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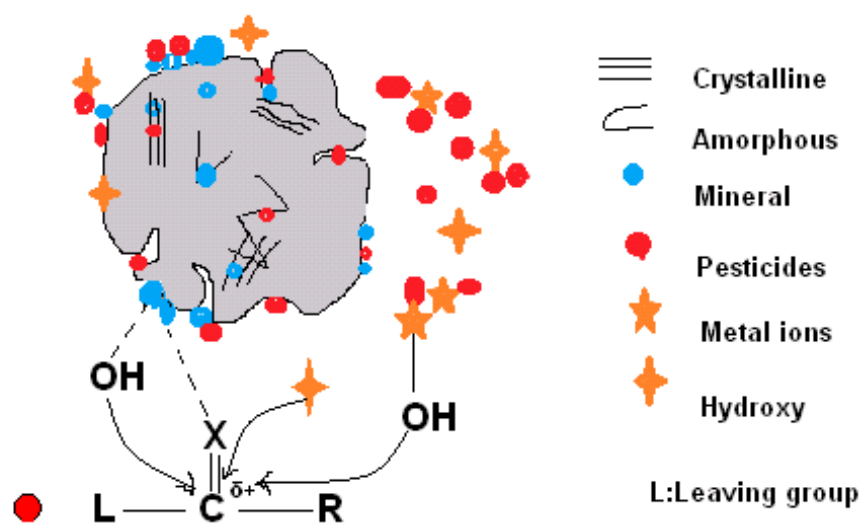
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Highlights of the manuscript:

- High ash content biochar can increase solution pH and released metal ions.
- Ash in biochar can combine pesticide through specific interactions.
- Composition and structure of biochar is favor for the hydrolysis of pesticides.

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1 Adsorption and Catalytic Hydrolysis of Carbaryl and Atrazine on Pig Manure –
2 Derived Biochars: Impact of Structural Properties of Biochars

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26 **Adsorption and Catalytic Hydrolysis of Carbaryl and Atrazine on Pig Manure –**
27 **Derived Biochars: Impact of Structural Properties of Biochars**

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32 **Abstract:** Biochars were produced from pig manure to elucidate the influence of biochars
33 with high ash contents on the fate of pesticides. Adsorption and catalytic hydrolysis of
34 carbaryl and atrazine on original biochars and deashed biochars were investigated. The
35 two pesticides were substantially adsorbed by the biochars, with organic carbon
36 normalized sorption coefficient (K_{oc}) values of $10^{2.65}$ - $10^{3.66}$ L/kg for carbaryl and
37 $10^{1.90}$ - $10^{3.57}$ L/kg for atrazine at C_e of 0.5 mg/L. Hydrophobic effect alone could not explain
38 the sorption, and several other processes including pore-filling and π - π electron
39 donor-acceptor interactions were involved in pesticide adsorption. Adsorption increased
40 greatly on the deashed biochar, indicating that some organic sorption sites in the original
41 biochars were blocked or difficult to access due to their interactions with inorganic moiety.
42 The pesticides were found to hydrolyze faster in the presence of biochars, and in the
43 presence of biochar pyrolyzed at 700 °C, carbaryl and atrazine were decomposed by
44 71.8% and 27.9% in 12 h, respectively. The elevated solution pH was the main reason for
45 the enhanced hydrolysis; however both the mineral surface and dissolved metal ions
46 released from the biochars were confirmed to catalyze the hydrolysis.

47 **Keywords:** Biochar; Carbaryl; Atrazine; Sorption; Hydrolysis

48 1. Introduction

49 Biochar is a carbonaceous material produced from oxygen-limited pyrolysis of
50 biomass [1]. Interest in biochars has recently increased because the application of
51 biochars on farmland has been advocated to provide a means of sequestering carbon and
52 increasing soil fertility [2-4]. Biochars show extraordinarily strong adsorption affinities for
53 contaminants [5-8]. The adsorption of hydrophobic organic chemicals (HOCs) on biochars
54 and other carbonaceous materials has been well documented [5, 9, 10], with adsorption
55 coefficients (K_d) of 10^3 - 10^8 L/kg. The highly aromatic and porous structures in biochars,
56 which favor both hydrophobic effect and specific interactions, have been suggested to
57 contribute to the high adsorption coefficients [11,12].

58 Until now, the majority of studies concerning the adsorption of organic chemicals
59 have been limited on biochars derived from plant residues [13-17]. Plant-derived biochars
60 commonly contain low amounts of inorganic moiety (ash), so the effect of ash on HOC
61 adsorption has usually been neglected. Livestock manure is an important agricultural
62 waste biomass, and using such manure to produce biochar is a possible approach to
63 recycling livestock manure [18]. Biochars derived from livestock manures may have quite
64 different compositions compared to those from plant residues. One important difference is
65 that biochars derived from livestock manures usually contain high ash content [19-20].
66 The high content of ash and its interaction with organic moieties may influence the
67 interactions of biochars with organic pollutants. The influence of the interaction between
68 inorganic and organic moieties on sorption capacity has been extensively studied on
69 natural geosorbents [21, 22], with conflictive results. Generally, mineral surfaces inhibit

70 the non-specific adsorption of organic moieties by reducing the accessibility of organic
71 sorption sites [23]; this is particularly true for the partitioning of apolar organics [24].
72 However, mineral surfaces can also enhance sorption of polar organic chemicals through
73 specific interactions, such as electron donor-acceptor [25] and H-bonding interactions [26].
74 For ionizable organics, they may provide sites for additional interactions of cation or anion
75 binding. Though these mechanisms have been proposed previously, the influence of
76 biochars of high ash content on the fate of organic pollutants has seldom been studied
77 [27-29].

78 Pesticides are common soil pollutants. Adsorption on biochars has been studied for
79 some pesticides, such as diuron [30], triazine [31], and dieldrin [32]. Though the
80 adsorption capacity of biochars for polar pesticides was not as large as those for apolar
81 HOCs [33], biochars are generally good adsorbents for pesticides, with K_d values ranging
82 between 10^1 and 10^3 L/kg [30-32]. Hence, the application of biochars to soils can have
83 great impact on the fate of pesticides. Moreover, modern pesticides are designed to be
84 easily degraded. Hydrolysis is an important mechanism for abiotic degradation of
85 pesticides. Many factors influence pesticide hydrolysis, which include the pH, dissolved
86 ions as well as the presence of clay and metal oxides that can catalyze the hydrolysis
87 reaction [34-36]. Manure-derived biochar contains a high ash content, which is expected
88 to influence the hydrolysis of pesticides. However, there is no prior study on this aspect of
89 biochars.

