

Radioimmunotherapy with ^{153}Sm -CEA monoclonal antibody in nude mice bearing human colon carcinoma: an experimental study

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Abstract: Objective To observe the therapeutic effect of ^{153}Sm -labeled CEA monoclonal antibody (mAb) in nude mice bearing human colon carcinoma. **Methods** Fifteen nude mice were subjected to subcutaneous inoculation of human colon carcinoma cells, and 3 days later, they were divided into 3 groups with equal number to receive single high-dose injection of 11.1 MBq ^{153}Sm -CEA mAb (therapy group), 11.1 MBq $^{153}\text{SmCl}_3$ (therapy control group), or 100 μl normal saline (non-treatment control group). The tumor-inhibiting effect of ^{153}Sm -labeled CEA mAb was evaluated in terms of body weight changes and tumor volume variation 4 weeks after the treatment. Histological analysis of tumors were performed in all the groups after the all other observations were completed. **Results** ^{153}Sm -CEA mAb had a significant anti-tumor effect, with a tumor inhibition rate of 74.29% at 4 weeks after treatment, while for $^{153}\text{SmCl}_3$, the inhibition rate was only 15.90%. Rapid tumor growth was observed in non-treatment control group. No significant difference in the body weight changes was noted between the 3 groups. Histopathological examination revealed tumor necrosis as the evidence for radioactive damage in therapy group, which was not observed in non-treatment control group. **Conclusions** ^{153}Sm -CEA mAb has a strong selective inhibitory effect against colon carcinoma and may be potentially used as an agent in radioimmunotherapy.

Key words: radioimmunotherapy; Samarium radioisotopes; antibodies, monoclonal; Neoplasm, nude mice

The radionuclides now in use for radioimmunotherapy (RIT) have exceeded 10 kinds, but no one single agent has merited general recognition as the best for RIT. In earlier studies iodine-131 was often adopted, but its unreliable physical property and rapid in vivo deiodination of the radiolabeled antibodies limited its practical application. Radioactive metallic nuclides such as Y-90 was subsequently chosen for RIT. Problems were not less fatal with Y-90: the radiation induced damages to the bone marrow and liver rendered it risky to administer an effective radiation dose. It has been generally agreed that the selection of radionuclides for RIT must incorporate the primary consideration of those that are capable of β particle emission, especially those emitting intermediate-energy β particles and γ rays (with γ ray energy below 300 keV) suitable for imaging without excessive radiation to cause damage to normal tissues while at the same time, applicable in in vitro

radioimmunoimaging (R域) studies.

Recently there has been increasing interest in utilization of ^{153}Sm labeling for therapeutic agents. ^{153}Sm labeled EDTMP has been used in human for the diagnosis and treatment of bone carcinoma. In 1989 Boniface first reported the radiolabeling of monoclonal antibodies (mAb) with ^{153}Sm using bifunctional chelate cyclic DTPA anhydride (cDTPAa) in the study of imaging and biodistribution in a rat model system. This preliminary study indicated the feasibility of using radioimmunoscintigraphy in combination with radioimmunotherapy in a clinical setting. Until now, however, no similar report has been available in this country addressing the application of ^{153}Sm in radioimmunoimaging and radioimmunotherapy. As a radiolanthanide, ^{153}Sm possesses excellent physical characteristics for radioimmunotherapy, capable of emitting β ray at $E_{\text{max}} = 640$ (30%), 710 (50%), and 810 (20%) keV with a half-life ($T_{1/2}$) of 1.95 days. In addition, it also emits a 103-keV γ ray that is suitable for γ camera for target absorbed dose assessment. Produced by nuclear reactor with high yield and highly specific activity initiated by

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neutron activation, ^{153}Sm is a very attractive radioisotope for RIT^[1]. We therefore selected ^{153}Sm that suits nationwide application in our investigation into the radiolabeling of anti-CEA mAb with ^{153}Sm by cDTPAa, and observed the therapeutic effect of ^{153}Sm -CEA mAb in nude mice bearing human colon carcinoma.

MATERIAL AND METHODS

Reagents

Anti-CEA mAb was obtained from Shanghai Institute of Immunology, and cyclic DTPA anhydride (cDTPAa) from Sigma Chemical Co. (St. Louis, MO). $^{153}\text{SmCl}_3$ was supplied by Department of Isotopes, China Institute of Atomic Energy. Sephadex G-50 was imported from Pharmacia Co. and divided by Factory of Shanghai Chemical Reagent. All the chemical reagents used in this study were of analytical grade, and prepared with ion-depleted water.

