

## 小球藻细胞活性物质的提取及对啤酒酵母的生理效应\*

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**Extraction of active substance from *Chlorella vulgaris* cells and its physiological effects on *Saccharomyces cerevisiae*.** HU Kaihui, ZHOU Shanyong (School of Life Sciences, Fujian Agriculture and Forestry University, Fuzhou 350002, China). -Chin. J. Appl. Ecol., 2005, 16(8): 1573~1576.

The study showed that freezing-thawing method could have a higher cellwall breakage rate, and a light injury to the active substance, which could not only increase the cell fission and growth, delay the death of *Saccharomyxete* cells, but also enhance CO<sub>2</sub> production rate and quicken fermentation process when the cells were exposed to the medium with 0.5% active substance.

**Key words** *Chlorella vulgaris*, Active substance, Extraction, *Saccharomyxete*, Physiological effects.

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### 1 引言

小球藻(*Chlorella vulgaris*)为绿藻门普生性单细胞藻类,因其具有极为丰富均衡的营养成分,常作为医疗保健品<sup>[3,5,28]</sup>,或用于食品、轻工业以及饲料添加剂,并可用于重金属的生物吸附<sup>[7,13,16,21,23]</sup>、污水处理<sup>[10,20,24]</sup>及环境评价<sup>[17~19,27,29]</sup>等方面。小球藻生长因子(Chlorella Growth Factor, CGF)又称小球藻精,为其细胞活性物质,含有氨基酸、核酸、多糖、多肽、蛋白质、酶、维生素、矿物质等成分<sup>[3]</sup>,被誉为“类荷尔蒙”<sup>[11]</sup>。国外对其医疗功能有较多的研究,其药理作用主要有激活淋巴细胞、增强人体免疫能力、活化人体细胞、加快儿童生长发育、抵抗外来疾病入侵和促进人体受伤组织修复等,CGF对有机物、重金属中毒具有迅速康合作用,还能防治胃溃疡、高血压和心血管等疾病<sup>[2,12]</sup>。近来,小球藻生长因子提取技术及其生理活性研究成为英、美、日等国家和我国台湾地区的研究热点<sup>[4,26,30]</sup>。本文在比较几种小球藻破壁方法的基础上,研究其活性提取物对啤酒酵母(*Saccharomyxete cerevisiae*)的生理效应,以期为今后将其应用于酵母工业、提高酵母菌产率、缩短生产周期及降低生产成本等提供理论依据。

### 2 材料与方

#### 2.1 材料

以粉核小球藻(*Chlorella vulgaris*)(福建莆田神州生物公司提供)、啤酒酵母(*Saccharomyxete cerevisiae*)(本校微生物实验室保存)为供试材料。粉核小球藻培养基为NH<sub>4</sub>Cl 0.475 g, KH<sub>2</sub>PO<sub>4</sub> 0.136 g, CaCl<sub>2</sub> 0.02 g, EDTA 0.001 g, Mg-SO<sub>4</sub> 0.024 g, Na<sub>2</sub>CO<sub>3</sub> 0.002 g,加水至1 000 ml,调pH7.0, 121℃高压灭菌30 min。啤酒酵母采用麦芽汁培养基:称取500 g麦芽粉,加水至2 000 ml,于水浴锅内65℃糖化2 h,双层纱布过滤,滤液煮沸30 min,4层纱布过滤,用糖度计测

定外观浓度为12 Bx,分装,121℃湿热灭菌30 min备用。崩解液:NaCl 0.9 g, Na<sub>2</sub>HPO<sub>4</sub>·2H<sub>2</sub>O 2.61 g, 柠檬酸·H<sub>2</sub>O 0.36 g,加水至100 ml,121℃高压灭菌30 min。