90 The objective of this study was to obtain a thorough understanding about the effects
91 of biochars with high ash content on the fate of pesticides. Two biochar samples were

92 produced from pig manure by pyrolysis at different temperatures. One carbamate
93 insecticide, carbaryl and one S – triazine herbicide, atrazine were selected as model
94 pesticides. The adsorption and the catalytic hydrolysis of these two pesticides were
95 studied on both original and deashed biochars. The mechanisms of adsorption and
96 catalytic hydrolysis were then discussed.

97 **2. Experimental Section**

98 **2.1 Materials.**

99 Carbaryl (98%) was obtained from Haili Chemical Company (Hunan, China), and
100 atrazine (97%) was obtained from Rainbow Chemical Company (Shandong, China). The
101 pesticides were used in batch experiments without purification. Analytical standards for
102 the pesticides were purchased from Accustandard (Connecticut, US). Other chemicals
103 used in this study were all analytical or HPLC grade.

104 **2.2 Biochars.**

105 Pig manure stock was collected from a hoggery in Jixian County, Tianjin, China. The
106 pig manure was air-dried and ground to pass through a 2-mm sieve. The manure was
107 then heated at 350 °C or 700 °C in a ceramic pot covered with a tight-fitting lid (where
108 oxygen was soon exhausted) in a preheated muffle furnace for 2 h [37]. The produced
109 biochars were ground to pass through 0.038 - 0.075 mm (200 - 400 mesh) sieves and
110 stored in amber glass bottles. Based on the temperature they were heated at, the
111 biochars were designated as BC350 and BC700.

112 To remove the ash (deashing) of the biochars, the samples of the original biochars
113 were treated with acid [38]. Batches consisting of 20 g of biochar were mixed and shaken

114 in 200 mL of 1 M HCl for 10 h. That treatment was carried out 4 times for each batch. The
115 entire procedure was then repeated using 200 mL of a 1 M: 1 M HCl – HF solution. The
116 treated biochars were rinsed with de-ionized water several times to remove residual acid
117 and soluble salts until the solution pH was neutral. The resulting biochar samples were
118 freeze-dried and stored in desiccators for later use. Base on the original heating
119 temperatures, these deashed biochar samples were referred to as DABC350 and
120 DABC700.

121 **2.3 Characterization of Biochars.**

122 Bulk organic elemental composition (C, H, and N) of the biochar samples was
123 determined by an element analyzer (Elementar Vario EL, Germany). Ash content of the
124 samples was measured by the residual weight after heating the biochars at 750 °C for 6 h
125 [37], and the O content was calculated by mass difference. Surface elemental composition
126 of biochars was quantified using an X-ray photoelectron spectrometer (XPS) (ULVAC-PHI
127 PHI 5000 VersaProbe II, Japan). Surface area and pore volume were measured by a
128 surface area analyzer (Quantachrome NOVA 2200e, USA). Solid-state ^{13}C NMR spectra
129 were obtained by using the cross-polarization and total-sideband suppression (cptoss)
130 method on a 7 mm magic angle-spinning (MAS) probe (BRUKER AVANCE III, Germany).
131 Fourier transform infrared spectroscopy (FTIR) spectra of biochars were recorded
132 between 4000 and 400 cm^{-1} wave numbers using an FTIR spectrometer (Perkin Elmer
133 1725 X, USA).

134 **2.4 Batch Sorption Experiments.**

135 A total of four sorbents, i.e., BC350, BC700, DABC350, and DABC700, were studied.

136 Sorption isotherms for each sorbent were determined at six different initial solution
137 concentrations, with duplicate measurements at each concentration point. An aliquot of 50
138 mg of each sorbent was weighted into a 40 mL vial (US EPA) with a
139 polytetrafluoroethylene (Teflon)-lined screw cap, followed by 40 mL of a background
140 solution. The background solution contained 5 mM CaCl₂ to maintain a constant ionic
141 strength and 200 mg/L HgCl₂ (pH 6.5) to inhibit aerobic biodegradation. After
142 pre-equilibration for 24 h, designated amounts of pesticide stock solutions were spiked
143 into each vial, with the volume percentage of methanol kept below 0.2% to minimize
144 possible cosolvent effects on sorption. Initial concentrations of carbaryl were 1, 2.5, 5, 8,
145 20, 40 mg/L; while those of atrazine were 2, 4, 8, 10, 20, 30 mg/L. All of the vials were left
146 with minimal head space and sealed with Teflon film. The vials were put in a shaker
147 operated at 150 rpm and 20 – 24 °C in the dark for 24 h. The equilibration time was
148 predetermined, and 24 h was confirmed to be enough to reach an apparent equilibrium.
149 After equilibration, the vials were centrifuged at 3000 rpm for 20 min. Before analyzing by
150 high performance liquid chromatography (HPLC), the supernatant was filtered through
151 0.45 µm PTFE syringe filters (Shanghai ANPLE, China), and external standards were also
152 filtered to correct the solute loss due to filtration (less than 4%). The results of control
153 samples without biochar indicated that the losses of pesticides by evaporation,
154 photochemical decomposition, and sorption to vials, etc. were less than 4%.

155 Details of pretreatment and analysis of samples were described in Supplementary
156 Information (SI).

157 **2.5 Hydrolysis in the Presence of Biochars.**

158 Hydrolysis of the two pesticides was investigated in suspensions containing the
159 original biochars. Aliquot of 50 mg BC350 and BC700 were suspended in 40 mL
160 background solution (the same as in sorption experiment) in US EPA vials. After 24 h of
161 pre-equilibrium, pesticide stock solution was added to each vial to obtain spiked
162 concentrations of 5 mg/L. The vials were capped and shaken in the dark at 150 rpm at 20
163 – 24 °C for 7 d. Pesticide concentrations in the liquid phase and on biochar particles were
164 then measured. In addition, hydrolysis in the presence of 500 mg of original biochar was
165 tested to investigate the influence of biochar dose. The residue pesticides both in
166 aqueous and on solid phase were measured using the methods described in the SI and
167 the hydrolysis extents of the pesticides were calculated based on mass balance. The pH
168 of each suspension was monitored during the entire experiment.

