液中油溶性杂质较多,随着稀释剂的增加,进入有机相的杂质增多而妨碍对褐藻糖胶的萃取;此外  $CaCl_2$ 的存在也不利于萃取. 随着稀释剂的增加,萃取剂的浓度亦随之降低,这说明对实际浸提液,萃取剂的浓度效应是比较强的.

#### 3.3 从不同浸提液中萃取褐藻糖胶

图 3 为萃取剂浓度对从不同浸提液中萃取褐藻糖胶的影响. 图中曲线 1~3 所用溶液为鼠尾藻超声浸提液,其中曲线 1~2 的溶液含有 0.5~ mol/L  $CaCl_2$ ,曲线 3~ 的溶液则无  $CaCl_2$ ,显然  $CaCl_2$  的存在不利于萃取. 曲线 4~ 所用溶液为海带的热酸水浸提液,曲线 5~ 为海带超声浸提液. 比较曲线 1~ 与 2~ 可知,在低萃取剂浓度下,甲苯作为稀释剂比乙酸戊酯作为稀释剂萃取效果稍好;从褐藻原料来看,比较曲线 1~ 3~ 与 4~ , 5~ 可知,从海带中浸提的褐藻糖胶比从鼠尾藻中浸提的褐藻糖胶有更好的可萃性,这说明不同来源的褐藻糖胶在性质上可能有所不同;而从提取方法来看,通过比较

曲线 4 与 5 可知,超声强化浸提的多糖比常规热酸水浸提的多糖更容易萃取,其原因可能是由于超声浸提接触时间短,妨碍萃取的杂质较少.



- 1,2. 1.6 g/L fucoidan, leached from Sargassum thunbergii by hot acidic aqueous solutions
- 3. 0.56 g/L fucose, from *Sargassum thunbergii* by ultrasonic breakage technology
- 2.71 g/L fucoidan, from Laminaria japonica by hot acidic aqueous solutions
- 0.66 g/L fucose, from Laminaria japonica by ultrasonic breakage technology

#### 图 3 从海藻浸提液中萃取褐藻糖胶

Fig.3 Effect of N263 concentration on liquid—liquid extraction of fucoidan leached from different seaweeds by different extraction procedures

### 3.4 褐藻糖胶的反萃

从配制液中萃取褐藻糖胶,无机盐浓度是最主要的影响因素<sup>[7]</sup>,因此萃取的褐藻糖胶可用无机盐水溶液反萃回水相. 用无机盐水溶液反萃从实际海藻浸提液萃取的褐藻糖胶,结果见表 1.

表 1 从海藻浸提液中萃取褐藻糖胶及其反萃的实验结果

Table 1 Extraction of fucoidan extracted from seaweeds and the stripping by aqueous inorganic salt solutions

	Liquid-liquid extraction				Stripping				
No.	Poly-saccharide conc. in aqueous (g/L)	Extractant conc. in organic (%, ω)		Poly-saccharide conc. in organic (g/L)	Phase ratio (O/A)	Stripping agent (mol/L)	Stages	Poly-saccharide conc. in aqueous (g/L)	Stripping (%)
1	1.567	10 N263 (φ)	66.9 (1/10)	10.48	2/1	CaCl <sub>2</sub>	1 2	6.741 2.507	32.2 12.0
2	1.493	10 N263 (φ)	75.7 (1/5)	5.649	3/1	2 NaCl	1 2	6.885 1.312	40.6 7.7
					3/1	2 CaCl <sub>2</sub>	1 2	10.63 2.506	62.7 14.8
3	0.514	10 TOA (φ)	23.6 (1/2.5)	0.303	4/1	2 NaCl	1 2	1.046 0.117	86.4 9.6
4	0.537	1 (7401)	68.4 (1/5)	1.837	2/1	2 NaCl	1 2	2.642 0.259	71.9 7.1
5	0.486	1 (7401)+4.5 TOA	69.4 (1/5)	1.684	2/1	2 NaCl	1 2	2.327 0.231	69.1 6.9
		1 (7401)+9 TOA	77.3 (1/5)	1.878	2/1.25 2/1	2 NaCl	1 2	2.260 0.303	75.2 8.1
6	0.560	2 (7401)+8 TOA	79.4 (1/5)	2.223	2/1.2	2 NaCl	1 2	2.204 0.575	59.5 15.5
7	0.567	1.5(7401)+8.5 TOA	98.5 (1/2.5)	1.401	4/1	2 NaCl	1 2	4.015 0.907	71.7 16.2
8	0.570	3 (7401)	97.0 (1/5)	2.763	2/1	2 NaCl	1 2	3.914 0.467	70.8 8.5
		3 (7401)+7 TOA	97.2 (1/5)	2.768	2/1	2 NaCl	1 2	4.483 0.366	81.0 6.6
9	0.657	3 (7401)+7 TOA	97.4 (1/5)	3.200	2/1	2 NaCl	1 2	3.981 0.872	62.2 13.6
10	0.603	3 (7401)+7 TOA	97.5 (1/5)	2.945	1/1	2 NaCl	1 2	2.409 0.090	81.8 3.1
11	0.585	6 (7401)+4 TOA	95.7 (1/10)	5.595	1/1	2 NaCl	1 2	4.240 0.217	75.8 3.9

Notes: Nos.1~5 are leached from *Sargassum thunbergii* and Nos.6~11 from *Laminaria japonica*. Nos.3~6 are by hot acidic aqueous solutions and Nos.1~2 and 7~11 by ultrasonic breakage technology. Nos.1~2 are analyzed as fucoidan and Nos.3~11 as fucose, respectively.