#### 2.2 小球藻细胞活性物质的提取

**2.2.1 纯种培养与收集** 按1:10接种小球藻,28℃光照培养1周,长至浓绿,无菌条件下取500 ml摇匀,离心10 min(4 000 r·min<sup>-1</sup>),弃上清液,沉淀用无菌水离心洗涤3次,得纯净无菌藻泥。

**2.2.2 破壁方法** 1)机械匀浆破壁法:用无菌水将无菌藻泥稀释,使小球藻的密度达到1 010 CFU·ml<sup>-1</sup>,无菌条件下用高速组织捣碎机匀浆30 s(12 000 r·min<sup>-1</sup>),3次重复,取样,显微镜下用血球计数板计数,统计完整细胞数;2)机械研磨破壁法:将无菌藻泥在研钵中加少量石英砂研磨30 min;3)超声波破壁法:用超声波破碎仪进行处理;4)冻融破壁法:用崩解液将无菌藻泥稀释,将样品置4℃冰箱预冻2 h,再置-22℃冰箱冷冻过夜,次日取出,加水融化,融化至一半时剧烈振摇30 min,再放回-22℃冰箱冷冻过夜,重复12次。

**2.2.3 细胞活性物质的提取** 将破壁后的小球藻转移至无菌离心管中,8 000 r·min<sup>-1</sup>离心15 min,上清液移至无菌透析袋透析除盐,得细胞活性物质浓缩液。用1 ml无菌注射器以0.5 ml·支<sup>-1</sup>分装至2 ml安培瓶中,冷冻干燥无菌封口,得细胞活性物质冻干品(图1)。

#### 2.3 啤酒酵母生长测定

**2.3.1 泡沫变化** 取7支装有9 ml麦芽汁培养基的试管,每支分别接种1 ml啤酒酵母菌液,按2.0%、1.0%、0.5%、0.25%、0.1%、0.05%加入细胞活性物质冻干品,摇匀,28℃恒温箱培养48 h,观察泡沫高度变化,3次重复。

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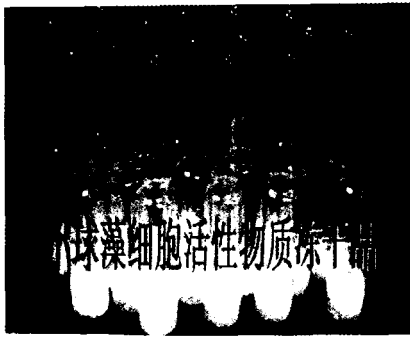


图1 小球藻活性物质冻干品

Fig.1 Products of active substance from *Chlorella vulgaris* by freezing-dry method.

**2.3.2 CO<sub>2</sub> 失重测定** 取6瓶装有100 ml 麦芽汁培养基的三角瓶, 每瓶分别接种1 ml 啤酒酵母菌液, 取1瓶作为对照, 另5瓶分别加入细胞活性物质1、0.5、0.25、0.1和0.05 g, 摇匀, 28℃恒温箱培养, 每6 h 称重一次, 直至恒重, 3次重复。

**2.3.3 菌体量测定** 取3支装有9 ml 麦芽汁培养基的试管, 每支分别接种1 ml 啤酒酵母菌液, 取1支作为对照, 另2支分别加入0.1和0.05 g 的细胞活性物质冻干品, 摇匀, 28℃恒温箱培养, 于24和48 h 取样计数, 3次重复。

**2.3.4 酵母菌死亡率测定** 取3支装有9 ml 麦芽汁培养基的试管, 每支分别接种1 ml 啤酒酵母菌液, 取1支作为对照, 另2支分别加入0.1和0.05 g 的细胞活性物质冻干品, 摇匀, 28℃恒温箱培养, 发酵7 d 后, 置4℃冰箱保存, 于30、60和90 d 取样制片(加次甲基蓝染色), 并用血球计数板统计染色酵母菌数, 计算酵母死亡率, 3次重复。啤酒酵母菌死亡率 = 染色酵母菌数/总菌数 × 100%。

**2.3.5 比生长率计算**  $U = (\ln N_{t_2} - \ln N_{t_1}) / (t_2 - t_1)$

## 2.4 数据处理

实验数据采用 SPSS 软件分析和处理<sup>[6, 14, 22]</sup>。

## 3 结果与分析

### 3.1 不同破壁方法的效果比较

小球藻体积小, 细胞壁坚固, 破壁较难, 破壁成为提取活性成分的关键。从表1可以看出, 超声波法、冻融法与机械匀浆法、机械研磨法相比差异达到极显著水平, 机械匀浆法与机械研磨法差异不显著。可见, 以冻融法破壁效果最佳(图2)。

表1 不同小球藻破壁方法的效果

Table 1 Effect of different methods to break cell wall

方法 Methods	匀浆法 Homogenate	研磨法 Grinding	超声波法 Ultrasonic wave	冻融法 Freeze-thawing
破壁率(%) Broken rates	11A	6A	30B	90C

破壁率 = (细胞总数 - 完整细胞数) / 细胞总数 × 100% Broken rates = (Total number of cell - complete cell number) / total number of cell × 100%; 表中数据为3次平均值 Data in the table were the average values of three replications; 不同字母表示差异显著 ( $P < 0.01$ ) Different letters represent significantly different at 0.01 level. 下同 The same below.