169 **2.6 Mechanisms behind the Enhanced Hydrolysis.**

170 To investigate mechanisms for the enhanced catalytic hydrolysis of the pesticides in
171 the presence of biochars, several hydrolysis experiments were designed. Firstly,
172 hydrolysis was performed in background solutions without biochars at pH values of 6.5
173 and 9.1, which were similar to the pH values measured in the suspensions with 50 and
174 500 mg of biochar, respectively. The pH values of the background solutions were adjusted
175 using phosphate or boric acid/ borax buffer solution as appropriate. Secondly, hydrolysis
176 was performed in biochar suspensions (500 mg) with the solution pH adjusted to neutral
177 pH (6.8) before the pesticides were spiked. Thirdly, hydrolysis tests were performed in
178 biochar leachate. For this set of tests, 500 mg samples of biochar were shaken in 40 mL

179 background solutions at 150 rpm and 20 - 24 °C for 24 h. The biochars were then
180 removed from the suspensions by filtration through 0.45 μ m membranes. The leachate
181 solutions were divided into two groups, with the pH of one group adjusted to neutral (6.8)
182 and the other not adjusted. Other manipulations were the same as described above.

183 **3 Results and Discussion**

184 **3.1 Characterization of Biochars.**

185 The bulk chemical compositions of the original biochars derived from pig manure at
186 the two temperatures and the corresponding deashed biochars are shown in Table 1. The
187 ash content of the biochars was greater at higher pyrolyzing temperature (45.3% for
188 BC350 and 66.8% for BC700). These are much higher than the common ash contents of
189 biochars derived from plant sources [37]. The carbon content of BC700 (25.2%) was lower
190 than that of BC350 (31.6%). This finding is not in agreement with an earlier study on
191 biochars with low ash contents [33]. As the charring temperature increased, some
192 amorphous carbon was burn into carbon dioxide [39], resulting in the higher ash content
193 and lower carbon content in BC700 compared to BC350.

194 At higher temperatures, some of the amorphous carbon could also be transformed
195 into aromatic carbon [37]. The atomic ratios of H/C and [(O + N)/ C] are recognized as
196 indices for aromaticity and polarity, respectively [40]. In the present study, bulk aromaticity
197 of the biochar increased and polarity decreased with charring temperature, as indicated
198 by the decreasing H/C and [(O + N)/ C] ratios.

199 The ^{13}C NMR spectra showed that the proportion of alkyl carbon in the total organic
200 carbon of the biochars decreased from 30% to 13%, while the aromatic carbon proportion

201 slightly increased from 37% to 43% as the charring temperature increased. Deashing
202 treatment efficiently reduced the ash content of biochars (Table 1). As a result, organic
203 matter in the deashed biochars greatly increased (> 93%). The bulk polarity of the
204 deashed biochars decreased as indicated by the reducing $[(O + N)/ C]$; while the
205 aromaticity increased as indicated by the reducing H/C. Deashing also increased the
206 number of acidic functional groups, such as phenolic and carboxyl groups (Table 1). This
207 observation is in agreement with previous findings showing that alkaline functional groups
208 are decreased by acid treatment [41].

209 Surface elemental compositions, surface areas and pore volume of the biochars are
210 presented in Table 2. The surface elemental composition of the original biochars was very
211 complex, with carbon as the dominant element, followed by oxygen. The results also
212 revealed the presence of mineral elements such as Al and Si on the surfaces of original
213 biochars. The surface polarity $[(O + N)/ C]$ of BC350 was smaller than that for BC700,
214 which was opposite to the trend in the polarity for the bulk material. The different trends in
215 bulk and surface polarity between BC350 and BC700 indicated their heterogeneous
216 structure.

217 All surface element contents decreased after deashing except for C. The elements of
218 Al and P even could not be detected after deashing. Similar to the bulk biochar, the
219 surface polarity of the deashed biochars decreased, as indicated by the decreasing $[(O +$
220 $N)/ C]$. The reduction degree of polarity on the surface was greater than that of the bulk
221 biochar, suggesting that mineral oxides and salts (containing N, P, and S) were the main
222 components of ash covering the surface of biochars. The inorganic fraction was not

223 removed completely after deashing, indicating that some minerals were masked by
224 organic matter or were located deeper in the biochar.

225 The surface area of biochars increased from 23.8 to 32.6 m²/g as charring
226 temperature increased. This increase was attributed to a greater transformation in the
227 elemental composition with increasing temperature. After deashing, the surface area and
228 pore volume increased due to the removal of surface minerals, and greater change
229 occurred in the BC700. The characteristics of surface area and pore volume of biochar
230 depend much on pyrolyzing temperature. At low temperature, amorphous carbon is the
231 main carbon, and pores are not well developed. Some pores are blocked by aliphatic or
232 volatile constituents that also reduce surface area. Under high temperature, amorphous
233 carbons condense to crystalline carbons, more pores are formed, and aliphatic and
234 volatile constituents are removed [37]. This is why greater change occurred in BC700 with
235 more micropores as compared to BC350 after the removal of inorganic constituents by
236 deashing.

237 The FTIR fingerprints appear to be quite different for biochars derived at different
238 temperatures (Figure 1). In BC700, hydroxyl was oxidized as indicated by the loss of peak
239 intensity at 3665 cm⁻¹, while aliphatic carbon was transformed as indicated by the
240 reduction in intensity of peaks at 2885 – 2850 cm⁻¹ and 1740 - 1700 cm⁻¹ [37]. After
241 deashing, the amount of carboxyl groups increased slightly, as indicated by the increasing
242 height of peaks at 1500 – 1450 cm⁻¹ and 1350-1300 cm⁻¹. These results are consistent
243 with the ¹³C NMR results (Table 1).

244 **3.2 Sorption Isotherms.**

245 Sorption isotherms of carbaryl and atrazine on the original and deashed biochars are
 246 presented in Figure 2. The adsorption data were fitted to the Freundlich model given by Q_e
 247 $= K_f C_e^n$, where Q_e (mg/kg) and C_e (mg/L) are the adsorbed and aqueous – phase
 248 equilibrium concentrations, respectively; K_f ((mg/kg)/ (mg/L)ⁿ) is the affinity coefficient; and
 249 n is the linearity index. All samples showed nonlinear isotherms except atrazine on BC350,
 250 and the nonlinearity of sorption isotherm was greater at higher pyrolyzing temperature
 251 (Table 3). Biochars could effectively sorb carbaryl and atrazine, and the corresponding K_f
 252 values of on BC700 were $10^{2.85}$ and $10^{2.77}$, respectively, being significantly higher
 253 ($P < 0.001$) than those on BC350 ($10^{2.06}$ and $10^{1.40}$). After deashing, the sorption capacity of
 254 biochars increased significantly ($P < 0.001$). The K_f of carbaryl and atrazine increased to
 255 $10^{3.35}$ and $10^{3.15}$ on DABC350, and to $10^{3.70}$ and $10^{3.55}$ on DABC700, respectively. Hence,
 256 among the two biochars, BC700 had greater sorption capacity; and deashing further
 257 enhanced the sorption.

