

论著

## 五没食子酰基葡萄糖对卵巢癌HO-8910细胞凋亡调控基因表达与胱天蛋白酶凋亡途径的影响

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收稿日期 2011-11-7 修回日期 2012-3-15 网络版发布日期 2012-8-21 接受日期

**摘要** 目的 探讨五没食子酰基葡萄糖(PGG)诱导卵巢癌HO-8910细胞凋亡的作用及诱导胱天蛋白酶凋亡途径的机制。方法 PGG 10, 20, 40和80  $\mu\text{mol} \cdot \text{L}^{-1}$ 处理HO-8910细胞48, 72和96 h后, MTT法检测细胞存活率; Hoechst 33258染色观察HO-8910细胞核形态改变, Annexin V-FITC/PI双染流式细胞术检测细胞凋亡率; Western 印迹法检测细胞内胱天蛋白酶原及活性形式; RT-PCR检测凋亡调控基因Bax、Bcl-2、Bcl-XL、凋亡抑制因子1(CIAP-1)、CIAP-2、存活蛋白、神经元凋亡抑制蛋白(NIAP)、X连锁凋亡抑制蛋白(XIAP)和细胞周期蛋白D1 mRNA表达。结果 PGG 10~80  $\mu\text{mol} \cdot \text{L}^{-1}$ 分别作用48, 72和96 h, 随浓度的增加, 细胞存活率明显降低,  $r$ 分别为0.93, 0.95和0.86 ( $P < 0.05$ )。PGG 40  $\mu\text{mol} \cdot \text{L}^{-1}$ 使HO-8910细胞的细胞核染色质固缩, 出现凋亡形态学改变, 早期凋亡率从正常对照组的(0.6±0.1)%分别增加到(3.4±1.1)%, (9.8±3.7)%和(19±4.5)%, 对晚期凋亡率影响不明显。PGG 20~80  $\mu\text{mol} \cdot \text{L}^{-1}$ 使HO-8910细胞内胱天蛋白酶3, 胱天蛋白酶7和胱天蛋白酶9及其底物多聚腺苷二磷酸核糖聚合酶(PARP)的剪切水平增加, PGG 20~80  $\mu\text{mol} \cdot \text{L}^{-1}$ 均抑制死亡受体FAS的蛋白表达水平并使胱天蛋白酶8总剪切水平降低。PGG 20~80  $\mu\text{mol} \cdot \text{L}^{-1}$ 抑制HO-8910细胞中细胞周期蛋白D1, Bcl-2, Bcl-XL和NIAP mRNA的表达, 上调CIAP-1 mRNA的表达, 对基因Bax, CIAP-2和XIAP mRNA表达影响不明显; PGG 20  $\mu\text{mol} \cdot \text{L}^{-1}$ 抑制存活蛋白基因mRNA的表达, 但是增加处理浓度却上调存活蛋白基因mRNA的表达。结论 PGG可能通过抑制凋亡抑制基因Bcl-2和Bcl-XL的表达从而诱导HO-8910细胞内胱天蛋白酶9依赖的内源性凋亡途径, 并诱导细胞凋亡。

**关键词** [五没食子酰基葡萄糖](#) [卵巢癌HO-8910细胞](#) [细胞凋亡](#) [Bcl-2家族](#) [凋亡抑制因子](#) [胱天蛋白酶](#)

分类号 [R285.5](#)

## Effect of pentagalloylglucose on expression of apoptosis regulator genes and caspase-dependent apoptosis pathways in HO-8910 cells

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### Abstract

**OBJECTIVE** To explore the effect of pentagalloylglucose(PGG) on the apoptosis of ovarian cancer cells and the mechanism underlying the induced caspase-dependent apoptosis pathways in ovarian cancer HO-8910 cells. **METHODS** HO-8910 cells were cultured with PGG 10,20,40 and 80  $\mu\text{mol} \cdot \text{L}^{-1}$  for 48, 72 or 96 h. The cell survival rate was detected by MTT assay. The morphological alteration of nucleus of HO-8910 cells was observed under a fluorescence microscope after Hoechst 33258 staining and the apoptosis ratio was determined by Annexin V-FITC/PI double-staining combined with flow cytometry assay. Western blotting was employed to detect the pro-caspases or active caspases(c-caspases). The mRNA expression of apoptosis regulator genes Bcl-2, Bcl-XL, CIAP-1, CIAP-2, survivin, NIAP, XIAP and cycle regulator gene cyclin D1 was visualized by RT-PCR. **RESULTS** PGG inhibited the growth *in vitro* of HO-8910 cells in a time- and concentration-dependent manner during 48, 72 and 96 h treatment ( $P < 0.05$ ); the correlation coefficient was 0.93, 0.95 and 0.86, respectively. PGG treatment induced nuclear chromatin pycnosis of HO-8910 cells and the morphological alteration of nuclei in apoptosis. The ratio of apoptosis in HO-8910 cells rose with lengthened treatment and increased doses of PGG. PGG 20-80  $\mu\text{mol} \cdot \text{L}^{-1}$  promoted the cleavage of caspases 3, 7 and 9 and the related PARP substrate in HO-8910 cells. However, PGG 20-80  $\mu\text{mol} \cdot \text{L}^{-1}$  inhibited the protein expression of the death receptor FAS and the cleavage of caspase 8 at 24 and 36 h, but down-regulated the mRNA expression of cyclin D1, Bcl-2, Bcl-XL and NIAP genes at 12 h. It had no effect on Bax, CIAP-2 and XIAP mRNA, but up-regulated the mRNA expression of CIAP-1. The effect of PGG on the surviving mRNA was not concentration-dependent. **CONCLUSION** PGG might induce caspase 9-dependent endogenous caspase pathways by inhibiting the expression of apoptosis inhibitor genes, and trigger HO-8910 cell apoptosis. It's likely that PGG has repressed the caspase 8-dependent apoptosis pathway in HO-8910 cells.

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**Key words** [pentagalloylglucose](#) [HO-8910 cells](#) [apoptosis](#) [Bcl-2 family](#) [apoptosis-inhibiting factors](#)  
[caspases](#)

DOI: 10.3867/j.issn.1000-3002.2012.04.012

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