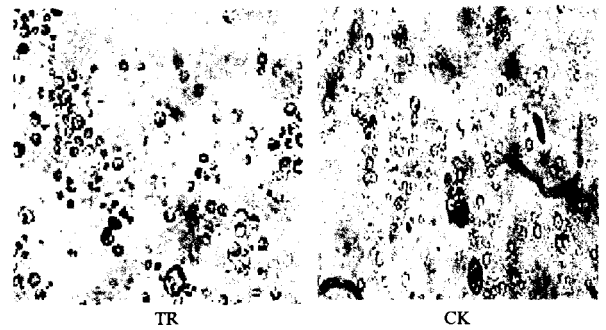


图2 冻融法的破壁效果

Fig.2 Effect of freeze-thawing method on *C. vulgaris* cell wall.

### 3.2 细胞活性物质对发酵过程酵母菌数的影响

从表2可以看出, 不同浓度小球藻活性物质对酵母的生长具有显著影响, 其比生长速率明显高于对照。在培养基中添加0.5%的提取物, 发酵24 h 后啤酒酵母菌数是对照的1.8倍, 48 h 后是对照的6.8倍。说明细胞活性物质具有提高酵母菌细胞分裂速度和促进生长的作用(图3)。

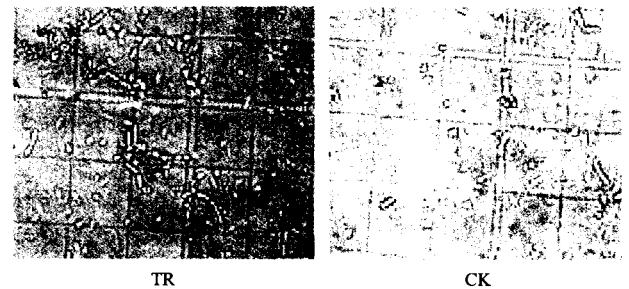


图3 细胞活性提取物对酵母生长的影响

Fig.3 Effects of adding active substance from *C. vulgaris* on the growth of *S. cerevisiae*.

表2 细胞活性物质对发酵过程啤酒酵母菌数的影响

Table 2 Effect of active substance from cells on the amount of *S. cerevisiae* in fermentation process

项目 Treatments	比生长速率 Specific growth rate ( $u \cdot d^{-1}$ )		细胞数 Numbers ( $10^5 CFU \cdot ml^{-1}$ )		
	0~24 h	24~48 h	0 h	24 h	48 h
CK	2.23	2.78	0.12	1.12	18
0.25%	2.44	4.21	0.12	1.38	93
0.5%	2.72	4.22	0.12	1.83	125

### 3.3 细胞活性物质对啤酒酵母发酵过程中产生泡沫的影响

从图4和图5可以看出, 细胞活性物质具有促进啤酒酵母的发酵、提高啤酒酵母产气量的作用, 且泡沫高度随活性物质添加量的增加而增加。当添加量达到0.5%以上时, 增加量趋于平缓。

### 3.4 细胞活性物质对啤酒酵母发酵过程中CO<sub>2</sub>失重影响

由图6可见, 在加入细胞活性物质的培养基中, 啤酒酵母发酵产生的CO<sub>2</sub>与对照不同, 并且随着添加量的增加, CO<sub>2</sub>失重增加。当添加量达到0.5%以上时, 失重增加量趋于平缓, 表明适量的小球藻活性提取物有利于提高酵母对糖类物质的利用和产酒精能力。



图4 细胞活性物质对啤酒酵母发酵过程中产生泡沫的影响  
Fig.4 Effect of active substance from *C. vulgaris* on foamy height in the fermentation process of *S. cerevisiae*.