从表可知,从海带中浸提的褐藻糖胶显然比从鼠尾藻中浸提的褐藻糖胶更易于萃取,而超声

强化浸提的多糖比常规热酸水浸提的多糖更容易萃取. 从反萃数据看,比较 No.1 与 2 可知,有机相中褐藻糖胶的浓度高不利于反萃(也可比较 No.10 与 11 的数据),而无机盐的浓度高则有利于反萃. 从 No.3 的结果可知,虽然 TOA 对褐藻糖胶的萃取不太理想,但 TOA 萃取的褐藻糖胶较易于反萃. 因此研究了混合萃取剂的萃取和反萃性能. 比较 No.4 与 5 的数据可看出 随着有机相中 TOA 浓度增加,褐藻糖胶的萃取率和反萃率都有所增加,因此 TOA 的加入既有利于褐藻糖胶的萃取,也有利于用无机盐反萃褐藻糖胶. 从反萃相比看,相比小一般有利于反萃,但不利于多糖的富集.

3.5 乙醇分步沉淀法与溶剂萃取法制备褐藻糖胶的比较

采用溶剂萃取技术从鼠尾藻浸提液中制备了褐藻糖胶,并与乙醇沉淀法制备的多糖进行了比较,其制备步骤为:

取鼠尾藻浸提液,加热浓缩至原体积的约 1/4,冷却后搅拌下加无水乙醇至  $30\%(\varphi)$ ,离心后弃去沉淀,上清液中继续加无水乙醇至  $60\%(\varphi)$ ,混合物置冰箱中过夜,离心后取沉淀,经洗涤、干燥得粗多糖,经分析含岩藻糖 19.3%.

取鼠尾藻浸提液,用含  $7401\ 5\%(\varphi)$ 的乙酸戊酯溶液以 1/5 的相比(O/A)进行萃取,萃取的多糖用  $2\ \text{mol/L}\ CaCl_2$ 溶液以 1/2 的相比(O/A)反萃  $2\ 次$ ,合并反萃液,反萃液中加无水乙醇至  $60\%(\varphi)$ ,混合物置冰箱中过夜,离心取沉淀 1,上清液中继续加乙醇至  $80\%(\varphi)$ ,放置  $3\ h$  后,离心取沉淀 2. 沉淀 1 和 2 经洗涤、干燥得到固体多糖,经分析,分别含岩藻糖 20.4%和 43.1%. 可见,萃取法所得多糖纯度高于乙醇分步沉淀法所得多糖的纯度.

# 4 结论

- (1) 用季铵盐萃取剂从实际海藻浸提液中萃取褐藻糖胶,萃取时间的影响不大,可在  $5\sim10~{
  m min}$ 内迅速达到平衡;而溶剂加入量对萃取的影响很大,溶剂加入越多,萃取率越低.
- (2) 从海带浸提液中萃取褐藻糖胶比从鼠尾藻浸提液中萃取更容易;而超声破碎法与常规热酸水浸提法相比,从前者浸提液中的萃取优于后者.
  - (3) TOA 的加入既有利干萃取也有利干反萃.
  - (4) 采用溶剂萃取法制备的褐藻糖胶的纯度可达 40%以上,优于常规乙醇沉淀法.

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Abstract: Liquid-liquid extraction of fucoidan leached from brown alga was studied in the present work. Effects of contact time, diluent volume added and extractant concentration were investigated. Results show that the liquid—liquid extraction of fucoidan from brown alga with N263 is rapid and the equilibrium between two phases can be reached in 5~10 min, and the extraction of fucoidan is significantly influenced by diluent volume added and the more the diluent volume added, the lower the percent extraction. This phenomenon is much different from the prepared solutions in which the diluent amount added has little effect on the liquid-liquid extraction. For different brown alga, the results show that the liquid-liquid extraction is more easily for fucoidan leached from Laminaria japonica than from Sargassum thunbergii, and moreover, an ultrasonic breakage technology is more favorable than hot acidic aqueous solutions for the liquid-liquid extraction. Furthermore, the stripping of fucoidan with an aqueous inorganic salt solution was investigated. The results indicated that fucoidan in the organic phase can be stripped by an aqueous inorganic salt solution, the percent stripping increases with increasing salt concentration and decreasing volume ratio of organic to aqueous phases. The effects of addition of TOA (tri-n-octyl amine) into the organic phase on the liquid-liquid extraction and stripping of fucoidan were also studied and it is shown that the addition of TOA to the organic phase is favorable to both the liquid-liquid extraction of fucoidan and the stripping with aqueous inorganic salt solutions. Finally, fucoidan was prepared from Sargassum thunbergii with liquid-liquid extraction technology and its purity (higher than 40%) is much higher than that (about 20%) with alcohol precipitation.

Key words: water extract from brown seaweeds; fucoidan; liquid-liquid extraction; stripping