



植物病理学报 » 2013, Vol. 43 » Issue (4): 401-410 DOI:

植物病害及其防治

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脂肽类抗生素fengycin在枯草芽孢杆菌NCD-2菌株抑制番茄灰霉病菌中的功能分析

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The fengycin lipopeptides are major components in the *Bacillus subtilis* strain NCD-2 against the growth of *Botrytis cinerea*

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摘要 枯草芽孢杆菌NCD-2菌株及其无细胞发酵液对多种植物病原真菌具有较强的拮抗活性。为了明确NCD-2菌株抑菌作用机理,本研究利用快速蛋白液相色谱技术(FPLC)结合抑菌活性测定,对NCD-2菌株的脂肽类抑菌物质进行分离、纯化,经质谱鉴定,该抑菌物质为芬莽素(fengycin)。从NCD-2菌株中克隆出fengycin合成酶基因fenC及其上游启动子序列,通过同源重组技术对fenC基因进行了内缺失突变,同NCD-2野生型菌株相比,fenC基因缺失突变子丧失了fengycin的合成能力,同时该突变子显著降低了对番茄灰霉病菌的拮抗活性。进一步在离体叶片上比较了NCD-2野生型菌株和突变子之间脂肽提取物和菌体细胞对番茄灰霉病的保护作用,结果发现,突变子不论是脂肽提取物还是菌体细胞显著降低了对番茄灰霉病的防治作用。

关键词: 枯草芽孢杆菌 番茄灰霉病菌 fengycin 突变

Abstract: The *Bacillus subtilis* strain NCD-2 and its cell-free supernatant showed strong inhibitive activity against the growth of phytopathogens. The NCD-2 lipopeptide extracts showed a strong inhibitive activity against the growth of *Botrytis cinerea*. The major antifungal activity compounds were isolated from the lipopeptides by Fast Protein Liquid Chromatography (FPLC), and identified as fengycin based on the profiles of matrix assisted laser desorption/ionization-time of flight mass spectrometry (MALDI-TOF). The antifungal activity of fengycin was supported by in-frameless deletion mutagenesis and it targeted to suppress the biosynthesis of fengycin production. Compared to the strain NCD-2 wild type, the fengycin-deficient mutant strain decreased the antifungal capability significantly. Further bioassay indicated that the lipopeptides extracted from the fengycin-deficient mutant not only lost the inhibitive activity against the growth of *B. cinerea* *in vitro*, but also significantly decreased the control effect against tomato grey mold on detached tomato leaves. The results could be confirmed by assays using bacterial suspension between strain NCD-2 wild type and the fengycin-deficient mutant.

Key words: *Bacillus subtilis* *Botrytis cinerea* fengycin mutagenesis

收稿日期: 2012-09-04;

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引用本文：

董伟欣,李宝庆,李社增等. 脂肽类抗生素fengycin在枯草芽孢杆菌NCD-2菌株抑制番茄灰霉病菌中的功能分析[J]. 植物病理学报, 2013, 43(4): 401-410.

DONG Wei-xin, LI Bao-qing, LI She-zeng et al. The fengycin lipopeptides are major components in the *Bacillus subtilis* strain NCD-2 against the growth of *Botrytis cinerea*[J]. Acta Phytopathologica Sinica, 2013, 43(4): 401-410.