258 It is well known that organic carbons play a crucial role in the sorption of organics. To
 259 evaluate the sorption capacity of organic carbons in different biochars, the organic carbon
 260 normalized sorption coefficients (K_{oc}) at equilibrium concentrations of 0.5 and 20 mg/L
 261 were calculated (Table 3) by $K_{oc} = Q_e / (C_e f_{oc})$, where f_{oc} (C contents in Table1) is organic
 262 carbon content of the sorbent [42,43]. Firstly, the K_{oc} values were $10^{2.65}$ - $10^{4.00}$ L/kg for
 263 carbaryl and $10^{1.90}$ - $10^{3.85}$ L/kg for atrazine at C_e of 0.5 mg/L, being greater for biochars
 264 acquired at higher temperature. This suggests that the organic carbon in biochars
 265 generated at different temperatures have different sorption capacity. Secondly, K_{oc}
 266 increased after deashing, suggesting that the removal of ash enhanced the accessibility of

267 sorption sites in organic moiety. Thirdly, K_{oc} decreased at higher solute concentration (20
268 mg/L), suggesting the heterogeneous nature of sorption sites. The sites with high sorption
269 affinity may have been saturated in face to high solute concentration. The K_{oc} in the
270 present study are higher than literature values of carbaryl ($10^{2.02}$ - $10^{2.59}$) in soil [44] and
271 atrazine ($10^{1.91}$ - $10^{2.42}$) in biochar amended soil [45].

272 3.3 Sorption Mechanisms.

273 To elucidate the controlling factor in biochar on sorption, K_{oc} values were correlated
274 with the structural parameters of the biochars. No significant correlation was found
275 between K_{oc} values of pesticides on biochars and the hydrophobic carbon contents (alkyl
276 and aromatic moieties, Table 1) in biochars. As hydrophobic carbon components,
277 aromatic carbon can provide sorption sites for hydrophobic effect and pore-filling, and
278 some studies have found that K_{oc} correlates positively with aromatic carbon contents in
279 sorbents [24, 46]. However, in this study, due to the high content of ash, the aromatic
280 carbon content of biochars is positively correlated with the O-alkyl and carboxyl carbon
281 content (Pearson correlation coefficient, $r = 0.983, 0.988, p < 0.05$). The surrounding polar
282 O-alkyl and carboxyl groups would have reduced the accessibility of aromatic domains to
283 pesticides. Hence, hydrophobic partition is not the only factor affecting pesticide sorption
284 on biochars, and other specific interactions may be involved.

285 To reveal the possible effect of porosity on sorption, surface area-normalized
286 adsorption isotherms of the two pesticides were drawn (Figure A.1 in Supplementary
287 Information). It can be seen that the normalized sorption on BC350 was obviously lower
288 than those on other biochars. This could be attributed to a lack of micropores in BC350

289 compared to other biochars (Table 2). In addition, the normalized adsorption for atrazine
290 was slightly higher than that for carbaryl at low solute concentration; this could be related
291 to the smaller molecular size of atrazine compared to carbaryl (0.61 nm versus 0.71 nm,
292 Figure A.3 in Supplementary Information). These findings suggested that pore-filling effect
293 was a main mechanism for pesticide sorption on the biochars and contributed more at low
294 solute concentrations.

295 Strong adsorption of HOCs on micro- and mesoporous carbon is usually driven by
296 specific interactions [10-11, 24-25]. In the present study, the chlorine substituent in
297 atrazine and the ester group in carbaryl have electron-withdrawing natures, making the
298 associated aromatic π -structures electron acceptors, which could interact with the
299 aromatic carbon (electron-donors) on the biochar surface through $\pi-\pi$ electron
300 donor-acceptor (EDA) interactions. Previous studies have shown that $\pi-\pi$ interactions
301 are much stronger than Van der Waals forces [21]. Therefore, the high adsorption affinity
302 between the pesticides and the biochars may have resulted from specific interactions
303 between the pesticide molecules and the biochar surface.

304 Moreover, the polarity of sorbents has been elucidated to exhibit negative effect on
305 the sorption of organic compounds. Recent literature proposed that surface polarity is a
306 more important parameter affecting adsorption than bulk polarity [24]. In the present study,
307 K_{oc} values at two equilibrium concentrations (0.5 and 20 mg/L) on different biochars
308 displayed an insignificant negative correlation ($P>0.05$) with the bulk polarity index [(O +
309 N)/ C] for both pesticides, while no clear correlation was observed for surface polarity
310 (Figure A.2 in Supplementary Information). The surface polarity of BC700 was the highest

311 due to its highest ash content; while it exhibited a substantially high sorption capacity. This
312 is because the two pesticides can be adsorbed to mineral ash through specific
313 interactions. Carbaryl is a neutral insecticide that contains an ester group. It has been
314 reported that clay minerals can adsorb carbamate insecticide through site-specific
315 interactions between the carbamate functional group and exchangeable cations on solid
316 surface [47]. Atrazine is a weak base, with a pK_a of 1.68 [48]. At the pH used in this study,
317 atrazine existed as neutral molecules, therefore, they could form weak hydrogen bonds to
318 the clay surface or carboxyl groups through their heterocyclic nitrogen atoms [49]. In
319 addition, the surface of BC700 was not covered completely by ash, and uncovered
320 aromatic sites were available for sorption and contributed to the high sorption capacity.

