# 简报 Short Communications

# The roles of carboxylesterase and AChE insensitivity in malathion resistance development in brown planthopper

# LIU Ze-Wen, HAN Zhao-Jun\*

(Key Laboratory of Monitoring and Management of Plant Diseases and Insects, the Ministry of Agriculture, Nanjing Agricultural University, Nanjing 210095, China)

Abstract: Malathion resistance of brown planthopper (Nilaparvata lugens Stål) was selected in laboratory and the successive changes in carboxylesterase (CarE) and AChE were also analyzed. The results showed that the speed of resistance selection varied with generations. The biggest change of LD<sub>50</sub> occurred between the 3rd – 5th generations. The rise of carboxylesterase activity was correlated with the change in malathion resistance for the first 5 generations, while change in AChE sensitivity between 6th – 8th generations had a higher correlation with the resistance development. Therefore, it was concluded that carboxylesterase activity increase play important role in the early stage of the resistance development and the AChE insensitivity at the late stage.

Key words: malathion; brown planthopper; carboxylesterase; acetylcholinesterase

# 1 Introduction

The brown planthopper, Nilaparvata lugens Stål (BPH), is a major rice pest in many parts of Asia. Extensive use of insecticides has selected for resistance in populations of this pest from different countries and areas (Nagata et al., 1979; Nagata, 1982; Gao et al., 1987). In order to manage resistance, the resistance mechanism had been studied. Esterases played an important role in the resistance of BPH to organophosphorus (OP) insecticides. Ozaki (1969) first reported that OP resistant BPH had higher carboxylesterase activity. Thereafter several papers reported similar results (Hama and Hosoda, 1983; Sun et al., 1984; Tranter and Emden, 1984; Kim and Hwang, 1987; Park and Choi, 1991). The insensitivity of acetylcholinesterase (AChE) may also be important in the OP resistance (Hama and Hosoda, 1983; Park and Emden, 1991). In order to confirm these results and find the roles of different mechanisms in resistance development, we selected BPH with malathion in laboratory and checked the changes in LD50, esterase and AChE at each generation.

## 2 Materials and Methods

## 2.1 Insect

The brown planthopper, Nilaparvata lugens, used in this study was first collected from the experimental field of Jiangsu, Nanjing. Then was reared on rice seedlings in laboratory, at  $25 \pm 1^{\circ}\text{C}$ , 16L/8D.

# 2.2 Chemicals

Malathion (99.9%) used in the topical treatment and biochemical analysis was provided by Professor Toru Nagata (Ibaraki University, Japan). Malathion (90%) used for resistance selection was provided by Shanghai Pesticide Company. Malaoxon (62.8%) was provided by Jiangsu Pesticide Research Institute. α-Naphthyl acetate (α-NA) was purchased from Shanghai First Chemical Company, fast blue RR salt and acetylcholine iodide (ACHI) from Fluka Chemical Company; DTNB, 5,5'-dithiobis-2-nitrobenzoic acid, from Huamei Biological Engineering Company.

### 2.3 Resistance selection

Resistance was selected by spraying insecticides on

基金项目:"973"国家重点基础研究项目(J20000162);江苏省科技攻关计划(BE2001345)

作者简介: 刘泽文, 男, 1977年11月生, 在读博士生, E-mail: jemunson@njau.edu.cn

<sup>\*</sup> 通讯作者 Author for correspondence, E-mail: zjhan@njau.edu.cn

seedlings infested with BPH. The seedling in soilless cultured were placed in the selection cage (28 cm × 28 cm × 43 cm), then 100 - 200 3rd instar larvae were placed in the cage. 2 hours later, the insecticide about LC70 in doses was sprayed on seedlings with insects using the pocket sprayer (purchased from Hongxing Company, Zhejiang Province). Then the cage was placed in observing room, at  $25 \pm 1$  °C , 16L/8D. More than 2 000 larvae were treated for each generation.