- [1] Montesinos E. Antimicrobial peptides and plant disease control [J]. FEMS Microbiology Letters, 2007, 270(1): 1-11. 
- [2] Jacques P. Surfactin and other lipopeptides from *Bacillus* spp. [J]. Biosurfactants, 2011, 20: 57-91. 
- [3] Stein T. *Bacillus subtilis* antibiotics: structures, syntheses and specific functions [J]. Molecular Micro-biology, 2005, 56(4): 845-857. 
- [4] Arrebol E, Jacobs R, Korsten L. Iturin A is the principal inhibitor in the biocontrol activity of *Bacillus amyloliquefaciens* PPCB004 against postharvest fungal pathogens [J]. Journal of Applied Microbiology, 2010, 108(2): 386-395. 
- [5] Deleu M, Paquot M, Nylander T. Effect of fengycin, a lipopeptide produced by *Bacillus subtilis*, on model biomembranes [J]. Biophysical Journal, 2008, 94(7): 2667-2679. 
- [6] Maget-Dana R, Thimon L, Peypoux F, et al. Surfactin/iturin A interactions may explain the synergistic effect of surfactin on the biological properties of iturin A [J]. Biochimie, 1992, 74(12): 1047-1051. 
- [7] Tao Y, Bie X, Lv F, et al. Antifungal activity and mechanism of fengycin in the presence and absence of commercial surfactin against *Rhizopus stolonifer* [J]. The Journal of Microbiology, 2011, 49(1): 146-150. 
- [8] Ongena M, Duby F, Jourdan E, et al. *Bacillus subtilis* M4 decreases plant susceptibility towards fungal pathogens by increasing host resistance associated with differential gene expression [J]. Applied Microbiology and Biotechnology, 2005, 67(5): 692-698. 
- [9] Ongena M, Jourdan E, Adam A, et al. Surfactin and fengycin lipopeptides of *Bacillus subtilis* as elicitors of induced systemic resistance in plants [J]. Environmental Microbiology, 2007, 9(4): 1084-1090. 
- [10] Li S Z, Lu X Y, Ma P, et al. Evaluation of biocontrol potential of a bacterial strain NCD-2 against cotton verticillium wilt in field trials (in Chinese) [J]. Acta Phytopathology Sinica (植物病理学报), 2005, 35(5): 451-455.
- [11] Meng L H, Li S Z, Guo Q G, et al. Primary analysis on the antifungal activity of the protein produced by *Bacillus subtilis* NCD-2 against *Verticillium dahliae* (in Chinese) [J]. Acta Agriculturae Boreali-Sinica (华北农学报), 2008, 23(1): 189-193.
- [12] Wei Y H, Wang L C, Chen W C, et al. Production and characterization of fengycin by indigenous *Bacillus subtilis* F29-3 originating from a potato farm [J]. International Journal of Molecular Sciences, 2010, 11(11): 4526-4538. 
- [13] Liu N, Guo Q G, An H, et al. Identification of biocontrol bacterial strain BAB-1 against tomato grey mold and optimization of fermentation conditions (in Chinese) [J]. Journal of Agricultural Science and Technology (中国农业科技导报), 2009, 11(2): 56-62.
- [14] Arguelles-Arias A, Ongena M, Halimi B, et al. *Bacillus amyloliquefaciens* GA1 as a source of potent antibiotics and other secondary metabolites for biocontrol of plant pathogens [J]. Microbial Cell Factories, 2009, 8(63): 1-12. 
- [15] Leenders F, Stein T H, Kablitz B, et al. Rapid typing of *Bacillus subtilis* strains by their secondary metabolites using matrix-assisted laser desorption/ionization mass spectrometry of intact cells [J]. Rapid Communications In Mass Spectrometry, 1999, 13(10): 943-949. 3.0.CO;2-0 target="_blank">> 
- [16] Martin P, Lohr J, Dean D. Transformation of *Bacillus thuringiensis* protoplasts by plasmid deoxyribonucleic acid [J]. Journal of Bacteriology, 1981, 145(2): 980-983.
- [17] Arnaud M, Chastanet A, Debarbouille M. New vector for efficient allelic replacement in naturally nontransformable, low-GC-content, gram-positive bacteria [J]. Applied and Environmental Microbiology, 2004, 70(11): 6887-6891. 
- [18] Wang Y, Deng C, Peng Q, et al. Effect of quorum sensing response regulator *nprR* deletion on expression of Cry protein in *Bacillus thuringiensis* (in Chinese) [J]. Acta Microbiological Sinica(微生物学报), 2010, 50(11): 1550-1555.
- [19] De Faria A F, Stefani D, Vaz B G, et al. Purification and structural characterization of fengycin homologues produced by *Bacillus subtilis* LSFM-05 grown on raw glycerol [J]. Journal of Industrial Microbiology and Biotechnology, 2011, 38(7): 1-9. 
- [20] Wu C Y, Chen C L, Lee Y H, et al. Nonribosomal synthesis of fengycin on an enzyme complex formed by fengycin synthetases[J]. Journal of Biological Chemistry, 2007, 282: 5608-5616.