321 After deashing, the sorption increased. To estimate the influence of ash and its
322 interaction with organic moieties in adsorption, the adsorption capacity of the deashed
323 biochars was taken as the adsorption capacity of the organic matter fraction in the original
324 biochars. Estimated isotherms fractionized by organic matter content in the original
325 biochars were then drawn (Figure 2). For carbaryl, the estimated isotherm for BC350 is
326 well above its measured isotherm; while the one for BC700 is close to the measured
327 isotherm. Similar trend was observed for atrazine. These results indicate that though the
328 ash could combine with the pesticides, the ash also reduced the sorption onto organic
329 matter by covering sorption sites or making the sorption sites more difficult to access. As a
330 result, the overall contribution of ash to adsorption in the original biochar, which was a
331 complex of ash and organic matter, was very weak or even negative [25]. The difference
332 between the measured and predicted isotherms was more obvious for BC350 compared

333 to BC700. This is because BC350 contained more amorphous carbons, which exhibit
334 mainly partition and weak interaction; while BC700 contained crystalline carbons of
335 specific interactions [50, 51]. Hence, the adsorption affinity of BC350 was weak compared
336 to BC700, and the negative effect of inorganic moiety in BC350 was greater than that in
337 BC700.

338 **3.4 Hydrolysis.**

339 Carbaryl was hydrolyzed markedly in the suspension of the original biochars ($P < 0.05$),
340 and the hydrolysis occurred in two phases (Figure 3). The 7 d hydrolysis extents ranged
341 between 23.5% and 90.6%, being greater at higher biochar dose and in the presence of
342 biochar acquired at higher pyrolysis temperature ($P < 0.05$, Table 4). The hydrolysis of
343 atrazine was much weaker than that of carbaryl ($P < 0.05$). The hydrolysis extents of
344 atrazine in the presence of different doses of the original biochars were less than 25%
345 except in the presence of 500 mg BC700, where a 7 d hydrolysis extent of 63.4% was
346 achieved (Figure 3b, Table 4). Deashing of the chars decreased the hydrolysis extents of
347 the two pesticides. The 7 d hydrolysis extents ranged between 14.3% and 16.6% for
348 carbaryl and 8.5% and 10.2% for atrazine in suspensions containing 500 mg deashed
349 biochars (Figure A.4, Table 4).

350 **3.5 Mechanisms behind the Enhanced Hydrolysis.**

351 The pH is a crucial factor affecting the hydrolysis. Both the residue minerals and
352 aromatic structure in biochars could lead to an elevation in pH [52]. As shown in Table 4,
353 at the low biochar dose (50 mg), the pH of the suspensions was weakly basic (7.1 for
354 BC350 and 7.6 for BC700). The pH increased significantly as the biochar dose increased

355 to 500 mg (7.9 for BC350 and 9.1 for BC700). Hydrolysis of the carbamate ester bond in
356 carbaryl is typically base-catalyzed [53]. Hence, the elevated pH could explain the
357 enhanced hydrolysis of carbaryl in the presence of biochar. Atrazine is a moderately
358 persistent herbicide that can be hydrolyzed in strong acidic or alkaline solutions [54]. The
359 pH values of biochar suspensions were only weakly basic except for BC700 at the dose of
360 500 mg, and these weakly basic suspensions could not efficiently catalyze the hydrolysis
361 of atrazine. The pH values of suspensions containing 500 mg deashed biochars were 6.5
362 and 6.6 for DABC350 and DABC700, respectively. This could partially explain the
363 decreasing hydrolysis of the pesticides in suspensions of the deashed biochars.

364 To confirm the effect of elevated pH on hydrolysis, the hydrolysis of the two pesticides
365 was examined in background solution with adjusted pH (Figure A.5). Two pH values (6.5
366 and 9.1) were selected based on the pH values of the biochar suspensions. Carbaryl was
367 hydrolyzed slowly at pH 6.5, with a 7 d hydrolysis extent of 14.5%; however, at pH of 9.1,
368 the hydrolysis became faster and 86.5% of carbaryl decomposed after 24 h. The
369 hydrolysis extent of atrazine was very weak, with less than 20% decreasing within 7 d
370 even at pH 9.1.

371 The hydrolysis extent of the pesticides in biochar suspensions differed a lot from
372 those in pure background solutions with the same pH values, and both enhancement and
373 inhibition occurred. The reduced hydrolysis in the presence of biochar, such as the case
374 for carbaryl in suspension containing 500 mg of BC700, could be explained by the
375 reduced availability of sorbed pesticides for chemical reaction. This result is consistent
376 with our other study, in which we found that Fenton oxidation of pyrene was significantly

377 reduced after pyrene was sorbed onto soils [55]. However, hydrolysis was markedly
378 greater in the presence of biochar for some cases, such as for atrazine in the suspension
379 containing 500 mg of BC700 ($P < 0.05$). This result indicated that factors other than pH may
380 catalyze the hydrolysis of pesticides.

381 To find the catalytic effects of other factors, the hydrolysis of the two pesticides was
382 examined in the suspension containing 500 mg biochar with pH adjusted to neutral (6.8)
383 so that the effects of pH could be excluded (Figure A.6 and Table 4). Hydrolysis of both
384 pesticides occurred in neutral biochar suspensions, and the hydrolysis was greater in the
385 presence of BC700 compared to BC350. The 7 d hydrolysis extent was 59.3% and 33.5%
386 for carbaryl and atrazine in the neutral suspensions containing 500 mg of BC700,
387 respectively. Previous studies found that clay and metal oxides can catalyze the
388 hydrolysis in several ways [56]. Bound metal atoms present at solid surface may
389 coordinate a hydrolysable moiety by forming complexes with pesticides, which would
390 facilitate the nucleophilic attack of water molecules. In addition, hydroxo groups bound to
391 the surface may act directly as nucleophiles. Furthermore, adsorption on the surface (or in
392 vicinal water surrounding the surface) leads to accumulation of reactants (organic
393 compounds and nucleophiles), thus facilitating the reaction. The original biochars used in
394 this study, especially BC700, had high ash contents. This could explain the catalytic
395 capacity of the biochars for pesticide hydrolysis.