Tumor model

Balb/cnu/numice (female, body weight of 20g) received subcutaneous xenograft in the thigh with 5×10^6 LoVo cells. The tumors growing to approximately 1cm in diameter was cut into tiny pieces and suspended in normal saline, aspirated and injected (approximately 0.2 ml) subcutaneously into the forelimb of BALB/C nude mice (4 to 5 weeks old). The healing of the wound normally took 12h. After the tumors had grown to the volume of 0.5 to 1.0cm³, the mice were used for subsequent study of pretargeting radioimmunoimaging and biodistribution. The study of the therapeutic effect was started on the third day following tumor inoculation.

Conjugation of anti-CEA mAb with DTPA

Coupling of cyclic anhydride of DTPA (cDTPAa) with anti-CEA mAb was performed according to the method described by Hnatowich^[2]. Briefly, cDTPAa was suspended in chloroform (1mg/ml), a aliquot of which was taken with a molar ratio of DTPA:mAb at 20:1 and added into an acid-washed vial for evaporation under a stream of high-purity dry nitrogen. Anti-CEA mAb (200 μg) was then added into the vial, thoroughly shaken for 1 min and allowed at room temperature for

15-20 min for reaction, which was terminated by acetic acid. Separation of the DTPA-CEA mAb conjugate from free DTPA was achieved by a mini-Sephadex G50 chromatography. The immunoreactivity of the DTPA-CEA mAb conjugate was assessed using indirect enzyme-linked immunosorbent assay (ELISA).

^{153}Sm labeling of anti-CEA mAb

$^{153}\text{SmCl}_3$ at a dose of approximately 40MBq (with specific activity of 22.2 GBq/ml) was mixed with purified CEA mAb-DTPA conjugate (0.1 ml), and incubated at room temperature for 20 min. Paper chromatography was carried out with Xinhua No. 1 filter paper (30% ammonium nitrate-treated) as the supporter and the mixture of tributyl phosphate, butanone, and acetic ether (in a proportion of 4:10:3) as the developing agent, to determine the labeling efficiency and radiochemical purity. The immunoreactivity of the labeled mAb was tested with indirect ELISA.

In vitro stability of ^{153}Sm -DTPA-CEA mAb

Following coupling and purification, a 0.3 ml aliquot of the labeled mAb was mixed with mouse serum of the same volume at 37 $^{\circ}\text{C}$ for 24h. Sampling of the mixture was performed at 12 and 24h respectively for the determination of ^{153}Sm release rate from labeled mAb using Sephadex G-50 chromatography.

Radioimmunotherapy

Treatment with a single high dose was adopted. Fifteen tumor-bearing mice were randomly divided into 3 groups (5 in each group). The mice in group A received 11.1MBq of ^{153}Sm -DTPA-CEA mAb (100 μg) via intra-peritoneal injection, and mice in group B was given ^{153}Sm -DB₂ at the dose of 11.1MBq (100 μg) to serve as the therapeutic control group. Intraperitoneal injection with 100 μg normal saline was administered in mice in Group C as the non-treatment control group. The length (a) and width (b) of tumors were measured with a sliding caliper once a week for one month, and the tumor volume (V) was calculated according to the formula: $V = 1/6 \pi a^2 b$. All the mice were weighed on the day of injection and then once a week after it for one month. The inhibition rate (IR) of tumor growth was

calculated according to the formula: $IR = (\text{Mean tumor volume of non-treatment control group} - \text{Mean tumor volume of therapeutic group}) / \text{Mean tumor volume of non-treatment control} \times 100\%$.

Histological examination

Pathological examination was also performed in these mice after all the above observations were completed. The mice were sacrificed, their organs isolated and weighed, and then fixed in 10% formalin solution and embedded in paraffin before sections 4 μm in thickness (stained with hemalum-eosin-safran) were prepared for routine histological examination.

RESULTS

Quality control and *in vitro* stability of the labeled compounds

The labeling efficiency of ¹⁵³Sm-DTPA-CEA mAb was 56%, with a specific activity of 15.54 GBq/mol, a radiochemical purity above 95% and immunoreactivity of approximately 50%. After mixed with mouse plasma for 12 and 24 h at room temperature, the labeled mAb showed a ¹⁵³Sm-release rate of $5.47 \pm 2.64\%$ and $9.13 \pm 0.29\%$, respectively.