### 2.4 Bioassay

2期

The bioassay followed the micro topical application technique reported by Nagata (1982). Macropterous adult females of 3 - 5 day-old were used as test animals in this study. A droplet of 0.04 µL acetone solution of insecticides was applied topically to the dorsal surface of the thorax of each female adult that had been anesthetized with carbon dioxide using a hand microapplicator (Burkard Manufacturing Co. Ltd, Richmansworth England). Thirty insects were treated at each concentration, and every treatment was repeated 3 times. Controls used acetone alone instead of insecticide solution. The treated insects were reared on the seedlings soilless cultured in the rearing cage (50 cm  $\times$  38 cm  $\times$  80 cm), at 25  $\pm$  1°C, 16L/ 8D. The results were checked in 24 hours.

#### 2.5 Determination of carboxylesterase activity

Ten 3rd instar larvae were homogenized in a glass homogenizer with 1 000 µL of 0.02 mol/L phosphate buffer (pH 7.0) using the method of Hung et al. (1990) adapted for use in a microplate reader. The homogenate was centrifuged at 4 000 x g and 4°C for 30 min, and the supernatant was used as the source of the carboxylesterase. In a well of the microplate, 100  $\mu$ L of the supernatant was put in, followed by addition of 100 µL of mixed solution of 2 mmol/L  $\alpha$ -naphthyl acetate and 1.5 mmol/L Fast Blue RR Salt (containing 10 5 mol/L eserine). Then the carboxylesterase activity was measured at 450 nm on the Microplate Reader (MODEL 550, BIO-RAD). Five replications were made for each generation.

### 2.6 Determination of the $I_{50}$ of malaoxon to AChE

Ten 3rd instar larvae were homogenized in a glass homogenizer with 2 mL of 0.02 mol/L phosphate buffer (containing 0.1% Triton X-100, pH 7.0) using the method of Park and Choi (1991) adapted for use in a microplate reader. The homogenate was centrifuged at

 $10\ 000 \times g$  and 4% for 30 min, and the supernatant was used as the source of AChE. Six concentrations of malaoxon solution were made by diluting malaoxon using acetone. 5 µL of insecticide solution of each concentration was mixed with 95 µL of the solution of AChE, and the mixture was placed in wells of microplate for 1 hour. The control was set by using phosphate buffer instead of insecticide solution. Then the 100  $\mu$ L DTNB (300  $\mu$ mol/L) and 100  $\mu$ L ATCHI (1.5 mmol/L) were added successively. The residual activity of AChE was measured at 405 nm on the Microplate Reader.

#### 3 Results

# The change of LD<sub>50</sub> of malathion against BPH and CarE activity after resistance selection

The topical LD<sub>50</sub> values of malathion against BPH were given in Fig. 1. The LD<sub>so</sub> of F<sub>0</sub> (the field population unselected) was 0.111  $\mu g$ /female, which is 10.07 times of S  $^{(}$  the susceptible strains in laboratory $^{()}$ , which shows that the field population was already with low level of resistance to malathion. From  $F_1$  to  $F_8$ , we can find that the change of LD<sub>50</sub> between two successive generations was different. The change of  $LD_{50}$  from  $F_0$  to  $F_2$  was small, and the  $LD_{50}$  of  $F_2$  was 0.301  $\mu g/female$ . The change from F<sub>3</sub> to F<sub>5</sub> was the biggest among the all generations selected, and the LD<sub>50</sub> of  $F_5$  reached 1.69  $\mu g/fe$ male. But the change from  $F_6$  to  $F_8$  became small again, with the LD<sub>50</sub> of 1.81  $\mu$ g/female to 2.03  $\mu$ g/female.

Fig. 1 also showed that carboxylesterase activity increased in successive generations after selection just like the change of  $\mathrm{LD}_{50}$ . The correlation index between carboxylesterase activity and LD<sub>50</sub> of each generation reached 0.9929. But from  $F_6$  to  $F_8$  it was only 0.8798. These means that carboxylesterase played an important role in resistance development at the early stage.