396 Moreover, biochars may release metal ions into solution, which can catalyze
397 hydrolysis via similar mechanisms as a mineral surface [56]. First, metal species may
398 facilitate the attack of a nucleophile, analogous to proton catalysis. Second, dissociation of

399 a leaving group may be promoted by interaction of the leaving group with a metal center.
400 Third, a nucleophile bound to metal centers (metal hydroxo species) may have a higher
401 nucleophilicity as compared to its nucleophilicity in bulk water. To illustrate the possible
402 effect of the dissolved ions, the hydrolysis of two pesticides in the leachate of 500 mg
403 original biochar was investigated (Figure A.7). Hydrolysis of carbaryl was more rapid than
404 that in the background solution at the same pH (Table 4). When the pH was adjusted to
405 neutral, the hydrolysis of carbaryl was still substantial, and a 7 d percentage hydrolysis of
406 40% was achieved. This result indicates that metal ions in the suspensions also contribute
407 to catalyze the hydrolysis of carbaryl. Metal ions were scanned by ICP-MS (Table A.1),
408 and alkali, alkali-earth, and transition metals were detected, with transition metals in
409 relatively lower amounts. After deashing, metal ions in the leachates were reduced by
410 three to five orders of magnitudes (Table A.1). The change in metal ions together with that
411 in pH could explain the decreasing hydrolysis in the presence of deashed biochars.
412 However, the hydrolysis of atrazine in the leachate of original biochars was weak, with all
413 samples less than 12%. This suggests that dissolved ions had less effect on atrazine
414 catalytic hydrolysis.

415 4 Conclusions

416 This research evaluated the impacts of biochars derived from pig manure, which have
417 high ash contents, on the fate of two pesticides (carbaryl and atrazine). In the original
418 biochar, which is a complex of organic matter and inorganic ash, pore-filling and specific
419 interactions contributed much to adsorption of the pesticides besides hydrophobic effects.
420 The ash can combine pesticides by specific interactions, however, its enhancement in

421 sorption was counteracted because the adsorption sites of organic moieties were masked
422 by ash. After deashing, sorption was enhanced significantly, and DABC700 has the
423 highest sorption capacity. Hydrolysis of carbaryl was enhanced in the suspensions of
424 biochar, due to the catalytic effects of elevated pH, released dissolved metal ions and the
425 mineral surface. However, hydrolysis of atrazine was mainly enhanced by high pH and
426 mineral surface. The influence of biochar on the fate of pesticides can vary greatly due to
427 different chemical compositions and steric conformations. Such influences should be
428 systematically studied.

429 **Support Information.** Details of pretreatment and analysis of the samples; One table
430 showing metal ions in the leachate of biochars; and seven figures, (1) Surface area -
431 normalized adsorption isotherms; (2) Correlations between K_{oc} of pesticides and polarity of
432 biochars; (3) Chemical structure and molecular size of the two pesticides; and (4-7)
433 Hydrolysis dynamics under different conditions.

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438

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592

Table 1. Bulk Elemental Composition and Chemical Composition of the Original and Deashed Biochars

sorbents	bulk elemental composition (%)						atomic ratio ^a		chemical shift (ppm) and carbon assignment					
	f_{om}^b	C	H	N	O	ash	H/C	(O+N)/C	0-50	50-109	109-145	145-163	163-190	190-220
									alkyl	O-alkyl	aromatic	phenolic	carboxyl	carbonyl
BC350	54.6	31.5	2.3	3.8	16.9	45.3	0.9	0.51	30	16	37	8	8	1
	7	8	6	0	3	3	0							
BC700	33.1	25.1	1.1	2.0	4.83	66.8	0.5	0.21	13	16	43	2	14	12
	6	6	2	5	4	3	0							
DABC350	98.7	66.6	4.4	7.5	20.0	1.27	0.8	0.32	30	12	33	10	10	5
	3	8	4	8	3	0	0							
DABC700	93.4	74.2	2.7	4.9	11.4	6.54	0.4	0.17	10	13	44	5	14	14
	6	2	9	7	8	5	0							

^a Calculated from atomic ratio of the bulk elements ^b Organic matter content. Noted that the bulk elemental composition and carbon assignment of the original and deashed biochars were determined by elemental analyzer and ¹³C NMR, respectively.

Table 2. Surface Elemental Composition, pH, CEC, Surface Area and Pore Volume Parameters for the Original and Deashed Biochars

sorbents	surface elemental composition (%)							atomic ratio ^a		S_{BET}^b (m ² /g)	V_t^c (cm ³ /g)	D_p^d (Å)	pH ^e	CEC (cmol/kg)
	C	N	O	Si	Al	P	S	O/C	(O+N)/C					
BC350	59.1	5.28	20.	1.4	2.09	3.5	1.08	0.26	0.33	23.8	0.053	88.3	8.3	96.8
	4		25	0		7								
BC700	54.8	4.26	22.	2.7	1.08	4.2	1.94	0.31	0.38	32.6	0.035	42.6	9.5	112.5
	2		98	4		5								
DABC350	76.2	6.22	16.	0.3	N.D. ^f	N.D	0.45	0.16	0.23	67.1	0.096	57.5	6.2	5.6
	3		72	7		.								
DABC700	81.1	2.93	13.	1.2	N.D.	N.D	1.10	0.13	0.16	218.1	0.315	57.8	6.4	8.7
	3		62	3		.								

^a Calculated from atomic ratio of the surface elements ^b Surface area determined by N₂ adsorption using the Brunauer – Emmett – Teller (BET) method. ^c Total pore volume, determined at $P/P_0=0.99$. ^d Average pore diameter. ^e pH of biochars measured in solution (biochar:water=2.5:1). ^f Not detected. Noted that surface element composition of the original and deashed biochars was measured by XPS.

Table 3. Freundlich Isotherm Parameters for the Sorption of the Two Pesticides onto Biochars

pesticides	Sorbents	$\lg K_f$	n	R^2	$\lg K_{oc}^a$	$\lg K_{oc}^b$
carbaryl	BC350	2.06±0.029	0.718±0.033	0.991	2.65	2.19
	BC700	2.85±0.018	0.302±0.015	0.990	3.66	2.54
	DABC350	3.35±0.045	0.615±0.058	0.965	3.64	3.03
	DABC700	3.70±0.071	0.448±0.075	0.874	4.00	3.11
atrazine	BC350	1.40±0.082	0.999±0.080	0.969	1.90	1.90
	BC700	2.77±0.030	0.330±0.023	0.976	3.57	2.50
	DABC350	3.15±0.022	0.615±0.037	0.983	3.44	2.83
	DABC700	3.55±0.032	0.427±0.041	0.957	3.85	2.93

^a K_d normalized by f_{oc} , L/kg, calculated from Freundlich model at $C_e = 0.5$ mg/L; ^b at $C_e = 20$ mg/L.

Table 4. The 7 d Hydrolysis Extents of the Two Pesticides and pH Values of Different Experimental Systems. Values with the same letter are not significantly different from each other ($p < 0.05$).