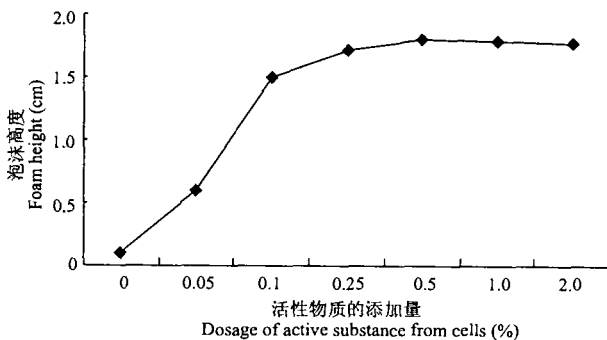


图5 泡沫高度与细胞活性物质添加量的关系  
Fig.5 Relationship between foamy height and active substance dosage.

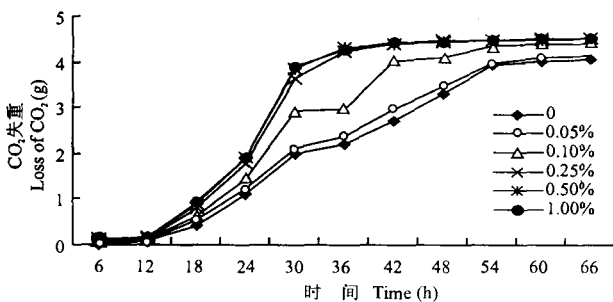


图6 啤酒酵母发酵过程中 CO<sub>2</sub> 失重与细胞活性物质添加量的关系  
Fig.6 Relationship between the loss of CO<sub>2</sub> and active substance dosage in the fermentation process of *S. cerevisiae*.

### 3.5 细胞活性物质对啤酒酵母菌死亡率的影响

从表3可以看出,加入0.25%和0.5%的细胞活性物质冻干品,酵母菌30 d后的死亡率分别比对照减少34%和50%,60 d后分别比对照减少20%和34%,90 d后分别比对照减少13%和27%,表明小球藻活性提取物能延长该酵母的保存期。

表3 细胞活性物质对啤酒酵母菌死亡率的影响  
Table 3 Effect of active substance from cells on the mortality rate of *S. cerevisiae*

处理 Treatments	酵母菌死亡率 Mortality rate of <i>S. cerevisiae</i> (%)		
	30 d	60 d	90 d
CK	21	30	45
0.25%	14	24	39
0.5%	10	21	33

## 4 讨 论

目前,小球藻的破壁提取主要采用机械匀浆法、机械研磨法、超声波法、酶解法以及酸碱浸提法等。酶解法效果较好,但成本较高,难以在生产上推广应用。机械匀浆法、机械研磨法、超声波法等由于小球藻的细胞壁为坚固的纤维素,且体积微小,破壁率低,不能充分利用。酸碱浸提法产率较高,但活性物质易被破坏。通过对小球藻几种破壁方法的比较发现,细胞冻融法破壁率较高,且能最大程度地保持小球藻活性成分的活性,活性物质损伤少,提取工艺简单。冻融法破壁率高的主要原因可能是反复冻融小球藻使其细胞壁结为冰晶,而后又解冻,连续处理多次使小球藻细胞壁被拉伤,通透性增加,细胞较易破裂<sup>[15,25]</sup>。

小球藻细胞活性物质含有氨基酸、多糖、蛋白质、维生素等功能成分,具有活化细胞,促进细胞生长和分裂作用<sup>[1,8,9,25,31]</sup>。对小球藻细胞活性物质对啤酒酵母生理效应的研究表明,小球藻细胞活性物质可显著增加啤酒酵母发酵过程中的酵母菌数,促进啤酒酵母的生长和繁殖,加速酵母发酵进程,增加酵母发酵的泡沫高度、产气量,缩短酵母菌发酵的延滞期,快速启动发酵,提前到达产气高峰。同时,小球藻提取物能提高酵母发酵过程抗杂菌能力,且细胞活性物质可以使酵母菌的降糖更彻底、糖的利用率更高。此外,添加小球藻细胞活性物质还可延缓酵母菌的死亡时间,在酵母工业、酿酒工业生产上具有良好的应用前景。今后还需进一步对小球藻活性成分进行分离鉴定,并深入探讨其活性成分对酵母菌的生理效应及其作用机理。

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