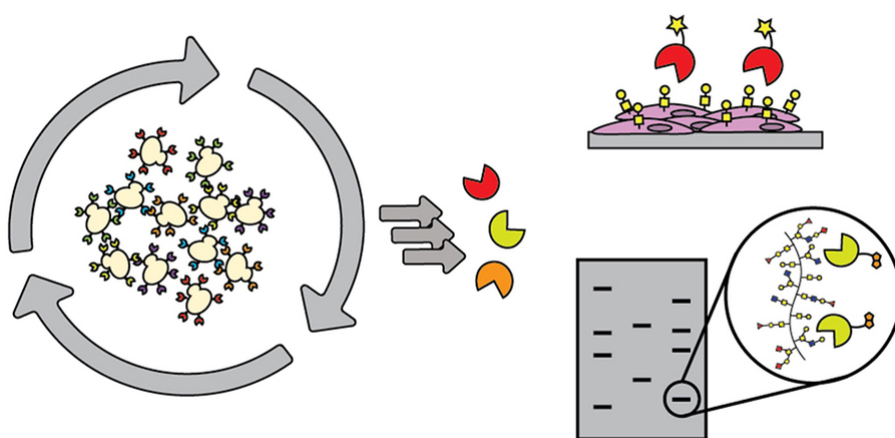


New tool can assist with identifying carbohydrate-binding proteins

Groundbreaking research can help alleviate the challenges affiliated with studying carbohydrates.

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A new tool from the Imperiali Lab uses directed evolution to generate glycan-binding proteins (GBPs) from small, hyper-thermostable DNA-binding protein.

Image courtesy of the researchers.

One of the major obstacles that those conducting research on carbohydrates are constantly working to overcome is the limited array of tools available to decipher the role of sugars. As a workaround, most researchers utilize lectins (sugar-binding proteins) isolated from plants or fungi, but they are large, with weak binding, and they are limited in their

specificity and in the scope of sugars that they detect. In a new [study published in ACS Chemical Biology](#), researchers in Professor [Barbara Imperiali's](#) group have developed a platform to address this shortcoming.

“The challenge with polymers of carbohydrates is that their biosynthesis is not template-driven,” says Imperiali, the senior author of the study and a professor in the departments of Chemistry and Biology. “Biology, medicine, and biotechnology have been fueled by technological advancements for proteins and nucleic acids. The carbohydrate field lags terribly behind and is desperately seeking tools.”

Identifying carbohydrate-binding proteins

Biosynthesizing carbohydrates requires every link between individual sugar molecules to be made by a particular enzyme, and there’s no ready way to decipher the structures and sequences of complex carbohydrates. Antibodies to carbohydrates can be generated, but doing so is challenging, expensive, and results in a molecule that is far larger than what is really needed for the research. An ideal resource for this field plagued with limited mechanisms would be discovery of binding proteins, of limited size, that recognize small chunks of carbohydrates to piece together a structure by using those binders, or methods to detect and identify particular carbohydrates within complicated structures.

The authors of this study used directed evolution and clever screen design to identify carbohydrate-binding proteins from proteins that have absolutely no ability to bind carbohydrates at all. Their findings lay the groundwork for identifying carbohydrate-binding proteins with diverse and programmable specificity.

Streamlining for collaboration

This advance will allow researchers to go after a user-defined sugar target without being limited by what a lectin does, or challenged by the abilities of generating antibodies. These results could serve to inspire future collaborations with engineering communities to maximize the efficiency of glycobiology’s yeast surface display pipeline. As it is, this pipeline works well for proteins, but sugars are far more difficult targets and require the pipeline to be modified.


In terms of future applications, the potential for this innovation ranges from diagnostic to, in the longer term, therapeutic, and paves the way for collaborations with researchers at MIT and beyond. For example, chemistry Professor [Laura Kiessling's](#) research group works

with *Mycobacterium tuberculosis* (Mtb), which has an unusual cell wall composition with unique, distinct, and exclusive sugars. Using this method, a binder could potentially be evolved to that particular feature on Mtb. Chemical engineering Professor [Hadley Sikes](#) develops paper-based diagnostic tools where the binding partner for a particular epitope or marker is laid down, and with the use of this discovery, in the longer term, a lateral flow assay device could be developed.

Laying the groundwork for future solutions

In cancer, certain sugars are overrepresented on cell surfaces, so theoretically, researchers can utilize this finding, which is also amenable to labeling, to develop a tool out of the evolved glycan binder for detection.

This discovery also stands to contribute significantly to improving cell imaging. Researchers can modify binders with a fluorophore using a simple ligation strategy, and can then choose the best fluorophore for tissue or cell imaging. The Kiessling group, for example, could apply small protein binders labeled with fluorophore to detect bacterial sugars to initiate fluorescence-activated cell sorting to probe a complex mixture of microbes. This could in turn be used to determine how a patient's microbiome has been disturbed. It also has the potential to screen the microbiome of a patient's mouth or their upper or lower gastrointestinal tract to read out the imbalance within the community using these types of reagents. In the more distant future, the binders could potentially have therapeutic purposes like clearing the gastrointestinal tract or mouth of a particular bacterium based on the sugars that the bacterium displays.

 [Paper: "Engineered Glycan-Binding Proteins for Recognition of the Thomsen-Friedenreich Antigen and Structurally Related Disaccharides"](#)