Experiment design	7 d degradation extent (%)		pH
	Carbaryl	Atrazine	
Suspension of 50 mg BC350	23.5±3.5a	12.6±1.5abc	7.1±0.1a
Suspension of 500 mg BC350	59.1±4.3b	21.2±4.2e	7.9±0.3c
Suspension of 50 mg BC700	67.9±3.3c	14.9±3.1bcd	7.6±0.2c
Suspension of 500 mg BC700	90.6±7.5d	63.4±6.5g	9.1±0.2d
Background solution without pH adjusted	14.1±5.6e	8.0±0.1a	6.5±0.1b
Background solution with pH adjusted	100±0.1f	16.1±3.0cd	9.1±0.3d
Suspension of 500 mg BC350 with pH adjusted	29.1±0.2g	21.8±2.3e	6.8±0.1ab
Suspension of 500 mg BC700 with pH adjusted	59.3±2.0b	33.5±0.1f	6.8±0.2ab
Suspension of 500 mg DABC350	14.3±0.8e	8.5±0.6a	6.5±0.3b
Suspension of 500 mg DABC700	16.6±2.3e	10.2±0.9ab	6.6±0.2b
Leachate of 500 mg BC350 without pH adjusted	65.2±0.6c	16.1±0.0cd	7.9±0.1c
Leachate of 500 mg BC700 without pH adjusted	100±0.1f	19.4±0.2de	9.1±0.1d
Leachate of 500 mg BC350 with pH adjusted	37.6±0.2h	8.5±1.7a	6.8±0.3ab
Leachate of 500 mg BC700 with pH adjusted	40.0±0.4h	11.8±1.4abc	6.8±0.3ab

Caption of Figures

Figure 1. FTIR spectra of biochars

Figure 2. Adsorption isotherms of two pesticides by original and deashed biochars. The lines represent the calculated Freundlich model fitting to adsorption data for the deashed biochar fractionalized by to the organic matter content in the original biochar (solid line for BC350; dotted line for BC700).

Figure 3. Hydrolysis of the two pesticides in suspensions of original biochars

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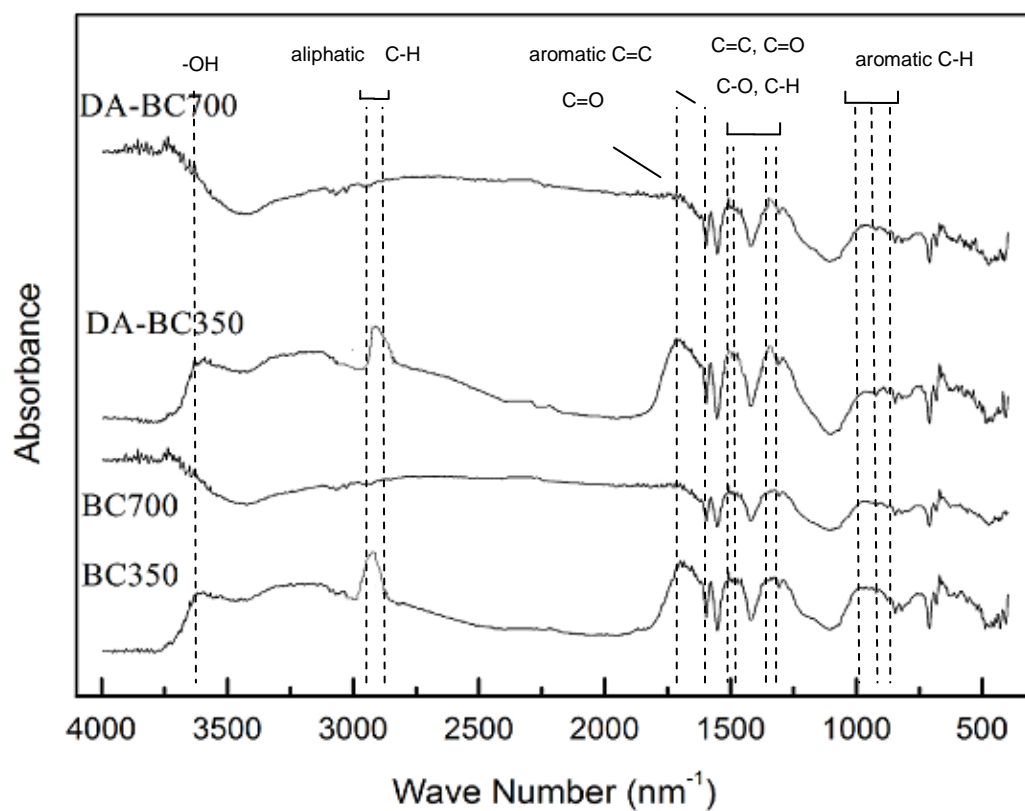


