

## Getting bacteria to do a plant's job

Researchers engineer microbes for low-cost production of precursor of anticancer drug Taxol and other pharmaceuticals.

Anne Trafton, MIT News Office

October 1, 2010

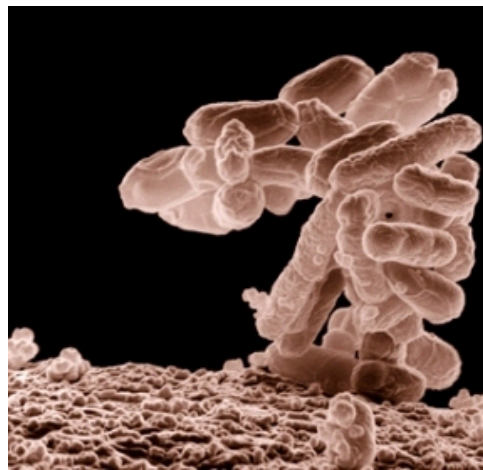
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Throughout human history, plants have been a source of potent medicines, including many cancer drugs discovered over the past few decades. However, it is quite difficult to discover such drugs and obtain them in large quantities from the plants or through chemical synthesis.



A close-up view of *E. coli*.

MIT researchers and collaborators from Tufts University have now engineered bacteria to produce large quantities of a critical compound that is a precursor of the anticancer drug Taxol, originally isolated from the bark of the Pacific yew tree. The bacteria can produce 1,000 times more of the precursor, known as taxadiene, than an engineered microbial strain.

The technique, [described in the Oct. 1 issue of \*Science\*](#), could bring down the manufacturing costs of Taxol and also help scientists discover potential new cancer and other diseases such as hypertension and Alzheimer's, said Greg Stephanopoulos, who led the team of MIT and Tufts researchers and is one of the authors of the paper.

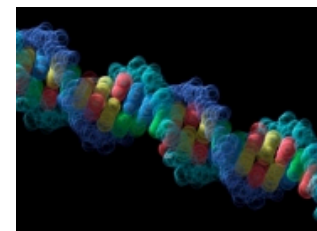
"If you can make Taxol a lot cheaper, that's good, but what really gets people excited is the prospect of using our platform to discover other therapeutic compounds, to be able to bring down the costs of declining new pharmaceutical products and rapidly escalating costs for drug development," said Stephanopoulos, the W.H. Dow Professor of Chemical Engineering at MIT.

Taxol, also known as paclitaxel, is a powerful cell-division inhibitor commonly used to treat ovarian, lung and breast cancers. It is also very expensive — about \$100,000 per dose, although the cost of manufacturing that dose is only a few hundred dollars. (Patients usually receive one dose.)

Two to four Pacific yew trees are required to obtain enough Taxol to treat one patient. In the 1990s, bioengineers came up with a way to produce it in the lab from plant cells, or by extracting key intermediates from plant material like the needles of the decorative yew. These methods generate enough material for patients, but cannot produce sufficient quantities for synthesizing variants that may be far more effective in treating cancer and other diseases. Organic chemists have succeeded in synthesizing Taxol in the lab, but these methods involve 35 to 50 steps and have a very low yield. They are not economical. Also, they follow a different pathway than the plants, which makes it impossible to produce the pathway intermediates and change them into new, potentially more powerful variations.

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“By mimicking nature, we can now begin to produce these intermediates that makes, so people can look at them and see if they have any therapeutic properties,” says Stephanopoulos. Moreover, they can synthesize variants of these intermediates that have therapeutic properties for other diseases.

### Improving efficiency

The complex metabolic sequence that produces Taxol involves at least 17 intermediates and is not fully understood. The team's goal was to optimize production of two Taxol intermediates, taxadiene and taxadien-5-alpha-ol. *E. coli* does not produce taxadiene, but it does synthesize a compound called IPP, which is two steps away from taxadiene. Those two steps normally occur only in plants. MIT postdoctoral associate Ajikumar Parayil recognized that the key to more efficient production of Taxol is an integrated pathway that does not allow potentially toxic intermediates to accumulate. To accomplish this, researchers took a two-pronged approach in engineering *E. coli* to produce taxadiene.

First, the team examined the IPP pathway, which has eight steps, and focused on those reactions that had been determined to be bottlenecks in the synthesis of taxadiene. There is not enough enzyme at those steps, so the entire process is slowed down. They then engineered the bacteria to express multiple copies of those four genes, bypassing the bottlenecks and speeding up IPP production.

To get *E. coli* to convert IPP to taxadiene, the researchers added two plant genes that were modified to function in bacteria, which code for the enzymes needed to perform the two reactions. They also varied the number of copies of the genes to find the most efficient combination. These methods allowed the researchers to boost taxadiene production 1,000 times over levels achieved by other researchers using engineered *E. coli*. The new strain produced 15,000 times more taxadiene over a control strain of *E. coli* to which they just added the two plant genes but did not optimize gene expression of either pathway.

Following taxadiene synthesis, researchers advanced the pathway by adding a critical step toward Taxol synthesis, the conversion of taxadiene to taxadien-5-alpha-ol. This is the first time that taxadien-5-alpha-ol has been produced in microbes. There are still several more steps to go before achieving synthesis of the intermediate from which Taxol can be chemically synthesized. “Though this is only a first step, it is a very promising development and certainly supports this approach and its potential,” says Blaine Pfeifer, assistant professor of chemical and biological engineering at MIT and author of the *Science* paper.

Now that the researchers have achieved taxadiene synthesis, there are still 18 to 20 steps to go before they can generate Taxol. In this study, they showed how to perform the first of those steps.

Joseph Chappell, professor of plant sciences at the University of Kentucky, says the team's yield of taxadiene is impressive, but the addition of several hydroxyl groups, which is required to produce baccatin III, will likely prove more difficult. “They've done one (hydroxylation), but it remains to be seen if they'll be able to couple this with other hydroxylation events needed to build the baccatin III molecule,” he said.

Stephanopoulos and Pfeifer expect that if this technique can eventually be used to manufacture Taxol, it would reduce significantly the cost to produce one gram of the drug. Researchers could also experiment with using these bacteria to create other drugs.