# 3.2 The change of $I_{50}$ of malaoxon against AChE after resistance selection

Fig. 2 showed that the biggest change of I<sub>50</sub> of malaoxon to AChE did not occur during the generations at which the biggest change of LD<sub>50</sub> occurred. There was not markable change of  $I_{50}$  from  $F_0$  to  $F_3$ , and the change was

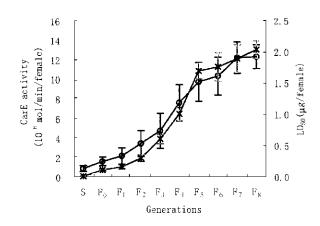


Fig. 1 The change of LD<sub>50</sub> of malathion against BPH and CarE activity after resistance selection

a little bigger from  $F_4$  to  $F_5$  than that from  $S_0$  to  $F_3$ . The biggest change of  $I_{50}$  occurred from  $F_6$  to  $F_8$ . The correlation index between  $I_{50}$  and  $LD_{50}$  was 0.8426, and from  $F_5$  to  $F_8$  reached 0.9954. These means the AChE insensitivity play much more important roles in resistance development at the later stage.

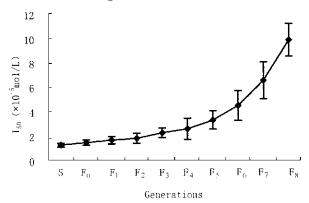


Fig. 2 The change of I<sub>50</sub> of malaoxon against AChE after resistance selection

Table 1 Three kinetic parameters in  $S_7$   $F_0$  and  $F_8$ 

Parameter	S	F <sub>0</sub>	F <sub>8</sub>
$K_{\rm m}$ (10 <sup>-6</sup> mol • mg - 1 • min - 1	) 5.178 ± 0.746 a	$6.242 \pm 1.357$ ab	$8.013 \pm 2.146 \text{ b}$
$V_{\text{max}}(\text{mmol} \cdot \text{L}^{-1})$	$1.222 \pm 0.104$ a	1.340 ± 0.366 a	$1.607 \pm 0.421$ a
$K_i$ (mol·min-1)	1.005 ± 0.122 a	1.118 ± 0.205 a	$1.694 \pm 0.405$ b

Notes: Different letters in the same horizontal column showed the significant difference at  $0.05 \ \mathrm{level}$ .

Table 1 showed that there were significant differences between S and  $F_8$  in  $K_m$  and  $K_i$  and between  $F_0$  and  $F_8$  in  $K_i$ . The significantly different  $K_m$  between S and  $F_8$  showed that the affinity of AChE in  $F_8$  declined significantly.  $K_i$  is one of the parameters for determining the in-

hibition effect of inhibitors against AChE and the AChE insensitivity. The results showed that the AChE insensitivity in  $F_8$  was significantly higher than that of S and  $F_0$ , which indicated that AChE insensitivity might be one of the important mechanisms for malathion resistance in BPH.

## 4 Discussion

Resistance selection with malathion showed that along with the increase of resistance of BPH, the carboxylesterase activity in the selected strain became higher and the AChE became more insensitive. These results, similar as previous reports, confirmed that both carboxylesterase activity increase and acetylcholinesterase insensitivity played important roles in the OP insecticide resistance of BPH (Hama and Hosoda, 1983).

From the related changes between LD<sub>50</sub> and carboxy-lesterase activity or AChE insensitivity in the successive selection generations, it was also found that these two mechanisms played different roles in the resistance development procession. At the early stage, the increase of the carboxylesterase activity was the main cause for resistance quick rise. At the late stage, it was the AChE insensitivity for resistance further growing. These suggested that the metabolizing resistance came first and the target resistance followed in the resistance development. This resistance development mode should be considered in rational pest resistance management.