- [21] Hu L B, Shi Z Q, Zhang T, et al. Fengycin antibiotic isolated from B-FS01 culture inhibits the growth of *Fusarium moniliforme* Sheldon ATCC 38932 [J]. FEMS Microbiology Letters, 2007, 272(1): 91-98. 
- [22] Vater J, Kablitz B, Wilde C, et al. Matrix-assisted laser desorption ionization-time of flight mass spectrometry of lipopeptide biosurfactants in whole cells and culture filtrates of *Bacillus subtilis* C-1 isolated from petroleum sludge [J]. Applied and Environmental Microbiology, 2002, 68(12): 6210-6219. 
- [23] Li B Q, Lu X Y, Guo Q G, et al. Isolation and identification of lipopeptides and volatile compounds produced by *Bacillus subtilis* strain BAB-1 (in Chinese) [J]. Scientia Agricultura Sinica(中国农业科学), 2010, 43(17): 3547-3554.
- [24] Toure Y, Ongena M, Jacques P, et al. Role of lipopeptides produced by *Bacillus subtilis* GA1 in the reduction of grey mould disease caused by *Botrytis cinerea* on apple [J]. Journal of Applied Microbiology, 2004, 96(5): 1151-1160. 
- [25] Zeriouh H, Romero D, Garcia-Gutierrez L, et al. The Iturin-like lipopeptides are essential components in the biological control arsenal of *Bacillus subtilis* against bacterial diseases of cucurbits [J]. Molecular Plant-Microbe Interactions, 2011, 24(12): 1540-1552. 
- [26] Liu J, Zhou T, He D, et al. Functions of lipopeptides bacillomycin D and fengycin in antagonism of *Bacillus amyloliquefaciens* C06 towards *Monilia fructicola* [J]. Journal of Molecular Microbiology and Biotechnology, 2011, 20(1): 43-52. 
- [27] Yanez-Mendizabal V, Zeriouh H, Vinas I, et al. Biological control of peach brown rot (*Monilia* spp.) by *Bacillus subtilis* CPA-8 is based on production of fengycin-like lipopeptides [J]. European Journal of Plant Pathology, 2012, 132(4): 1-11. 
- [28] Ke W J, Chang B Y, Lin T P, et al. Activation of the promoter of the fengycin synthetase operon by the UP element [J]. Journal of Bacteriology, 2009, 191(14): 4615-4623. 
- [1] 郑雪芳^{*}, 刘波^{*}, 林乃铨^{*}, 朱育菁, 车建美. 青枯雷尔氏菌无致病力突变菌株的构建及其防效评价模型分析[J]. 植物病理学报, 2013, 43(5): 518-531.
- [2] 侯毅平, 章四平, 王建新, 陈长军, 周明国^{*}. 枯草芽孢杆菌NJ-18对油菜菌核病的防治效果及其定殖动态[J]. 植物病理学报, 2013, 43(4): 411-417.
- [3] 王阳, 马东方, 张亮, 卢丽丽, 王美南, 井金学, 康振生. GUS基因插入导致的小麦条锈菌突变体遗传稳定性和毒性变异研究[J]. 植物病理学报, 2013, 43(1): 42-49.
- [4] 王洁, 刘金亮, 阴筱, 高瑞, 刘红梅, 李向东, 刘焕庭. HC Pro 3个参与抑制RNA沉默氨基酸位点的鉴定[J]. 植物病理学报, 2012, 42(3): 267-273.
- [5] 邹丽芳, 周丹, 刘之洋, 邹华松, 陈功友. 水稻黄单胞菌致病相关基因插入突变体系的建立[J]. 植物病理学报, 2012, 42(2): 176-185.
- [6] 陈析丰, 金杨, 马伯军. 水稻类病变突变体及抗病性的研究进展[J]. 植物病理学报, 2011, 41(1): 1-9.
- [7] 王洁, 刘金亮, 阴筱, 高瑞, 刘红梅, 李向东, 刘焕庭. HC Pro 3个参与抑制RNA沉默氨基酸位点的鉴定[J]. 植物病理学报, 0, (): 7-.