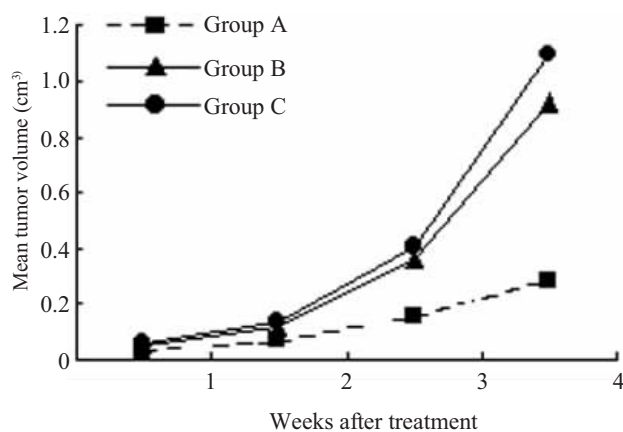


Fig.1

Fig.1 Growth curves of the implant tumors in the 3 groups

Changes in body weight of tumor-bearing nude mice

Before treatment, the body weight of the nude mice in Group C (non-treatment control) and Group B (therapeutic control) were 19.24 ± 1.49 g and 17.10 ± 1.40 g respectively, which increased to 24.60 ± 2.89 g and 19.84 ± 1.12 g respectively 4 weeks after the treatment. The body weight of the mice in Group A (therapy group) measured at the same 2 time points were 19.24 ± 1.58 g and 22.14 ± 1.23 g respectively, showing no significant body weight loss in comparison with the other 2 groups.

Dynamic observation of the tumor volume

In the first week after treatment, the tumor volume showed little difference between the groups, while in the second week slower tumor growth rate in Group A was noted. Till the fourth week, the tumor volume was significantly smaller in group A than in the other 2 groups ($P < 0.001$, Fig 1). When the tumor inhibition rate in Group C was considered to be zero, the tumor inhibition rate at 4 weeks after the therapy was as high as 74.29% in Group A, while only 15.90% in Group B, with significant difference between the latter 2 groups ($P < 0.01$, Fig 2).

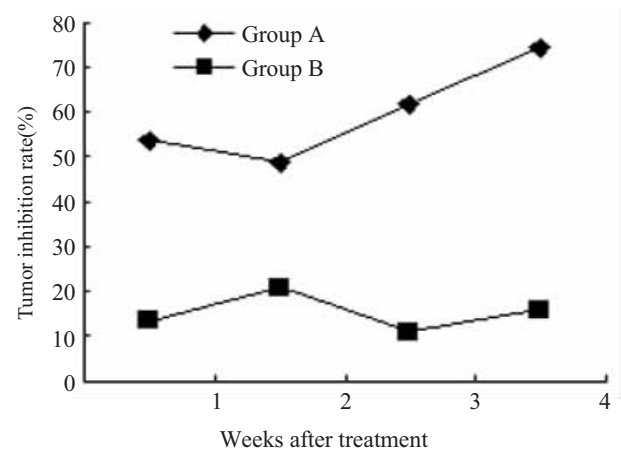


Fig.2

Fig.2 Curves for tumor growth inhibition in the 2 therapy groups

Histological examination

Samples of the tumor tissues were obtained from Group A at the end of the observation. Histopathological examination revealed evidences for degeneration

and necrosis of the tumor cells such as nucleus pycnosis and fragmentation. Some tumor tissues exhibited complete necrosis and lysis of the tumor cells to form liquefied cisternae. While the tumor cells in Group C

were characterized by absence of necrosis, with morphology typical of tumor cells.

DISCUSSION

Radiolabeled mAb against tumor-associated antigens for tumor diagnosis and therapy, ie. RIT and RIT, has attracted much attention from researchers specialized in tumor nuclear medicine, with great achievements already made therein owing much to the recent development of the mAbs with high affinity, improvement in labeling and imaging techniques, and optimized tumor microenvironment. The effect of targeting therapy depends heavily on the targeting ability of the carriers that delivers the irradiation damage by radionuclide to the target. In hundreds of the anti-tumor mAb already developed, Anti-CEA mAb merits special attention and has already entered clinical applications, for instance, the determination of CEA levels in serum and body fluid, immunohistochemical staining, tumor radioimmunoimaging, radioimmuno-guided surgery and radioimmunotherapy. RII and RIT with CEA mAb are of important significance for early diagnosis of tumors (such as colorectal carcinoma), and are also useful in the detection and therapy of tumor recurrence and metastasis. In the United States, anti-CEA mAb is the first approved anti-tumor antibody for clinical application. Studies of experimental and clinical application of anti-CEA mAb in RII for various types of tumors such as colorectal carcinoma have been conducted in China, and have verified the value of the antibodies in the diagnoses and treatment of CEA-positive tumors.

