

Journal of Andrology, Vol 19, Issue 5 585-594, Copyright © 1998 by The American Society of Andrology

JOURNAL ARTICLE

Involvement of reactive oxygen species in human sperm acrosome reaction induced by A23187, lysophosphatidylcholine, and biological fluid ultrafiltrates

E. de Lamirande, C. Tsai, A. Harakat and C. Gagnon
Urology Research Laboratory, Royal Victoria Hospital, and the Faculty of Medicine, McGill University, Montreal, Quebec, Canada.

Although recent evidence indicated that the production of reactive oxygen species (ROS) by human spermatozoa may be involved in the regulation of capacitation, very little is known about the role of ROS in the acrosome reaction. To address this issue, Percoll-washed spermatozoa were incubated in Ham's F-10 medium in the absence (no capacitation) or presence (capacitation) of fetal cord serum ultrafiltrate (FCSu) or progesterone. The effects of the ROS scavengers, superoxide dismutase (SOD), and catalase were then tested on the acrosome reaction induced by lysophosphatidylcholine (LPC), A23187, and ultrafiltrates from follicular fluid (FFu) and FCSu, as well as on the protein tyrosine phosphorylation associated with this process. 2-Methyl-6-(p-methoxyphenyl)-3,7-dihydroimidazo [1,2-a] pyrazin-3-one (MCLA)-amplified chemiluminescence was used to determine the extracellular superoxide (O_2^-) production from spermatozoa. The observations that both SOD and catalase reduced (in the case of LPC) or totally prevented (in the other cases) the acrosome reaction of capacitated spermatozoa and that hydrogen peroxide (H_2O_2) or ROS generated by the combination of xanthine and xanthine oxidase (O_2^- , which dismutates to H_2O_2) triggered the acrosome reaction indicated the involvement of ROS in this process. In fact, capacitated spermatozoa in which the acrosome reaction was induced by LPC, A23187, and FFu produced more O_2^- than noncapacitated spermatozoa treated with the same agents. A23187 and LPC had minor effects on protein tyrosine phosphorylation of noncapacitated spermatozoa. However, these inducers caused a decrease in tyrosine phosphorylation of Triton-soluble proteins (mainly those of 37, 42, and 47 kDa) from capacitated spermatozoa, a decrease more pronounced in the presence of SOD. On the other hand, there was a marked increase in tyrosine phosphorylation of few proteins (70 to 105 kDa) from the Triton-insoluble fraction, which was partly reversed by SOD (in the case of LPC and A23187) or catalase (in the case of A23187), or abolished in the presence of the two antioxidants (in the case of A23187). These data indicate that the acrosome reaction is associated with an extracellular O_2^- generation by spermatozoa and that both O_2^- and H_2O_2 may be involved in the regulation of this process. The mechanism by which these ROS act is unknown but may involve tyrosine phosphorylation of sperm proteins.

This Article

- ▶ [Full Text \(PDF\)](#)
- ▶ [Alert me when this article is cited](#)
- ▶ [Alert me if a correction is posted](#)

Services

- ▶ [Similar articles in this journal](#)
- ▶ [Similar articles in PubMed](#)
- ▶ [Alert me to new issues of the journal](#)
- ▶ [Download to citation manager](#)

Citing Articles

- ▶ [Citing Articles via HighWire](#)
- ▶ [Citing Articles via Google Scholar](#)

Google Scholar

- ▶ [Articles by de Lamirande, E.](#)
- ▶ [Articles by Gagnon, C.](#)
- ▶ [Search for Related Content](#)

PubMed

- ▶ [PubMed Citation](#)
- ▶ [Articles by de Lamirande, E.](#)
- ▶ [Articles by Gagnon, C.](#)



HUMAN REPRODUCTION UPDATE

▶ HOME

K. Tremellen

Oxidative stress and male infertility--a clinical perspective
Hum. Reprod. Update, May 1, 2008; 14(3): 243 - 258.

[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



Physiological Reviews

▶ HOME

K. Bedard and K.-H. Krause

The NOX Family of ROS-Generating NADPH Oxidases: Physiology and Pathophysiology

Physiol Rev, January 1, 2007; 87(1): 245 - 313.

