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### **Title**

Runx1 Regulates c-Myc Expression And The Expansion Of Hematopoietic Precursors In A C-terminally Dependent Manner

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Molecular and Cellular Biology

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## Abstract

Runx1 regulates the expression of several important target genes and plays critical roles in the process of hematopoiesis. Runx1 by itself is a poor regulator of transcription and instead nucleates transcription complexes through its C-terminus to transactivate or repress the expression of target genes. We generated a C-terminally deleted Runx1 construct (Runx1.d190), which lacks important co-factor sites, to further investigate the function of Runx1 in development. A potential role for Runx1 in regulating the expression of another potent transcriptional regulator, c-Myc, has been suggested by published studies, which show that Runx1 and c-Myc collaborate in oncogenesis. In these studies, we show that endogenous Runx1 binds to three Runx consensus sites upstream of the *c-Myc* transcriptional start site in Jurkat T cells and murine primary splenocytes. Retroviral transduction of Jurkat T cells with Runx1.d190 results in the increased transcription of *c-Myc* as determined by microarray analysis. In order to monitor *c-Myc* expression in response to early-acting and transient Runx1.d190, we generated a cell membrane-permeable TAT-Runx1.d190 fusion protein. Treatment of murine primary splenocytes with TAT-Runx1.d190 protein results in a transient increase in the transcription of *c-Myc* and a corresponding increase in *c-Myc* protein levels. This effect is dependent on the ability of Runx1.d190 to bind to DNA. These data demonstrate that Runx1 directly regulates *c-Myc* expression in a C-terminally and DNA-binding dependent manner. In these studies, we also investigate the effects of the truncation of Runx1 C-terminus on hematopoietic stem cells (HSCs). We found that treatment of bone marrow cells enriched for HSCs with TAT-Runx1.d190 results in a 12.5 fold increase in hematopoietic precursors compared to untreated precursors *in vitro* as determined by Colony Forming Cell assays. We also show that hematopoietic precursors treated with TAT-Runx1.d190 are able to differentiate normally both *in vitro* and *in vivo* and thus represent functional hematopoietic precursors. Our findings show that we are able to transiently expand hematopoietic precursors *in vivo* by treating the cells with a Runx1 construct that lacks the C-terminus. Collectively, this work demonstrates that Runx1 regulates the expression of *c-Myc* and the expansion of hematopoietic precursors in a C-terminally dependent manner.

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