Figure 1. FTIR spectra of biochars

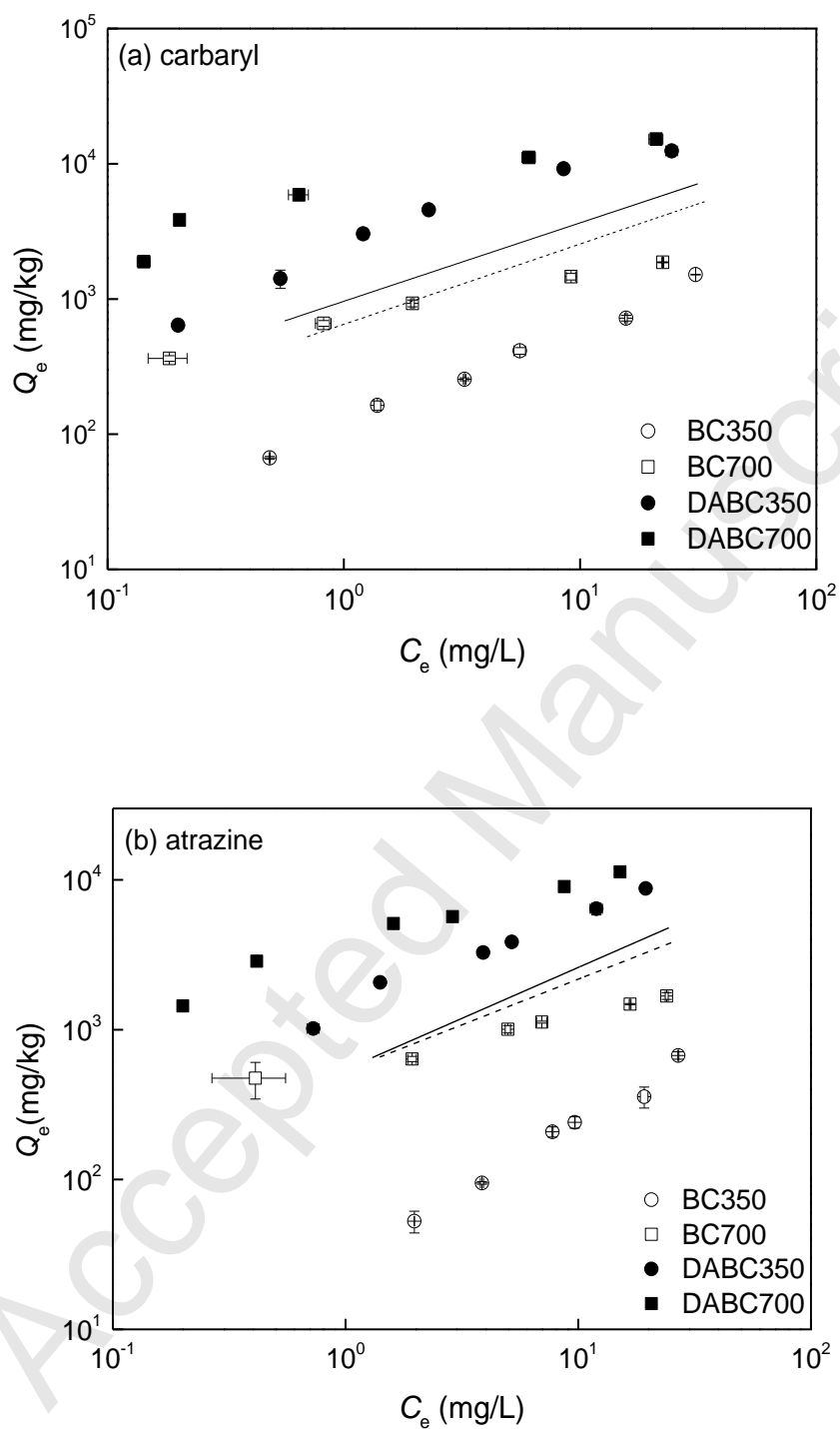


Figure 2. Adsorption isotherms of two pesticides by original and deashed biochars. The lines represent the calculated Freundlich model fitting to adsorption data for the deashed biochar fractionalized by the organic matter content in the original biochar (solid line for BC350; dotted line for BC700).

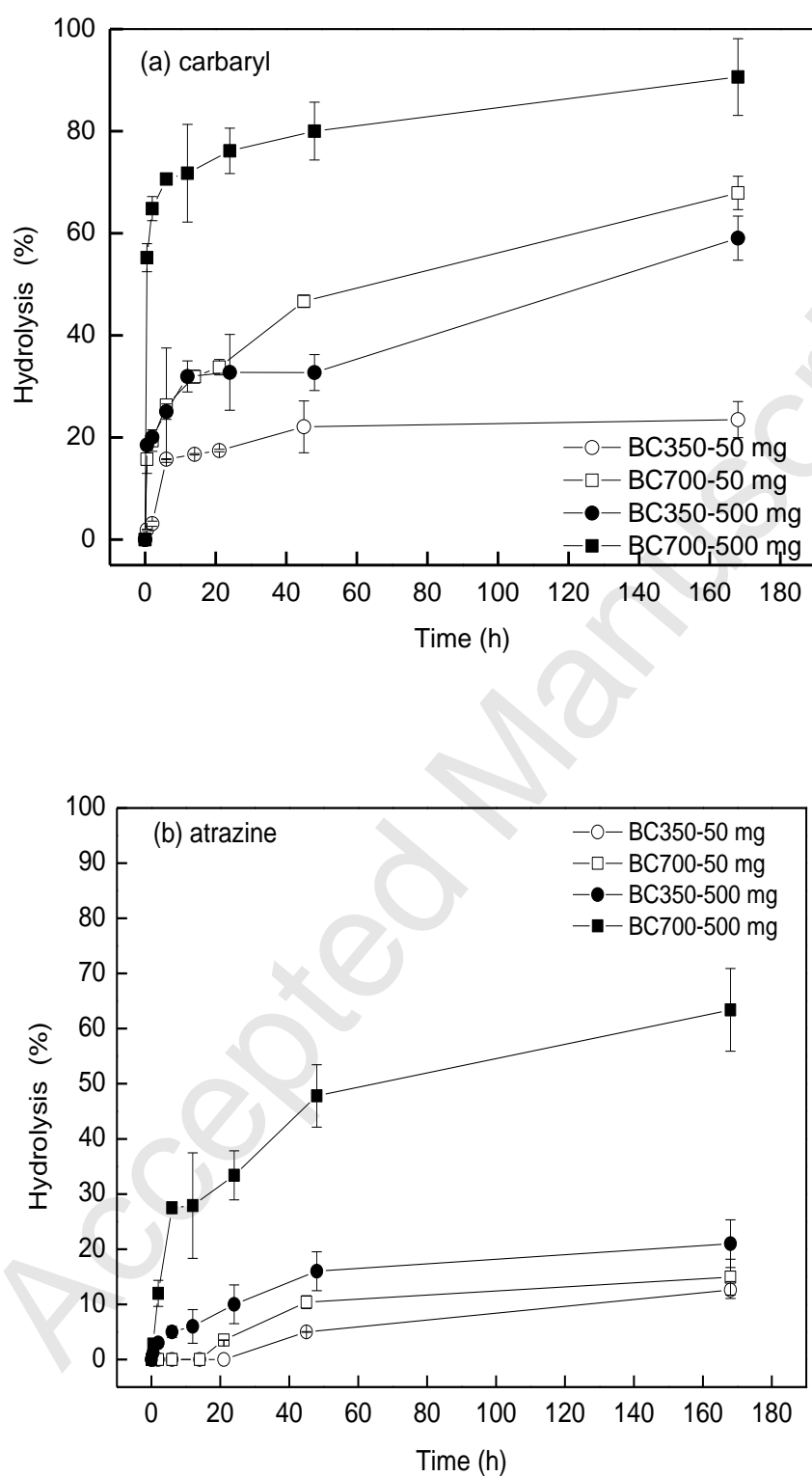


Figure 3. Hydrolysis of the two pesticides in suspensions of original biochars.