[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



Cardiovascular Research

▶ HOME

L. Moldovan, K. Mythreya, P. J. Goldschmidt-Clermont, and L. L. Satterwhite

Reactive oxygen species in vascular endothelial cell motility. Roles of NAD(P)H oxidase and Rac1

Cardiovasc Res, July 15, 2006; 71(2): 236 - 246.

[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



Journal of ANDROLOGY

▶ HOME

A. Agarwal, S. A. Prabakaran, and T. M. Said

Prevention of Oxidative Stress Injury to Sperm
J Androl, November 1, 2005; 26(6): 654 - 660.

[\[Full Text\]](#) [\[PDF\]](#)



HUMAN REPRODUCTION

▶ HOME

C.M. McVicar, D.A. O'Neill, N. McClure, B. Clements, S. McCullough, and S.E.M. Lewis

Effects of vasectomy on spermatogenesis and fertility outcome after testicular sperm extraction combined with ICSI

Hum. Reprod., October 1, 2005; 20(10): 2795 - 2800.

[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



BIOLOGY of REPRODUCTION

▶ HOME

C. O'Flaherty, E. de Lamirande, and C. Gagnon

Reactive Oxygen Species and Protein Kinases Modulate the Level of Phospho-MEK-Like Proteins During Human Sperm Capacitation

Biol Reprod, July 1, 2005; 73(1): 94 - 105.

[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



Molecular Human Reproduction

▶ HOME

L. Liguori, E. de Lamirande, A. Minelli, and C. Gagnon

Various protein kinases regulate human sperm acrosome reaction and the associated phosphorylation of Tyr residues and of the Thr-Glu-Tyr motif

Mol. Hum. Reprod., March 1, 2005; 11(3): 211 - 221.

[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



J. Roca, M. J. Rodriguez, M. A. Gil, G. Carvajal, E. M. Garcia, C. Cuello, J. M. Vazquez, and E. A. Martinez
Survival and In Vitro Fertility of Boar Spermatozoa Frozen in the Presence of Superoxide Dismutase and/or Catalase
J Androl, January 1, 2005; 26(1): 15 - 24.
[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



W.C.L. Ford
Regulation of sperm function by reactive oxygen species
Hum. Reprod. Update, September 1, 2004; 10(5): 387 - 399.
[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



N.C. Feki, P. Therond, M. Couturier, G. Limea, A. Legrand, P. Jouannet, and J. Auger
Human sperm lipid content is modified after migration into human cervical mucus
Mol. Hum. Reprod., February 1, 2004; 10(2): 137 - 142.
[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



J. Baumber, B. A. Ball, J. J. Linfor, and S. A. Meyers
Reactive Oxygen Species and Cryopreservation Promote DNA Fragmentation in Equine Spermatozoa
J Androl, July 1, 2003; 24(4): 621 - 628.
[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



N. Cormier and J. L. Bailey
A Differential Mechanism Is Involved During Heparin- and Cryopreservation-Induced Capacitation of Bovine Spermatozoa
Biol Reprod, July 1, 2003; 69(1): 177 - 185.
[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



L. Lefievre, K. N. Jha, E. de Lamirande, P. E. Visconti, and C. Gagnon
Activation of Protein Kinase A During Human Sperm Capacitation and Acrosome Reaction
J Androl, September 1, 2002; 23(5): 709 - 716.
[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



E. de Lamirande and C. Gagnon
The extracellular signal-regulated kinase (ERK) pathway is involved in human sperm function and modulated by the superoxide anion
Mol. Hum. Reprod., February 1, 2002; 8(2): 124 - 135.
[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



M. B. Herrero, E. de Lamirande, and C. Gagnon

Tyrosine nitration in human spermatozoa: a physiological function of peroxynitrite, the reaction product of nitric oxide and superoxide

Mol. Hum. Reprod., October 1, 2001; 7(10): 913 - 921.

[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)



S. C. Richer and W.C.L. Ford

A critical investigation of NADPH oxidase activity in human spermatozoa

Mol. Hum. Reprod., March 1, 2001; 7(3): 237 - 244.

[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)