Acknowledgements We would like to acknowledge the National Key Basic Research Program of China (973 Program, Grant No. J20000162). We also would like to acknowledge Professor Toru Nagata in Ibaraki University of Japan for his provision of the easy equipments for rearing the brown planthopper and Hand Microapplicator.

#### References

Gao H H, Wang Y C, Tan F J, You Z P, 1987. Studies on the sensitivity-level of the brown planthopper, *Nilaparvata lugens* Stål, to insecticides. *Journal of Nanjing Agricultural University*, 4 (Suppl.): 65 – 71. [高辉华, 王荫长, 谭福杰, 尤子平, 1987. 稻褐飞虱对杀虫剂敏感水平的研究. 南京农业大学学报, 4 (增刊): 65 – 71]

Hama H, Hosoda A, 1983. High aliesterase activity and low acetylcholinesterase sensitivity involved in organophosphorus and car-

- bamate resistance of the brown planthopper, Nilaparvata lugens Stål (Homoptera: Delphacidae). Applied Entomology and Zoology, 18 (4): 475 485.
- Hung C F, Kao C H, Liu C C, Lin J G, Sun C N, 1990. Detoxifying enzymes of selected insect species with chewing and sucking habits. *Journal of Economic Entomology*, 83 (2): 361 365.
- Kim J W, Hwang T C, 1987. Studies on resistance to organophosphorus insecticides in the brown planthopper, Nilaparvata lugens Stål ( □ ). Difference of the biochemical characteristic. Korean Journal of Plant Protection, 26 (3): 165 170.
- Nagata T, 1982. Insecticide resistance and chemical control of the rice planthopper, Nilaparvata lugens Stål. Bulletin of the Kyushu National Agriculture Experiment Station, 22 (1): 49 164.
- Nagata T, Masuda T, Moriya S, 1979. Development of insecticide resistance in the brown planthopper, Nilaparvata lugens (Stål)

- (Homoptera: Delphacidae). Applied Entomology and Zoology 14 (3): 264 269.
- Ozaki K, 1969. The resistance to organophosphorus of the green rice leafhopper, *Nephotettix cincticeps*, and the smaller brown planthopper. *Review of Plant Protection Research*, 2: 1 13.
- Park H M, Choi S Y, 1991. Changes in esterase activity and acetylcholinesterase sensitivity of insecticide-selected strains of the brown planthopper (Nilaparvata lugens Stål). Korean Journal of Plant Protection, 30 (2): 117 123.
- Sun C N, Chung T C, Dai S M, 1984. Insecticide resistance in the brown planthopper, *Nilaparvata lugens* Stål (Homoptera: Delphacidae). *Protection Ecology*, 7 (2/3): 167 181.
- Tranter B C, Emden H F, 1984. Esterase variation in the brown planthopper *Nilaparvata lugens* and its involvement in resistance to organophosphorus insecticides. *British Crop Production Conference*, 2: 521 526.

# 羧酸酯酶和乙酰胆碱酯酶在褐飞虱 对马拉硫磷抗性发展中的作用

刘泽文, 韩召军

(南京农业大学农业部病虫监测与治理基础实验室,南京 210095)

摘要:对室内筛选褐飞虱 Nilaparvata lugens Stål 对马拉硫磷抗性及羧酸酯酶、乙酰胆碱酯酶的连续变化进行了研究。结果表明,抗性发展在不同世代之间存在一定的变化。LD<sub>50</sub>的最大变化发生在第3代和第5代之间。在筛选的前5代,羧酸酯酶活性上升与马拉硫磷抗性变化存在很好的相关性,而乙酰胆碱酯酶敏感性在第6代和第8代间的变化与抗性发展存在很好的相关性。可见,羧酸酯酶活性的上升在抗性发展的早期阶段起重要作用,而乙酰胆碱酯酶不敏感在抗性发展的后期阶段起更要作用。

关键词: 褐飞虱; 羧酸酯酶; 乙酰胆碱酯酶

中图分类号: 0965.9 文献标识码: A 文章标号: 0454-6296(2003)02-0250-04