This present paper, for the first time in China, described the radiolabeling of CEA mAb with ^{153}Sm using the cDTPAa, along with the experimental study of radioimmuno-therapy in nude mice bearing human colon carcinoma. The results showed that ^{153}Sm -CEA mAb at the dose of 11.1 MBq had obvious inhibitory effect on colon carcinoma xenografted in nude mice. Tumor growth inhibition was observed 2 weeks after the therapy, and at the fourth week the tumor inhibition

rate reached 74.29%. The mechanism underlying the strong selective inhibitory effect of ^{153}Sm -CEA mAb against colon carcinoma may be that (1) α ray emitted by radionuclide may result in irreversible damage to cell genetic materials and DNA by means of direct and indirect ionizing radiation; (2) the labeled antibodies may specifically bind to the tumor cells through direct contact, permeation, or localization by gravitational force bond; (3) α ray produced by ^{153}Sm with proper strength of penetration may kill those tumor cells that are at a distance from the targeting site, antigen-negative tumor cells in the neighborhood, and those hard to reach through permeation by mAb in RIT.

RIT is not ideal, however, for treating massive solid tumors. The investigations have shown a negative correlation between the curative effect of RIT for solid tumors and the tumor bulk. The principal reason lies in the fact that the antibody uptake by the tumors may be affected by many factors such as increased interstitial pressure within the tumor, relatively decreased number of the blood capillaries and possible necrosis present in the increased tumor bulks. RIT is therefore considered particularly suited for treating sub-clinical microfocal recurrent tumors arising from previous tumor remnant or postoperative metastasis. At the same time, the α ray that ^{153}Sm emits with intermediate or low-energy is effective to kill tiny focal tumors. Based on the above consideration, we conducted our experiment with RIT, which was designed to initiate on the third day following tumor inoculation. Histopathological evidence revealed degeneration and necrosis of the tumor cells, which was rather extensive in some tumor tissues, suggesting the efficacy of ^{153}Sm -CEA mAb on tumors in early stages of development, when less tumor cells, lower heterogeneity of the tumor antigen expression and higher sensitivity to radioactivity are the major features. In addition, $^{153}\text{SmCl}_3$ itself has, to a certain degree, tumor-inhibiting effect at a rate of 15.90% at the fourth week of treatment, which may be attributed to unspecific radiation effect of this agent. But since $^{153}\text{SmCl}_3$ did not conjugate with mAb, effective in vivo

localization of tumor cells would not take place, and this unspecific radiation effect was understandably limited.

Based on literature review and the results of this investigation, we conclude that RIT with ¹⁵³Sm-CEA mAb may serve as an auxiliary method for tumor treatment, which is particularly suitable for small tumor or metastasis and can be useful in the prevention of tumor recurrence. With its considerable value in the diagnosis and therapy of colon carcinoma, ¹⁵³Sm-CEA mAb may become a new type of targeting therapy agent for RIT, already showing its potential in clinical applications.

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¹⁵³Sm-CEA 单抗对结肠癌移植瘤裸鼠模型放射免疫治疗的实验研究

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摘要 目的 观察 ¹⁵³Sm 标记抗 CEA 单抗 ¹⁵³Sm-CEAmAb 对肿瘤的抑制效应。探索 ¹⁵³Sm-CEAmAb 在荷人结肠癌移植瘤裸鼠模型中的放射免疫治疗作用。方法 体外培养的 LoVo 细胞株无菌接种于 15 只裸鼠前肢皮下。制备荷人结肠癌移植瘤裸鼠模型并分为 3 组。每组 5 只。在移植瘤接种后的第 3 天实施治疗。采用单次较大剂量治疗。渊 1.1 MBq/ 每鼠。袁治疗组注射 ¹⁵³Sm 标记 TPA 环酞法 冤抗 CEA 单抗。单纯注射 ¹⁵³SmCl₃ 的荷瘤裸鼠作为治疗对照组。袁治疗对照组注射 100 滋生理盐水。遥给药后定期测量裸鼠的体质量和肿瘤生长体积并进行肿瘤的组织病理学观察。遥结果 ¹⁵³Sm-CEAmAb 治疗组对肿瘤有明显的抑制作用。袁治疗第 4 周肿瘤生长抑制率达到 74.29%。袁而非治疗对照组肿瘤体积增长迅速。遥治疗对照组第 4 周肿瘤生长抑制率为 15.90%。袁与治疗组相比有显著性差异。渊 < 0.05。冤。遥治疗组与对照组相比。袁动物体质量没有显著性差异。遥组织病理学结果提示 ¹⁵³Sm-CEAmAb 治疗组肿瘤组织有坏死改变。袁而对对照组肿瘤细胞呈典型的癌细胞形态。遥结论 ¹⁵³Sm-CEAmAb 对人结肠癌的荷瘤裸鼠具有抑制增长作用。遥 ¹⁵³Sm-CEAmAb 作为一种新型核素的导向治疗剂。袁具有定位诊断和导向治疗结肠癌的双重作用。袁具有良好的应用前景。遥

关键词 放射免疫疗法 放射核素 抗体 袁克隆 肿瘤 袁裸鼠